DATA NOTE

The genome sequence of the Shaggy Mining Bee, *Lasioglossum villosulum* (Kirby, 1802) (Hymenoptera: Halictidae)

[version 1; peer review: 3 approved]

Steven Falk[1], University of Oxford and Wytham Woods Genome Acquisition Lab, Darwin Tree of Life Barcoding Collective,
Wellcome Sanger Institute Tree of Life Management, Samples and Laboratory team,
Wellcome Sanger Institute Scientific Operations: Sequencing Operations,
Wellcome Sanger Institute Tree of Life Core Informatics team,
Tree of Life Core Informatics collective, Darwin Tree of Life Consortium

[1]Independent researcher, Kenilworth, Warwickshire, England, UK

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Abstract

We present a genome assembly from an individual female *Lasioglossum villosulum* (Shaggy Mining Bee; Arthropoda; Insecta; Hymenoptera; Halictidae). The genome sequence has a total length of 458.49 megabases. Most of the assembly (87.76%) is scaffolded into 11 chromosomal pseudomolecules. The mitochondrial genome has also been assembled, with a length of 29.57 kilobases. This assembly was generated as part of the Darwin Tree of Life project, which produces reference genomes for eukaryotic species found in Britain and Ireland.

Keywords

Lasioglossum villosulum; Shaggy Mining Bee; genome sequence; chromosomal; Hymenoptera

This article is included in the Tree of Life gateway.

Open Peer Review

Approval Status ✓ ✓ ✓

	1	2	3
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1. **Christopher Wyatt**, University College London, London, UK
2. **Pilar De la Rúa**, University of Murcia, Murcia, Spain
3. **Eduardo Zattara**, Instituto de Investigaciones en Biodiversidad y Medioambiente, Río Negro, Argentina

Any reports and responses or comments on the article can be found at the end of the article.

Corresponding author: Darwin Tree of Life Consortium (mark.blaxter@sanger.ac.uk)

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Species taxonomy

Eukaryota; Opisthokonta; Metazoa; Eumetazoa; Bilateria; Protostomia; Ecdysozoa; Panarthropoda; Arthropoda; Mandibulata; Pancrustacea; Hexapoda; Insecta; Dicondylia; Pterygota; Neoptera; Endopterygota; Hymenoptera; Apocrita; Aculeata; Apoidea; Anthophila; Halictidae; Halictinae; Halictini; *Lasioglossum*; *Hemihalictus*; *Lasioglossum villosulum* (Kirby, 1802) (NCBI: txid88523)

Background

Lasioglossum villosulum (Kirby, 1802) is a small member of the Halictidae, informally called the "sweat bees". Individuals are typically smaller than honey bees, and may have pale hair bands on the abdomen, though these are not always present. In Britain, the species is widespread and locally common, occurring in a range of habitats including coastal soft rock cliffs (Falk & Lewington, 2015). In Wales and north-west England, it is primarily coastal. It is not currently considered scarce or threatened in Britain (Bees & Wasps and Ants Recording Society, 2006).

The species is bivoltine. Females are active from March to October, while males appear from late June until mid-October (Falk & Lewington, 2015). A peak in activity occurs in July and August, with occasional records into November (Bees & Wasps and Ants Recording Society, 2006).

Nesting biology is solitary, although aggregations are known. In some cases, two females share a nest entrance, but these are interpreted as communal rather than eusocial. The nest burrow is vertical with lateral branches ending in horizontal cells, which are provisioned and sealed following oviposition, with no contact between adult and larva (Bees & Wasps and Ants Recording Society, 2006).

Flowers visited include *Smyrnium olusatrum*, *Ranunculus* spp., *Cirsium arvense*, *Crataegus monogyna*, *Heracleum sphondylium*, and *Prunus* spp. (Bees & Wasps and Ants Recording Society, 2006), with a particular preference for yellow-flowered Asteraceae, especially *Picris hieracioides* and other Cichorioideae (Sjödin, 2010). It is subject to cleptoparasitism by *Sphecodes puncticeps* and *S. geoffrellus*, though no species is known to be host-specific (Bees & Wasps and Ants Recording Society, 2006; Falk & Lewington, 2015).

Although often overlooked in crop pollination, *L. villosulum* and other *Lasioglossum* species can be effective pollinators due to high visitation rates, despite their relatively small size and reduced hairiness (Coudrain *et al.*, 2022).

Social behaviour in the genus *Lasioglossum* is diverse. Although *L. villosulum* is regarded as solitary, the genus as a whole shows extensive variability in social traits, ranging from solitary to eusocial forms. Phylogenetic analyses suggest that eusociality in this group evolved early and has been lost repeatedly, including in *L. villosulum*, where social reversion is not necessarily accompanied by reversion to a univoltine life cycle (Gibbs *et al.*, 2012; Groom *et al.*, 2021).

We present a chromosome-level genome sequence for *Lasioglossum villosulum*, generated as part of the Darwin Tree of Life Project. Although numerous genomes exist for the family Halictidae, this assembly provides the first genome sequence for *Lasioglossum villosulum* (data obtained via NCBI datasets, O'Leary *et al.*, 2024). The assembly was produced using the Tree of Life pipeline from a specimen collected in Wytham Woods, Oxfordshire, United Kingdom (Figure 1).

Methods

Sample acquisition and DNA barcoding

The specimen used for genome sequencing was an adult *Lasioglossum villosulum* (specimen ID Ox002188, ToLID iyLasVill1; Figure 1), collected from Wytham Woods, Oxfordshire, United Kingdom (latitude 51.769, longitude –1.328) on 2022-05-19. The specimen was collected and identified by Steven Falk. For the Darwin Tree of Life sampling and metadata approach, refer to Lawniczak *et al.* (2022).

The initial identification was verified by an additional DNA barcoding process according to the framework developed by Twyford *et al.* (2024). A small sample was dissected from the specimen and stored in ethanol, while the remaining parts were shipped on dry ice to the Wellcome Sanger Institute (WSI) (see the protocol). The tissue was lysed, the COI marker region was amplified by PCR, and amplicons were sequenced and compared to the BOLD database, confirming the species identification (Crowley *et al.*, 2023). Following whole genome sequence generation, the relevant DNA barcode region was also used alongside the initial barcoding data for sample tracking at the WSI (Twyford *et al.*, 2024). The standard operating procedures for Darwin Tree of Life barcoding are available on protocols.io.

Nucleic acid extraction

Protocols for high molecular weight (HMW) DNA extraction developed at the Wellcome Sanger Institute (WSI) Tree

Figure 1. Photograph of the *Lasioglossum villosulum* (iyLasVill1) specimen used for genome sequencing.

of Life Core Laboratory are available on protocols.io (Howard *et al.*, 2025). The iyLasVill1 sample was weighed and triaged to determine the appropriate extraction protocol. Tissue from the whole organism was homogenised by powermashing using a PowerMasher II tissue disruptor.

HMW DNA was extracted in the WSI Scientific Operations core using the Automated MagAttract v2 protocol. DNA was sheared into an average fragment size of 12–20 kb following the Megaruptor®3 for LI PacBio protocol. Sheared DNA was purified by automated SPRI (solid-phase reversible immobilisation). The concentration of the sheared and purified DNA was assessed using a Nanodrop spectrophotometer and Qubit Fluorometer using the Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system. For this sample, the final post-shearing DNA had a Qubit concentration of 28 ng/μL and a yield of 1 120.00 ng, with a fragment size of 10.5 kb. The 260/280 spectrophotometric ratio was 1.94, and the 260/230 ratio was 1.72.

PacBio HiFi library preparation and sequencing

Library preparation and sequencing were performed at the WSI Scientific Operations core. Libraries were prepared using the SMRTbell Prep Kit 3.0 (Pacific Biosciences, California, USA), following the manufacturer's instructions. The kit includes reagents for end repair/A-tailing, adapter ligation, post-ligation SMRTbell bead clean-up, and nuclease treatment. Size selection and clean-up were performed using diluted AMPure PB beads (Pacific Biosciences). DNA concentration was quantified using a Qubit Fluorometer v4.0 (ThermoFisher Scientific) and the Qubit 1X dsDNA HS assay kit. Final library fragment size was assessed with the Agilent Femto Pulse Automated Pulsed Field CE Instrument (Agilent Technologies) using the gDNA 55 kb BAC analysis kit.

The sample was sequenced using the Sequel IIe system (Pacific Biosciences, California, USA). The concentration of the library loaded onto the Sequel IIe was in the range 40–135 pM. The SMRT link software, a PacBio web-based end-to-end workflow manager, was used to set-up and monitor the run, and to perform primary and secondary analysis of the data upon completion.

Hi-C

Sample preparation and crosslinking

The Hi-C sample was prepared from 20–50 mg of frozen tissue of the iyLasVill1 sample using the Arima-HiC v2 kit (Arima Genomics). Following the manufacturer's instructions, tissue was fixed and DNA crosslinked using TC buffer to a final formaldehyde concentration of 2%. The tissue was homogenised using the Diagnocine Power Masher-II. Crosslinked DNA was digested with a restriction enzyme master mix, biotinylated, and ligated. Clean-up was performed with SPRISelect beads before library preparation. DNA concentration was measured with the Qubit Fluorometer (Thermo Fisher Scientific) and Qubit HS Assay Kit. The biotinylation percentage was estimated using the Arima-HiC v2 QC beads.

Hi-C library preparation and sequencing

Biotinylated DNA constructs were fragmented using a Covaris E220 sonicator and size selected to 400–600 bp using SPRISelect beads. DNA was enriched with Arima-HiC v2 kit Enrichment beads. End repair, A-tailing, and adapter ligation were carried out with the NEBNext Ultra II DNA Library Prep Kit (New England Biolabs), following a modified protocol where library preparation occurs while DNA remains bound to the Enrichment beads. Library amplification was performed using KAPA HiFi HotStart mix and a custom Unique Dual Index (UDI) barcode set (Integrated DNA Technologies). Depending on sample concentration and biotinylation percentage determined at the crosslinking stage, libraries were amplified with 10–16 PCR cycles. Post-PCR clean-up was performed with SPRISelect beads. Libraries were quantified using the AccuClear Ultra High Sensitivity dsDNA Standards Assay Kit (Biotium) and a FLUOstar Omega plate reader (BMG Labtech).

Prior to sequencing, libraries were normalised to 10 ng/μL. Normalised libraries were quantified again and equimolar and/or weighted 2.8 nM pools. Pool concentrations were checked using the Agilent 4200 TapeStation (Agilent) with High Sensitivity D500 reagents before sequencing. Sequencing was performed using paired-end 150 bp reads on the Illumina NovaSeq 6000.

Genome assembly

Prior to assembly of the PacBio HiFi reads, a database of *k*-mer counts ($k = 31$) was generated from the filtered reads using FastK. GenomeScope2 (Ranallo-Benavidez *et al.*, 2020) was used to analyse the *k*-mer frequency distributions, providing estimates of genome size, heterozygosity, and repeat content.

The HiFi reads were assembled using Hifiasm (Cheng *et al.*, 2021) with the --primary option. Haplotypic duplications were identified and removed using purge_dups (Guan *et al.*, 2020). The Hi-C reads (Rao *et al.*, 2014) were mapped to the primary contigs using bwa-mem2 (Vasimuddin *et al.*, 2019), and the contigs were scaffolded in YaHS (Zhou *et al.*, 2023) with the --break option for handling potential misassemblies. The scaffolded assemblies were evaluated using Gfastats (Formenti *et al.*, 2022), BUSCO (Manni *et al.*, 2021) and MERQURY.FK (Rhie *et al.*, 2020).

The mitochondrial genome was assembled using MitoHiFi (Uliano-Silva *et al.*, 2023), which runs MitoFinder (Allio *et al.*, 2020) and uses these annotations to select the final mitochondrial contig and to ensure the general quality of the sequence.

Assembly curation

The assembly was decontaminated using the Assembly Screen for Cobionts and Contaminants (ASCC) pipeline. TreeVal was used to generate the flat files and maps for use in curation. Manual curation was conducted primarily in PretextView and HiGlass (Kerpedjiev *et al.*, 2018). Scaffolds were visually inspected and corrected as described by Howe *et al.* (2021). Manual corrections included 6 breaks, 31 joins, and removal of one haplotypic duplication. The curation process is documented at https://gitlab.com/wtsi-grit/rapid-curation. PretextSnapshot was used to generate a Hi-C contact map of the final assembly.

Assembly quality assessment

The Merqury.FK tool (Rhie *et al.*, 2020) was run in a Singularity container (Kurtzer *et al.*, 2017) to evaluate *k*-mer completeness and assembly quality for the primary and alternate haplotypes using the *k*-mer databases (k = 31) computed prior to genome assembly. The analysis outputs included assembly QV scores and completeness statistics.

The genome was analysed using the BlobToolKit pipeline, a Nextflow implementation of the earlier Snakemake version (Challis *et al.*, 2020). The pipeline aligns PacBio reads using minimap2 (Li, 2018) and SAMtools (Danecek *et al.*, 2021) to generate coverage tracks. It runs BUSCO (Manni *et al.*, 2021) using lineages identified from the NCBI Taxonomy (Schoch *et al.*, 2020). For the three domain-level lineages, BUSCO genes are aligned to the UniProt Reference Proteomes database (Bateman *et al.*, 2023) using DIAMOND blastp (Buchfink *et al.*, 2021). The genome is divided into chunks based on the density of BUSCO genes from the closest taxonomic lineage, and each chunk is aligned to the UniProt Reference Proteomes database with DIAMOND blastx. Sequences without hits are chunked using seqtk and aligned to the NT database with blastn (Altschul *et al.*, 1990). The BlobToolKit suite consolidates all outputs into a blobdir for visualisation. The BlobToolKit pipeline was developed using nf-core tooling (Ewels *et al.*, 2020) and MultiQC (Ewels *et al.*, 2016), with containerisation through Docker (Merkel, 2014) and Singularity (Kurtzer *et al.*, 2017).

Genome sequence report

Sequence data

PacBio sequencing of the *Lasioglossum villosulum* specimen generated 25.26 Gb (gigabases) from 2.44 million reads, which were used to assemble the genome. GenomeScope2.0 analysis estimated the haploid genome size at 411.83 Mb, with a heterozygosity of 0.79% and repeat content of 32.25% (Figure 2). These estimates guided expectations for the assembly. Based on the estimated genome size, the sequencing data provided approximately 58× coverage. Hi-C sequencing produced 112.38 Gb from 744.22 million reads, which were used to scaffold the assembly. Table 1 summarises the specimen and sequencing details.

Assembly statistics

The primary haplotype was assembled, and contigs corresponding to an alternate haplotype were also deposited in INSDC

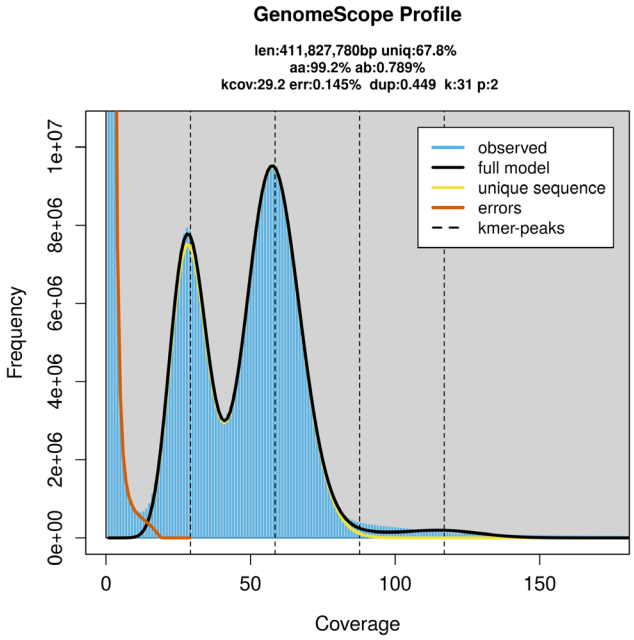

Figure 2. Frequency distribution of *k*-mers generated using GenomeScope2. The plot shows observed and modelled *k*-mer spectra, providing estimates of genome size, heterozygosity, and repeat content based on unassembled sequencing reads.

databases. The final assembly has a total length of 458.49 Mb in 191 scaffolds, with 177 gaps, and a scaffold N50 of 40.46 Mb (Table 2).

Most of the assembly sequence (87.76%) was assigned to 11 chromosomal-level scaffolds. These chromosome-level scaffolds, confirmed by Hi-C data, are named according to size (Figure 3; Table 3).

The mitochondrial genome was also assembled. This sequence is included as a contig in the multifasta file of the genome submission and as a standalone record.

The combined primary and alternate assemblies achieve an estimated QV of 61.2. The *k*-mer completeness is 81.28% for the primary assembly, 79.91% for the alternate haplotype, and 96.93% for the combined assemblies (Figure 4).

BUSCO v.5.5.0 analysis using the hymenoptera_odb10 reference set (n = 5 991) identified 95.8% of the expected gene set (single = 95.4%, duplicated = 0.3%). The snail plot in Figure 5 summarises the scaffold length distribution and other assembly statistics for the primary assembly. The blob plot in Figure 6 shows the distribution of scaffolds by GC proportion and coverage.

Table 4 lists the assembly metric benchmarks adapted from Rhie *et al.* (2021) the Earth BioGenome Project Report on Assembly Standards September 2024. The EBP metric, calculated

Table 1. **Specimen and sequencing data for BioProject PRJEB64955.**

Platform	PacBio HiFi	Hi-C
ToLID	iyLasVill1	iyLasVill1
Specimen ID	Ox002188	Ox002188
BioSample (source individual)	SAMEA110451629	SAMEA110451629
BioSample (tissue)	SAMEA110451839	SAMEA110451839
Tissue	whole organism	whole organism
Instrument	Sequel IIe	Illumina NovaSeq 6000
Run accessions	ERR11843428	ERR11837519
Read count total	2.44 million	744.22 million
Base count total	25.26 Gb	112.38 Gb

Table 2. **Genome assembly statistics.**

Assembly name	iyLasVill1.1
Assembly accession	GCA_963971365.1
Alternate haplotype accession	GCA_963971395.1
Assembly level	chromosome
Span (Mb)	458.49
Number of chromosomes	11
Number of contigs	368
Contig N50	3.09 Mb
Number of scaffolds	191
Scaffold N50	40.46 Mb
Organelles	Mitochondrion: 29.57 kb

for the primary assembly, is **6.7.Q60**, meeting the recommended reference standard.

Wellcome Sanger Institute – Legal and Governance

The materials that have contributed to this genome note have been supplied by a Darwin Tree of Life Partner. The submission of materials by a Darwin Tree of Life Partner is subject to the **'Darwin Tree of Life Project Sampling Code of Practice'**, which can be found in full on the Darwin Tree of Life website. By agreeing with and signing up to the Sampling Code of Practice, the Darwin Tree of Life Partner agrees they will meet the legal and ethical requirements and standards set out within this document in respect of all samples acquired for, and supplied to, the Darwin Tree of Life Project. Further, the Wellcome Sanger Institute employs a process whereby due diligence is carried out proportionate to the nature of the materials themselves, and the circumstances under which they have been/are to be collected and provided for use. The purpose of this is to address and mitigate any potential legal and/or ethical implications of receipt and use of the materials as part of the research project, and to ensure that in doing so we align with best practice wherever possible. The overarching areas of consideration are:

- Ethical review of provenance and sourcing of the material
- Legality of collection, transfer and use (national and international)

Each transfer of samples is further undertaken according to a Research Collaboration Agreement or Material Transfer Agreement entered into by the Darwin Tree of Life Partner, Genome Research Limited (operating as the Wellcome Sanger Institute), and in some circumstances, other Darwin Tree of Life collaborators.

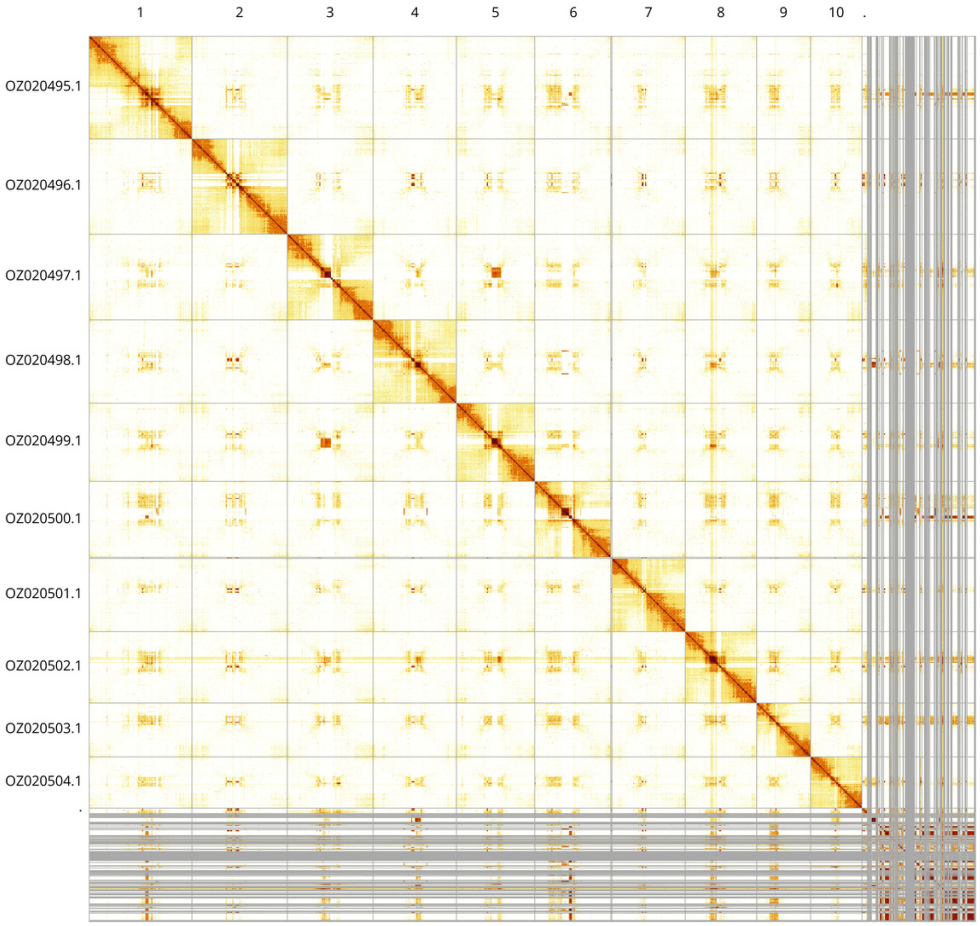

Figure 3. Hi-C contact map of the *Lasioglossum villosulum* genome assembly. Assembled chromosomes are shown in order of size and labelled along the axes. The plot was generated using PretextSnapshot.

Table 3. Chromosomal pseudomolecules in the primary genome assembly of *Lasioglossum villosulum* iyLasVill1.

INSDC accession	Molecule	Length (Mb)	GC%
OZ020495.1	1	53.28	40
OZ020496.1	2	49.27	40
OZ020497.1	3	44.37	41.50
OZ020498.1	4	43.09	41
OZ020499.1	5	40.46	42.50
OZ020500.1	6	39.93	40.50
OZ020501.1	7	37.82	40.50
OZ020502.1	8	36.91	41
OZ020503.1	9	27.86	39
OZ020504.1	10	26.61	41
OZ020505.1	11	2.79	39.50

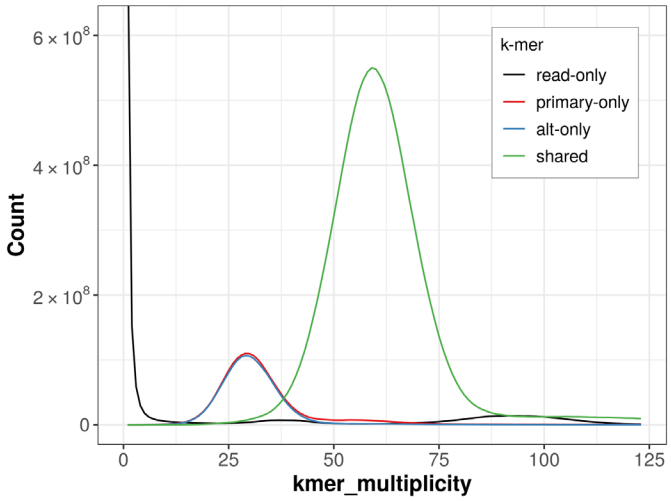

Figure 4. Evaluation of *k*-mer completeness using MerquryFK. This plot illustrates the recovery of *k*-mers from the original read data in the final assemblies. The horizontal axis represents *k*-mer multiplicity, and the vertical axis shows the number of *k*-mers. The black curve represents *k*-mers that appear in the reads but are not assembled. The green curve corresponds to *k*-mers shared by both haplotypes, and the red and blue curves show *k*-mers found only in one of the haplotypes.

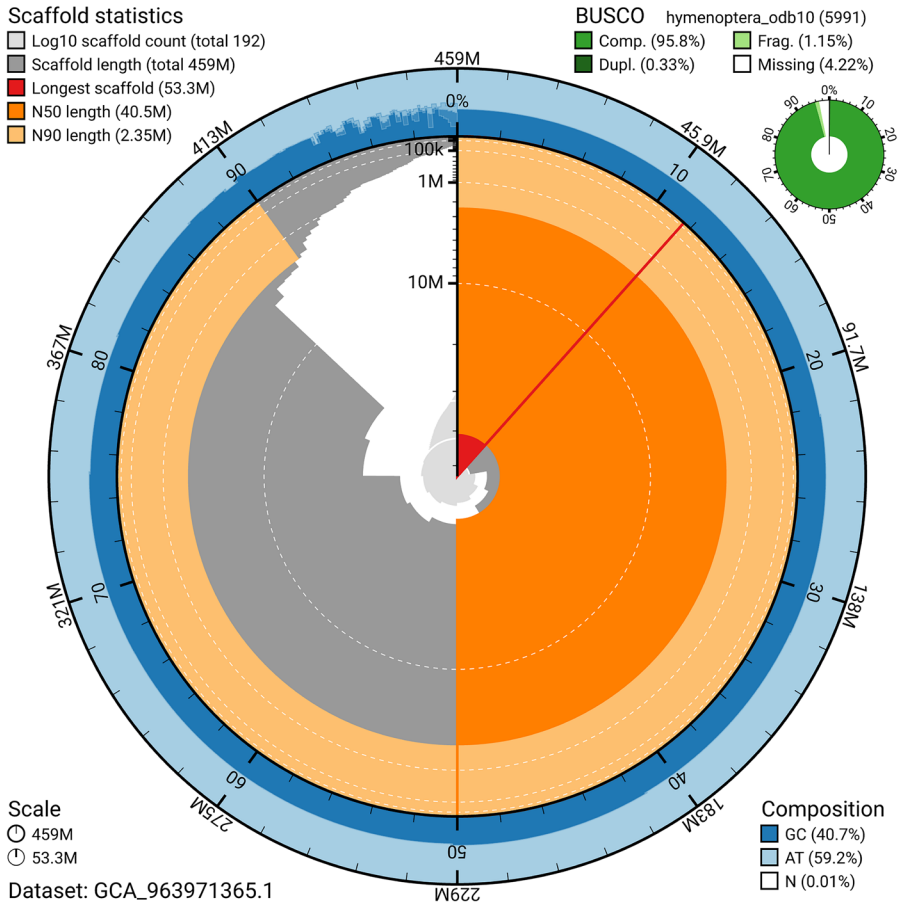

Figure 5. Assembly metrics for iyLasVill1.1. The BlobToolKit snail plot provides an overview of assembly metrics and BUSCO gene completeness. The circumference represents the length of the whole genome sequence, and the main plot is divided into 1 000 bins around the circumference. The outermost blue tracks display the distribution of GC, AT, and N percentages across the bins. Scaffolds are arranged clockwise from longest to shortest and are depicted in dark grey. The longest scaffold is indicated by the red arc, and the deeper orange and pale orange arcs represent the N50 and N90 lengths. A light grey spiral at the centre shows the cumulative scaffold count on a logarithmic scale. A summary of complete, fragmented, duplicated, and missing BUSCO genes in the hymenoptera_odb10 set is presented at the top right. An interactive version of this figure can be accessed on the BlobToolKit viewer.

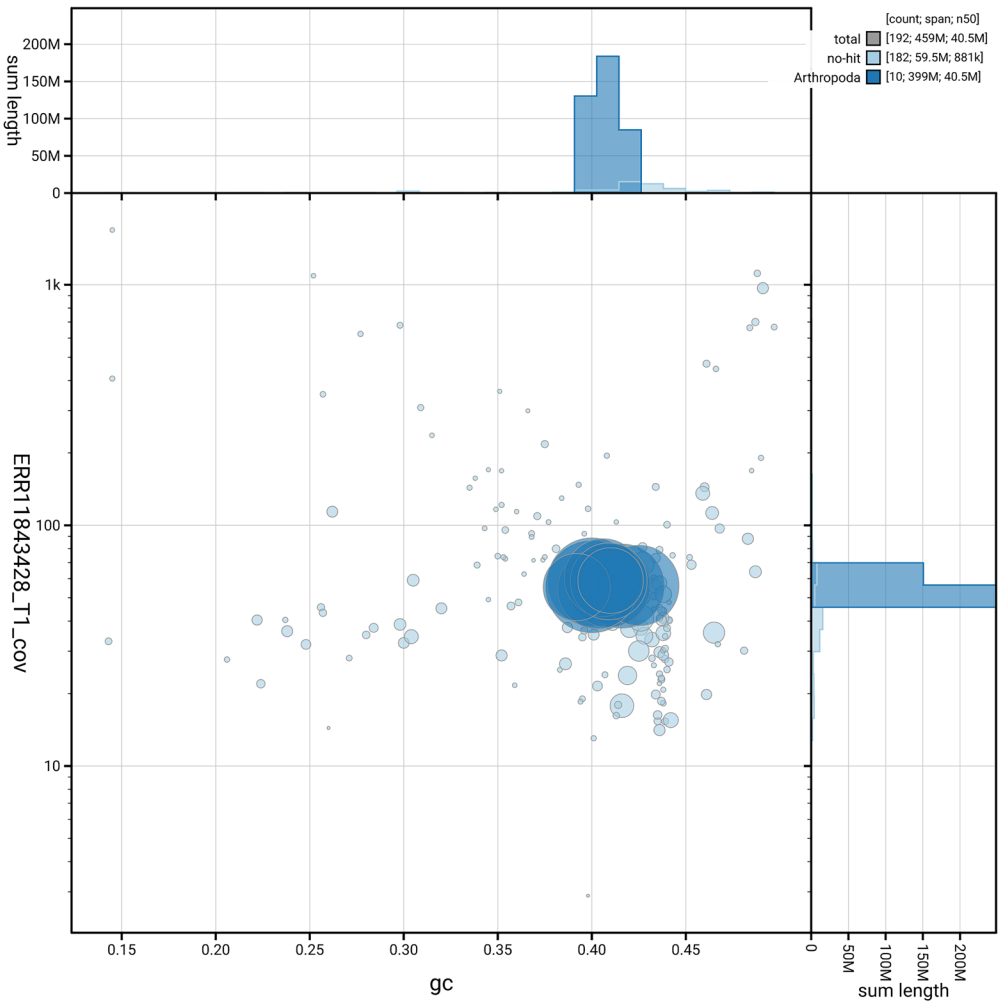

Figure 6. BlobToolKit GC-coverage plot for iyLasVill1.1. Blob plot showing sequence coverage (vertical axis) and GC content (horizontal axis). The circles represent scaffolds, with the size proportional to scaffold length and the colour representing phylum membership. The histograms along the axes display the total length of sequences distributed across different levels of coverage and GC content. An interactive version of this figure is available on the BlobToolKit viewer.

Table 4. Earth Biogenome Project summary metrics for the *Lasioglossum villosulum* assembly.

Measure	Value	Benchmark
EBP summary (primary)	6.7.Q60	6.C.Q40
Contig N50 length	3.09 Mb	≥ 1 Mb
Scaffold N50 length	40.46 Mb	= chromosome N50
Consensus quality (QV)	Primary: 60.6; alternate: 62.0; combined: 61.2	≥ 40
k-mer completeness	Primary: 81.28%; alternate: 79.91%; combined: 96.93%	≥ 95%
BUSCO	C:95.8% [S:95.4%; D:0.3%]; F:1.2%; M:3.1%; n:5 991	S > 90%; D < 5%
Percentage of assembly assigned to chromosomes	87.76%	≥ 90%

Data availability

European Nucleotide Archive: Lasioglossum villosulum (shaggy furrow bee). Accession number PRJEB64955. The genome sequence is released openly for reuse. The *Lasioglossum villosulum* genome sequencing initiative is part of the Darwin Tree of Life Project (PRJEB40665) and the Sanger Institute Tree of Life Programme (PRJEB43745). All raw sequence data and the assembly have been deposited in INSDC databases. The genome will be annotated using available RNA-Seq data and presented through the Ensembl pipeline at the European Bioinformatics Institute. Raw data and assembly accession identifiers are reported in Table 1 and Table 2.

Production code used in genome assembly at the WSI Tree of Life is available at https://github.com/sanger-tol. Table 5 lists software versions used in this study.

Table 5. Software versions and sources.

Software	Version	Source
BEDTools	2.30.0	https://github.com/arq5x/bedtools2
BLAST	2.14.0	ftp://ftp.ncbi.nlm.nih.gov/blast/executables/blast+/
BlobToolKit	4.3.9	https://github.com/blobtoolkit/blobtoolkit
BUSCO	5.5.0	https://gitlab.com/ezlab/busco
bwa-mem2	2.2.1	https://github.com/bwa-mem2/bwa-mem2
Cooler	0.8.11	https://github.com/open2c/cooler
DIAMOND	2.1.8	https://github.com/bbuchfink/diamond
fasta_windows	0.2.4	https://github.com/tolkit/fasta_windows
FastK	1.1	https://github.com/thegenemyers/FASTK
GenomeScope2.0	2.0.1	https://github.com/tbenavi1/genomescope2.0
Gfastats	1.3.6	https://github.com/vgl-hub/gfastats
GoaT CLI	0.2.5	https://github.com/genomehubs/goat-cli
Hifiasm	0.16.1	https://github.com/chhylp123/hifiasm
HiGlass	1.13.4	https://github.com/higlass/higlass
MerquryFK	1.1.2	https://github.com/thegenemyers/MERQURY.FK
Minimap2	2.24-r1122	https://github.com/lh3/minimap2
MitoHiFi	2	https://github.com/marcelauliano/MitoHiFi
MultiQC	1.14; 1.17 and 1.18	https://github.com/MultiQC/MultiQC
Nextflow	23.04.1	https://github.com/nextflow-io/nextflow
PretextSnapshot	N/A	https://github.com/sanger-tol/PretextSnapshot
PretextView	0.2.5	https://github.com/sanger-tol/PretextView
purge_dups	1.2.3	https://github.com/dfguan/purge_dups
samtools	1.19.2	https://github.com/samtools/samtools
sanger-tol/ascc	0.1.0	https://github.com/sanger-tol/ascc
sanger-tol/blobtoolkit	0.4.0	https://github.com/sanger-tol/blobtoolkit
sanger-tol/curationpretext	1.4.2	https://github.com/sanger-tol/curationpretext
Seqtk	1.3	https://github.com/lh3/seqtk
Singularity	3.9.0	https://github.com/sylabs/singularity
TreeVal	1.4.0	https://github.com/sanger-tol/treeval
YaHS	1.1a.2	https://github.com/c-zhou/yahs

Author information

Contributors are listed at the following links:

- Members of the University of Oxford and Wytham Woods Genome Acquisition Lab
- Members of the Darwin Tree of Life Barcoding collective
- Members of the Wellcome Sanger Institute Tree of Life Management, Samples and Laboratory team
- Members of Wellcome Sanger Institute Scientific Operations – Sequencing Operations
- Members of the Wellcome Sanger Institute Tree of Life Core Informatics team
- Members of the Tree of Life Core Informatics collective
- Members of the Darwin Tree of Life Consortium

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Open Peer Review

Current Peer Review Status: ✔ ✔ ✔

Version 1

Reviewer Report 07 January 2026

https://doi.org/10.21956/wellcomeopenres.27394.r140037

✔ **Eduardo Zattara**
Instituto de Investigaciones en Biodiversidad y Medioambiente, Río Negro, Argentina

This data note reports the sequencing and assembly of the genome of a solitary species of sweat bee. Besides providing an instrumental resource for research on this particular species, it will also be informative when compared with genomes of other hymenopterans about the genomic evolutionary correlates of traits like sociality, size, behaviour, food preferences, etc. The report includes adequate detail both regarding the methodologies for sequencing and assembly, and quality assessment.

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Genomics, evolution of development, pollination biology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 30 December 2025

https://doi.org/10.21956/wellcomeopenres.27394.r140029

Pilar De la Rúa
University of Murcia, Murcia, Spain

The authors present the research flow for obtaining the genome of Lasioglossum villosulum. It appears that they have achieved a fairly complete version according to the data presented (N50 and single-copy genes present according to my search). It seems that these articles use a common template that always raises similar questions for me:

- The photo is not very good and the morphology of the specimen studied cannot be seen properly.
- The small dissected sample refers to a leg?
- On page 10 the scientific name of the specimen must be written in italics

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Conservation genetics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 13 November 2025

https://doi.org/10.21956/wellcomeopenres.27394.r136801

Christopher Wyatt

University College London, London, UK

The authors present the genome of *Lasioglossum villosulum,* which is of likely near complete genome based on the N50 and busco single copy genes present.
The paper is clearly written and explained. I could not find any section that was unclear or needed correction. There is an appropriate background to the species with highly detailed methodology.

Is the rationale for creating the dataset(s) clearly described?
Yes

Are the protocols appropriate and is the work technically sound?
Yes

Are sufficient details of methods and materials provided to allow replication by others?
Yes

Are the datasets clearly presented in a useable and accessible format?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Genomics, Evolutionary Biology, Bioinformatics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.