

The Future of Epigenetics: Emerging Technologies and Clinical Applications

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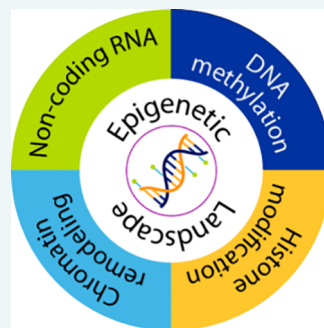
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ABSTRACT: Epigenetics, the study of heritable changes in gene expression that do not involve alterations to the DNA sequence, has emerged as a transformative field in biology and medicine, revealing how gene expression is modulated in response to internal and external cues. Its applications span from understanding fundamental biological processes and disease mechanisms to developing novel diagnostics and therapies, making it a cornerstone of modern biomedical research. This report explores data from the CAS Content Collection to examine recent transformative advances in epigenetic research, highlighting technological innovations that are revolutionizing our understanding of gene regulation and therapeutic applications. Environmental epigenetic research shows strong interest, demonstrating how external factors induce heritable epigenetic changes with implications for transgenerational inheritance. These findings bridge environmental exposures with health outcomes across generations, informing public health strategies and personalized medicine approaches. Disease applications span multiple pathologies, with cancer research leading epigenetic studies. Emerging applications in aging research, metabolic diseases, and autoimmune conditions demonstrate the field's expanding clinical relevance. Clinical translation has achieved significant success, with 13 FDA-approved epigenetic drugs primarily targeting hematological malignancies through HDAC inhibitors (6 drugs) and DNMT inhibitors (2 drugs). The robust clinical pipeline includes 37 ongoing trials across novel targets, with encouraging diversification beyond oncology into metabolic, neurological, and inflammatory disorders.

KEYWORDS: epigenetics, DNA methylation, histone modification, noncoding RNA, chromatin remodeling, epitranscriptomics



Epigenetics encompasses heritable changes in gene expression that occur without DNA sequence alterations, mediated through reversible modifications to DNA, histones, and RNA, as well as chromatin remodeling.^{1–6} These mechanisms include DNA methylation for gene silencing,^{7,8} histone modifications affecting chromatin accessibility, regulatory noncoding RNAs (ncRNAs),^{9,10} and structural chromatin changes that modulate transcriptional activity.^{11–13}

Epigenetic regulation is fundamental to developmental biology,^{14,15} disease pathogenesis (including cancer,¹⁶ diabetes,¹⁷ neurodegenerative,^{18,19} and autoimmune disorders),^{20–22} gene-environment interactions,²³ transgenerational inheritance,^{24,25} and therapeutic development.²⁶ By elucidating how genes are dynamically regulated beyond the static genome, epigenetics bridges genetics and environmental influences, offering insights into disease mechanisms and innovative therapeutic strategies.

The commercial significance of epigenetics has expanded dramatically, with the global epigenetics market valued at USD 1.84 billion in 2023 and projected to reach USD 6.77 billion by 2033.²⁷ This growth is driven by more than 10 U.S. FDA-approved epigenetic drugs including azacitidine (Vidaza) and vorinostat (Zolinza) for cancer treatment,²⁸ and with over 35 epigenetic therapies in clinical trials predominantly targeting

various malignancies but starting to show diversification beyond cancer. Major pharmaceutical companies including Merck,²⁹ and Celgene^{28,30} have invested significant capital and continue to do so in epigenetic drug development.

In this report, we explore data from the CAS Content Collection,³¹ the largest human-curated collection of published scientific information, to outline the progress made in epigenetic research, to identify key emerging concepts and challenges, and its societal impact, in an effort to understand how epigenetics shapes the future of gene regulation and inheritance. This comprehensive analysis examines the scientific foundations, clinical applications, and technological innovations shaping modern life sciences and healthcare.

The CAS Content Collection contains over 120,000 epigenetic-related publications (2000–2024), demonstrating a steep and continual growth in publications over the last two decades (Figure 1A). The field is dominated by journal articles

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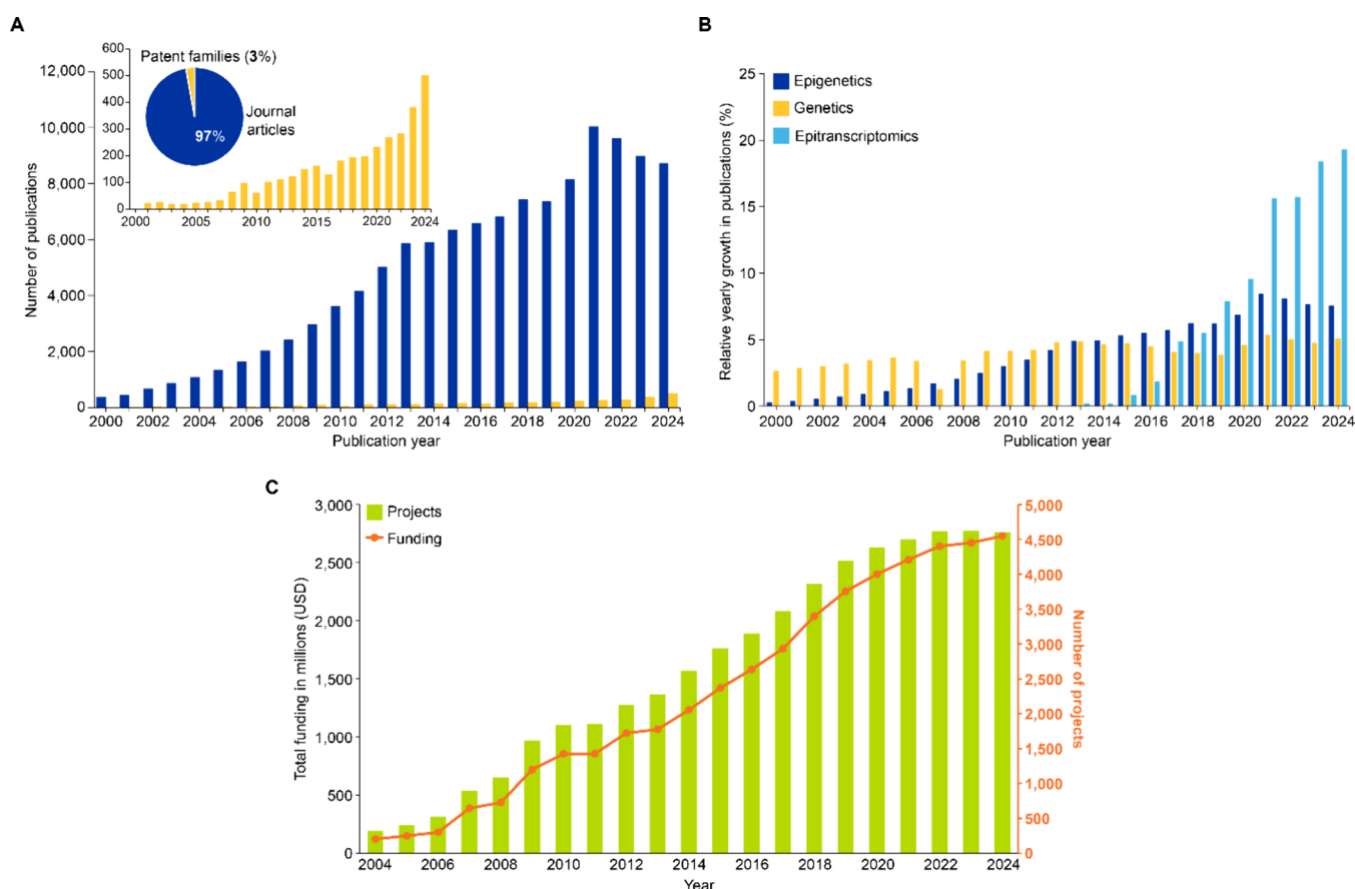


Figure 1. (A) Publication trends for epigenetic-related documents and (B) relative yearly growth in publications related to epigenetics, genetics, and epitranscriptomics. Data includes journal articles and patents extracted from the CAS Content Collection for the period 2000–2024. (C) Trends for number of projects funded and total funding granted to epigenetics related projects by the NIH (<https://reporter.nih.gov>).³³

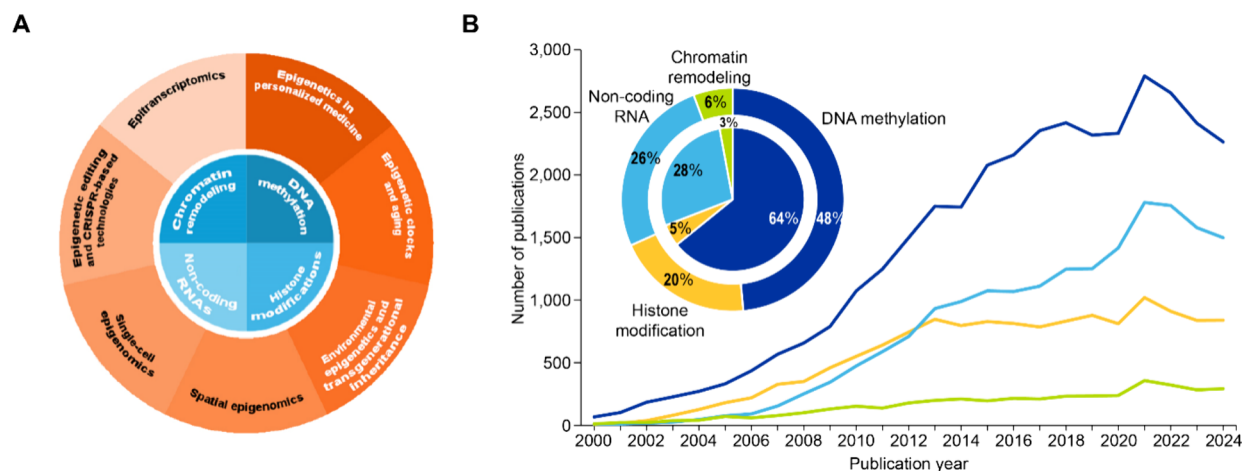


Figure 2. (A) Core mechanisms of epigenetic regulation (inner pie chart) and recent advances (outer donut chart). (B) Publication trends of epigenetic mechanisms based on journal and patent publications from the CAS Content Collection for the period 2000–2024. Outer donut and inner pie chart in the inset image represent journal and patent publications, respectively.

(97%), with patents comprising only 3% of publications (Figure 1A, inset), indicating that epigenetics remains primarily in the discovery and validation phase, though their sharp increase indicates growing commercial interest and translational potential in this rapidly evolving field. Notably, epigenetic publications have outpaced genetics publications since 2014 (Figure 1B). The emergence of epitranscriptomics as a distinct field³² especially noticeable after 2020 (Figure 1B), highlights

the expanding scope of epigenetic regulation beyond DNA and histone modifications. This growth trajectory aligns with substantial funding increases, as evidenced by total research funding climbing from USD 200 million in 2004 to over USD 4.5 billion in 2024, supporting approximately 2,700 projects annually³³ (Figure 1C). The consistent rise in both project numbers and funding levels, with average project funding increasing from USD 1.2 million to USD 1.7 million,

underscores sustained governmental and private sector commitment to advancing epigenetic research and its clinical translation.

For detailed insights on publication-related analysis please see Figures S1–S4 in Supporting Information. The analysis includes identification of leading research organizations in terms of research output and impact separately (Figure S1), as well as using a combination of both research output and impact (Figure S2), leading commercial and noncommercial patent assignees engaged in filing patents related to epigenetics and their geographical distribution (Figure S3), and leading scientific journals engaged in publishing epigenetic-related research (Figure S4).

CORE MECHANISMS OF EPIGENETIC REGULATION

Epigenetic mechanisms regulate gene expression through chemical modifications of DNA and chromatin structure without altering the DNA sequence. These mechanisms comprise four main classes: DNA methylation, histone modifications, ncRNA regulation, and chromatin remodeling (Figure 2), forming a layer of control in the cells that regulates gene expression and silencing.^{4,6}

Our analysis of epigenetic-related publications indicates that the early 2000s exhibited limited research output and represents the foundational era when epigenetics was being established as a field. Subsequently, all four mechanisms show accelerated growth, with DNA methylation experiencing the steepest growth, dominating the research landscape, both in terms of journal and patent publications, reflecting its status as the most established and clinically relevant epigenetic modification. This dominance is evident in the sustained growth reaching approximately 2,800 publications annually (Figure 2B). The dominance of DNA methylation research aligns with its clinical applications in cancer diagnostics³⁴ and therapeutics,³⁵ where methylation biomarkers^{36,37} and DNA methyltransferases (DNMT) inhibitors³⁸ have achieved regulatory approval.

ncRNAs and histone modifications also experienced growth, indicating increasing recognition of multiple regulatory layers. The growth in ncRNA research correlates with advances in RNA sequencing and CRISPR-based RNA targeting technologies, enabling functional characterization of previously inaccessible regulatory elements.^{39–43} The emergence of chromatin remodeling as a distinct research focus around 2008 reflects technological advances enabling genome-wide chromatin accessibility studies (Figure 2B).

The sustained interest across all mechanisms, despite recent plateaus, possibly reflective of a shift from discovery-based research to translational applications, suggests continued recognition of epigenetic targets for drug development, particularly in oncology and neurological disorders where epigenetic dysregulation is well-established.

Discussed briefly below are the four major mechanisms of epigenetic modifications.

DNA Methylation: The Primary Epigenetic Mark

DNA methylation is a fundamental epigenetic mechanism involving addition of methyl groups ($-\text{CH}_3$) to the cytosine rings at CpG dinucleotides in DNA, primarily resulting in transcriptional repression. This modification plays a critical role in X-chromosome inactivation,⁴⁴ genomic imprinting,⁴⁵ and transposon suppression.^{46,47}

DNA methylation is catalyzed by DNA methyltransferases (DNMTs). DNMT1 maintains existing methylation patterns

during replication,^{48,49} while DNMT3a/3b establishes new methylation patterns during development or in response to environmental cues.⁵⁰ DNMT3L, lacking catalytic activity and mainly expressed in early development, is restricted to the germ cells and thymus in adulthood.^{51,52} DNMTs target CpG islands, regions with high frequency of CpG dinucleotides, often located near gene promoters.^{8,53,54}

Aberrant methylation patterns have been identified in various diseases including cancer,⁵⁵ cardiovascular diseases,^{56,57} mental health disorders,⁵⁸ Alzheimer's disease,⁵⁹ autism,^{58,60} and metabolic syndromes.^{61,62} Hypermethylation of tumor suppressor gene promoters silences their expression promoting oncogenesis,⁶³ while global hypomethylation can activate oncogenes and cause genomic instability.^{64,65} DNA methylation patterns change with age,⁶⁶ including the accumulation of errors and loss of fidelity in methylation maintenance.^{67,68}

Histone Modifications: Dynamic Chromatin Regulation

Histone modifications are an epigenetic mechanism, since they can be inherited through cell division and can be influenced by environmental factors, potentially impacting development and disease states.⁶⁹

Post-translational histone modifications such as acetylation, methylation, phosphorylation, and ubiquitination, alter chromatin structure and gene accessibility. Histone acetyltransferases (HATs) and deacetylases (HDACs) regulate acetylation by catalyzing addition and removal of acetyl groups from histones, while methyltransferases (HMTs) and demethylases (HDMs) control methylation. These modifications primarily occur on lysine, arginine, serine, and threonine residues in histone tails, protrusions from the nucleosome core with increased accessibility.^{70–73}

Histone modifications affect the structure of chromatin, which is DNA packaged around histone proteins,^{74,75} regulating gene expression, DNA repair, and chromosome condensation during mitosis.^{74–76} Aberrant histone modifications can disrupt gene expression patterns and contribute to tumor development and metastasis,⁷⁷ and are linked to other diseases and disorders such as neurodegenerative⁷⁸ (Alzheimer's,^{79,80} Huntington's^{81,82}), autism,⁸³ aging-associated chromatin changes,⁸⁴ and immune dysregulation.^{85,86}

Histone modifications often work in combinations, forming a "histone code", with combinations of modifications on different residues synergizing or antagonizing to fine-tune chromatin states.^{87,88} Histone modifications are a dynamic and versatile system for regulating chromatin structure and gene expression, and their reversible nature makes them promising targets for therapeutic interventions.^{89–91}

Noncoding RNAs (ncRNAs): Epigenetic Regulators beyond the Genetic Code

ncRNAs constitute a diverse class of regulatory molecules that play critical regulatory roles in gene expression and chromatin dynamics without encoding proteins. These RNAs fine-tune gene processing by targeting mRNA for degradation or modulating transcriptional machinery.^{92,93}

While structural RNAs (tRNA, rRNA, small nuclear (snRNA), small nucleolar (snoRNA)) maintain essential cellular functions, three classes have emerged as primary epigenetic regulators: microRNAs (miRNAs), long noncoding RNAs (lncRNAs), and PIWI-interacting RNAs (piRNAs).^{10,94}

Our analysis of epigenetic-related publications indicates miRNAs dominated early investigations (2006–2014) appearing to reach a peak in 2021 (Figure 3). This trajectory reflects the

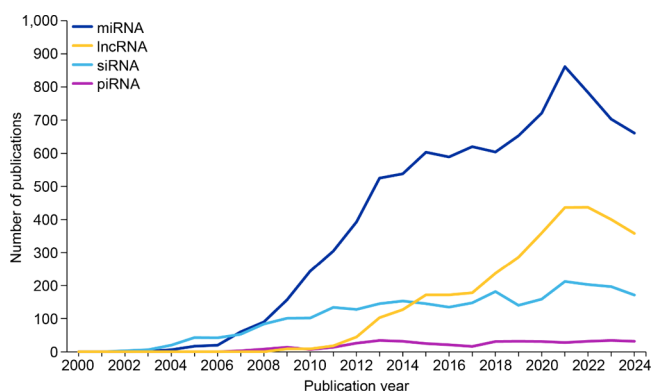


Figure 3. Publication trends of various ncRNAs in the epigenetics data set. Data includes journal articles and patents extracted from the CAS Content Collection for the period 2000–2024.

field's maturation from discovery to therapeutic application, with multiple miRNA-based therapies now in clinical trials.^{95,96} lncRNAs⁹⁷ emerged later but demonstrated remarkable growth from 2012 onward with this surge correlating with technological advances in RNA sequencing and functional characterization methods.^{98,99} Small interfering (siRNA) research appears to maintain steady output reflecting its established role in RNA interference therapeutics.^{100,101}

The modest but consistent publication trajectory of piRNA research (Figure 3) reflects the specialized nature of piRNA biology, which primarily operates in germline cells¹⁰² and early embryonic development,^{103–106} limiting its study to specific biological contexts compared to the ubiquitously expressed miRNAs and lncRNAs. However, emerging evidence suggests piRNAs function beyond germline contexts.¹⁰⁷ Somatic piRNA expression has been detected in neurons, where they regulate memory formation and storage¹⁰⁸ with implications in neurodegenerative diseases.^{109,110} In cancer, aberrant piRNA expression patterns correlate with genomic instability and tumor progression,^{111,112} with specific piRNAs serving as biomarkers for early detection.^{113,114} The stable but modest publication rate for piRNA research likely reflects the field's technical challenges: the complexity of the ping-pong amplification cycle, the requirement for specialized PIWI proteins, and the predominant germline expression pattern that necessitates specific model systems. As technologies for studying and manipulating piRNAs improve, we may witness an acceleration in publications similar to the lncRNA surge observed after 2012, particularly if somatic functions and therapeutic applications continue to emerge.

The overall publication trends demonstrate ncRNA research's evolution from mechanistic discovery (2000s) through functional characterization (2010s) to current therapeutic translation (2020s), with each RNA class following distinct developmental trajectories based on biological complexity and clinical applicability.

Mechanisms of ncRNA-Mediated Epigenetic Regulation. ncRNAs orchestrate epigenetic regulation through five principal mechanisms:^{115–121}

- (i) Chromatin remodeling: lncRNAs recruit chromatin-modifying complexes to specific genomic loci, influencing chromatin structure and gene expression. For example, X inactive specific transcript (XIST) mediates X-chromosome inactivation by recruiting polycomb repressive complex 2 (PRC2).

- (ii) Transcriptional regulation: miRNAs and lncRNAs modulate transcription by interacting with transcription factors or RNA polymerases. For example, the lncRNA HOX transcript antisense intergenic RNA (HOTAIR) recruits PRC2 to repress genes on a different chromosome.
- (iii) Post-transcriptional control: miRNAs bind to the 3' untranslated regions (UTRs) of target mRNAs (mRNAs), leading to degradation or translational inhibition. lncRNAs can act as molecular sponges to sequester miRNAs, preventing them from targeting mRNAs.
- (iv) RNA modification: Some ncRNAs guide RNA methylation (e.g., N6-methyladenosine, m⁶A) or editing, affecting RNA stability and translation.
- (v) Genome defense: piRNAs and siRNAs suppress transposable elements, protecting genomic integrity. siRNAs can guide heterochromatin formation in regions with repetitive sequences.

Chromatin Remodeling: Structural Gene Regulation

Chromatin remodeling dynamically modifies chromatin architecture between euchromatin (open, transcriptionally active) and heterochromatin (condensed, transcriptionally silent) states. The basic unit of chromatin is the nucleosome, composed of DNA wrapped around an octamer of histone proteins (H2A, H2B, H3, and H4).¹²² Chromatin remodeling is carried out by ATP-dependent complexes, such as switch/sucrose non-fermentable (SWI/SNF),¹²³ imitation switch (ISWI),^{124,125} chromodomain helicase DNA (CHD)-binding proteins,¹²⁶ and INO80 families,¹²⁷ utilizing ATP hydrolysis to power repositioning, ejection, or restructuring of nucleosomes.^{128–130}

Remodeling mechanisms include nucleosome sliding,^{131,132} histone eviction/exchange, and chromatin compaction/decompaction.^{129,133} These processes work in concert with DNA methylation and histone modifications to regulate gene expression, cell differentiation, and identity maintenance.^{134–136}

Chromatin remodeling dysregulation is linked to cancer,^{137,138} neurological disorders,¹³⁹ and developmental diseases.^{140,141} For example, mutations in chromatin remodelers like AT-rich interaction domain 1A (ARID1A) (a component of the SWI/SNF complex) are frequently observed in malignancies.^{142,143} Chromatin remodelers represent promising therapeutic targets for reversing aberrant gene expression.¹⁴⁴ Advances in single-cell technologies are revealing heterogeneity of chromatin states within cell populations, providing deeper insights into cell-specific regulation.^{145–147} Increasingly, chromatin remodeling is being recognized as a mediator of environmental influences (diet, stress, toxins, etc.) on gene expression and health.^{148–150} Future research directions include elucidating remodeler recruitment mechanisms, understanding ncRNA-chromatin interplay, and developing targeted epigenetic therapies.

RECENT ADVANCES IN EPIGENETIC MECHANISMS

Recent progress has elucidated how epigenetic modifications including DNA methylation, histone modifications, ncRNA regulation, and chromatin remodeling contribute to disease pathogenesis, particularly in cancer.^{151–156} These insights have driven the development of targeted epigenetic therapies, with emphasis on novel drug combinations that can simultaneously and effectively target multiple epigenetic pathways to enhance treatment efficacy. Emerging research on miRNAs¹⁵⁷ and other

ncRNAs^{119,158} as epigenetic regulators has identified new therapeutic targets, while advances in individual epigenetic profiling^{159,160} are aiding advancements in personalized medicine approaches.^{161–163}

Publication trends from the recent past demonstrate the rapidly expanding interest in these advances (Figure 4), with

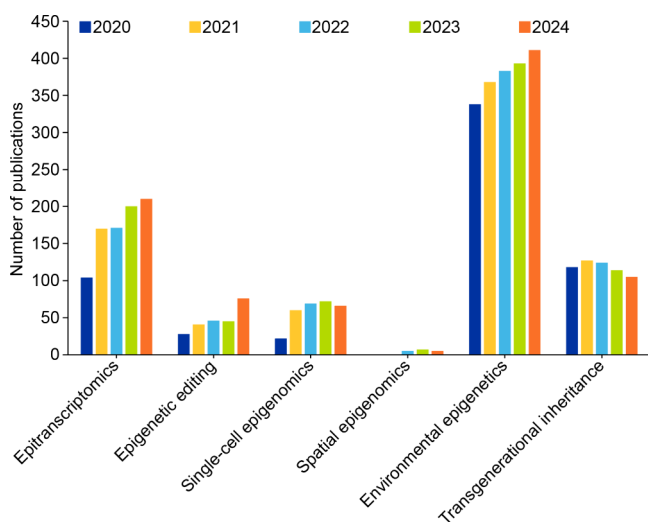


Figure 4. Publication trends for recent advances in epigenetic research. Data includes journal articles and patents extracted from the CAS Content Collection for the period 2000–2024.

environmental epigenetics seemingly leading research output, followed by epitranscriptomics showing consistent growth. Fields like epigenomic editing,^{164,165} single-cell epigenomics,¹⁶⁶ and transgenerational epigenetic inheritance^{25,167} appear to be growing at a moderate pace. Finally, newer fields like spatial epigenomics,¹⁶⁸ while still emerging with limited publications, represent cutting-edge frontiers in the discipline.

Epitranscriptomics

Epitranscriptomics is an emerging field in epigenetics that focuses on the study of chemical modifications to RNA molecules and their role in regulating gene expression and cellular functions.^{32,169} Just as epigenetics explores modifications to DNA and histones that influence gene activity without altering the underlying DNA sequence, epitranscriptomics investigates how RNA modifications affect RNA stability, translation, splicing, and other processes. This field has gained attention in recent years (Figure 1B and Figure 4) due to its potential to uncover new layers of gene regulation and its implications for health and disease offering new therapeutic and diagnostic opportunities.^{170,171}

Major RNA Modifications and Regulatory Machinery.

Over 170 distinct RNA modifications across mRNA, tRNA, rRNA, and ncRNAs have been identified, with N⁶-methyladenosine (m⁶A) being the most abundant and extensively studied modification in eukaryotic mRNA.^{172–174} Key modifications include: (i) m⁶A methylation at the N⁶ position of adenosine, influencing mRNA stability, splicing, export, and translation; (ii) m⁵C methylation at the N⁵ position of cytosine in mRNA and tRNA, affecting RNA stability and translation; (iii) pseudouridine (Ψ), an isomer of uridine, enhancing RNA stability and translation efficiency; and (iv) A-to-I editing – adenosine-to-inosine deamination, altering base pairing properties and affecting splicing and protein coding.^{172–174}

The epitranscriptomic machinery comprises three main protein classes: “writers” are enzymes that add modifications (e.g., METTL3/METTL14 complex for m⁶A methylation), “erasers” are enzymes that remove modifications (e.g., fat mass- and obesity-associated protein (FTO) and ALKB homologue 5, RNA demethylase (ALKBH5) for m⁶A demethylation), and “readers” are proteins that recognize and bind to modified RNA (e.g., YTH domain proteins for m⁶A recognition), which collectively regulate RNA modification dynamics.^{172,175–178}

Functional Roles and Disease Implications. RNA modifications regulate multiple cellular processes including mRNA stability and decay, translation control, alternative splicing, subcellular RNA localization, and immune recognition of self vs nonself RNA, playing critical roles in embryonic development, stem cell differentiation, and tissue-specific gene expression. Dysregulation of these modifications contributes to various pathologies. In cancer, m⁶A modifications are often altered in tumors¹⁷⁹ and are linked to tumor progression, metastasis,¹⁸⁰ and drug resistance.¹⁸¹ Aberrant RNA editing and modifications are associated with neurodegenerative diseases including Alzheimer’s,^{182,183} Parkinson’s,^{184–186} and amyotrophic lateral sclerosis (ALS).¹⁸⁷ Additionally, RNA viruses, including HIV and SARS-Cov-2, exploit host RNA modification machinery to regulate viral replication and immune evasion including epigenetic modifications.¹⁸⁸

Technological Advances and Therapeutic Applications. High-throughput sequencing technologies such as MeRIP-seq (m⁶A specific RNA immunoprecipitation sequencing)¹⁸⁹ and miCLI (m⁶A individual-nucleotide resolution cross-linking and immunoprecipitation)¹⁸⁹ enable genome-wide mapping of RNA modifications, complemented by mass spectrometry and chemical labeling techniques for modification detection and quantification.^{190,191} These advances facilitate therapeutic development targeting RNA modification enzymes.

Targeting components of RNA modification machinery (writers, erasers, and readers) is being explored as a strategy for treating diseases. For example, inhibitors of FTO, which are m⁶A demethylases, have shown promise in cancer therapy,^{192–194} while modified nucleotides in mRNA vaccines enhance stability and translation efficiency.^{195,196}

Future Directions. Current research focuses on deciphering the “epitranscriptome code,”¹⁹⁷ developing tools for precise *in vivo* manipulation of RNA modifications, and exploring modifications in ncRNAs.¹⁹⁸ Single-cell epitranscriptomics is revealing cell-type-specific RNA modification patterns and their functional consequences.^{199,200} Investigations into role of RNA modifications in circadian rhythms,^{201,202} stress responses,²⁰³ and environmental adaptation^{204,205} are currently being pursued and is expanding our understanding of RNA modification dynamics. As such, the integration of epitranscriptomic and epigenetic mechanisms represents a critical frontier in gene regulation research.

Epigenetic Editing and CRISPR-Based Technologies

The advent of CRISPR-Cas9 technology has revolutionized genetic engineering, and its application to epigenetics is no exception.^{206–208} Epigenetic editing involves the targeted modification of epigenetic marks without altering the DNA sequence, offering a powerful tool for studying gene regulation and developing novel therapies.

The CRISPR-Cas9 system employs guide RNAs (gRNAs) to direct catalytically inactive Cas9 (dCas9) fused with epigenetic effector domains to specific genomic loci. Key effector domains

include DNMT (e.g., DNMT3A), DNA demethylases (e.g., TET1), histone modifiers (e.g., p300, HDACs), and chromatin remodelers, enabling precise addition or removal of epigenetic modifications.^{209–211}

CRISPR-Cas9 technology enables selective activation or repression of gene expression by targeting regulatory elements including enhancers and promoters, allowing researchers to directly test the causal roles of specific epigenetic marks in gene regulation,^{212,213} cellular differentiation,^{214,215} and disease.²¹⁶ Additionally, it allows examination of epigenetic inheritance across cell divisions and development, while allowing generation of disease models driven by epigenetic dysregulation such as cancer, neurological, and metabolic disorders.^{208,211,217}

Therapeutic applications include silencing disease-causing genes (oncogenes, viral genes) and reactivating beneficial silenced genes (tumor suppressors) in cancer²¹⁸ and hypercholesterolemia²¹⁹ among others.^{214,220,221} This approach offers the potential of highly personalized therapies with minimal off-target effects and applications in cellular reprogramming for regenerative medicine. Key advantages of CRISPR-based epigenome editing include high targeting precision, versatility through interchangeable effector domains, and reduced mutagenic risk compared to gene editing.

Advancing this technology requires developing high-fidelity Cas variants and optimized gRNA designs to minimize off-target effects. Furthermore, multiplexed targeting will enable investigation of complex regulatory networks, while improved delivery methods (viral vectors, nanoparticles) will facilitate clinical translation.^{208,217,222} Large-scale functional screens using CRISPR-based epigenome editing may help identify key epigenetic regulators, and integration with immunotherapy or small-molecule drugs may enhance therapeutic efficacy. Clinical trials are currently evaluating safety and efficacy of these approaches (e.g., NCT06671093²²³).^{208,217,224}

Single-Cell Epigenomics

Single-cell epigenomic technologies enable profiling of epigenetic modifications at the resolution of individual cells, providing unprecedented insights into cellular diversity and function,^{225–227} and overcoming drawbacks of traditional bulk sequencing methods which looks at average epigenetic marks across millions of cells. Techniques including single-cell ATAC-seq (assay for transposase-accessible chromatin using sequencing) and single-cell ChIP-seq (chromatin immunoprecipitation sequencing) which profile chromatin accessibility and histone modifications in individual cells and are at the forefront of this trend.^{228–234}

Applications span from developmental biology including mapping epigenetic landscapes during embryogenesis,^{15,235,236} cellular and tissue differentiation,²³⁷ cell fate decisions,²³⁸ and immune system heterogeneity²³⁹ to cancer research encompassing uncovering intratumoral epigenetic heterogeneity relevant to clonal evolution,^{147,240,241} drug resistance,^{242,243} and metastasis.²⁴¹ The technology is also used to elucidate epigenetic changes in neurodevelopment,²⁴⁴ neurodegeneration,²⁴⁵ and immune cell differentiation,²⁴⁶ informing understanding of autoimmune diseases and immunotherapy responses. Future directions include integration with spatial transcriptomics,²⁴⁷ temporal profiling of epigenetic marks in response to cellular processes or environmental stimuli,²⁴⁸ and clinical applications for biomarker identification and discovery as well as development of personalized medicine.^{249,250}

Spatial Epigenomics

Spatial epigenomics is an emerging and cutting-edge technology in epigenetic research that combines the study of epigenetic modifications with spatial context within tissues unlike in single-cell approaches.²⁵¹ This approach bridges single-cell epigenomics and histology, revealing how epigenetic regulation varies across tissue regions and influences cellular organization, function, and communication in health and disease.^{251,252}

Applications include mapping tissue patterning during organogenesis guided by epigenetic changes, identifying spatially distinct epigenetic signatures within tumors (core versus invasive margins) which may drive metastasis or drug resistance, characterizing region-specific epigenetic changes in brain tissues or cell types providing insights into neurodevelopment, plasticity, and neurodegenerative diseases, understanding epigenetic regulation across immune cell niches, and influence of epigenetic states in tissue microenvironment.^{168,253,254}

Environmental Epigenetics

Environmental factors such as diet,²⁵⁵ stress, toxins, and lifestyle²⁵⁶ can induce epigenetic changes affecting gene expression through one of the many epigenetic mechanisms.^{257,258} These modifications contribute to the development of diseases such as cancer,²⁵⁹ metabolic disorders,²⁶⁰ and neurological conditions^{261,262} by altering cellular function and tissue homeostasis.²⁶³ Specific influences include: nutrient availability (e.g., folate, vitamin B12) affecting DNA methylation patterns;²⁶⁴ chemical exposures (e.g., bisphenol A (BPA), pesticides, air pollutants) altering epigenetic marks leading to long-term health consequences;^{265–267} psychological and physiological stress inducing epigenetic changes in stress-response genes and modulating mental health;^{268,269} and lifestyle factors (e.g., smoking,²⁷⁰ alcohol,²⁷¹ physical activity²⁷²) influencing disease risk via modulating epigenetic states.

Transgenerational Inheritance

Transgenerational inheritance refers to the transmission of environmentally induced epigenetic changes across multiple generations independent of alterations in the DNA sequence.^{273,274} This phenomenon has been observed in plants²⁷⁵ and animals²⁷⁶ suggesting that environmental exposures experienced by one generation can affect the health and development of subsequent generations. While reports do exist with regards to humans there are concerns about the extent and significance of it^{167,277,278} with the field plagued by challenges.^{279,280} Mechanisms include germline epigenetic modifications,²⁸¹ epigenetic memory persisting through developmental reprogramming, maternal-fetal crosstalk,²⁸² and sperm RNA-mediated information transfer.²⁸³

Example of transgenerational epigenetic inheritance includes the Dutch Hunger Winter (1944–1945): altered DNA methylation patterns and increased risk of metabolic disorders was observed in subsequent generations of individuals who were subject to famine exposure in Netherlands.^{284,285} Animal studies have also provided examples of transgenerational effects of endocrine disruptors^{286,287} (e.g., vinclozolin²⁸⁸) on reproduction, behavior, and disease susceptibility, while in humans paternal smoking²⁸⁹ and maternal stress²⁹⁰ have been linked to epigenetic changes in offspring.

While the mechanisms underlying transgenerational epigenetic inheritance are not fully understood, their effect on raising the risk profile for diseases such as obesity, diabetes, and cardiovascular disorders in descendants of exposed individuals

Table 1. Summary of Epigenetic Mechanisms, Biomarkers, U.S. FDA Approved Drugs and Clinical Trials for Various Diseases^a

	Epigenetic mechanisms				U.S. FDA approved drugs		Clinical trials			Biomarkers			
	DNA methylation	Histone modification	ncRNA	Chromatin remodeling	√/X	√/X	Highest phase of development			DNA methylation	Histone modification	ncRNA	Chromatin remodeling
Cancer	11,917	4,287	7,506	1,364	√	√	Phase III			2,202	422	1,530	134
Aging	2,545	793	945	220	X	X	-			479	52	158	19
Neurological	2,147	781	942	194	X	X	-			303	57	198	13
Blood disorders	1,790	759	913	265	X	X	-			182	41	119	20
Cardiovascular	905	391	712	84	X	X	-			143	31	141	6
Metabolic	2,142	722	1,080	111	X	X	-			279	40	160	10
Respiratory	544	202	299	30	X	X	-			74	19	54	4
Autoimmune	927	379	637	52	X	X	-			140	45	149	3
Gastrointestinal	154	59	107	9	X	X	-			30	10	25	2
Infectious	2,281	931	1,454	280	X	√	Phase I			278	7	166	13

^aThe numbers correspond to journal and patent publications from the CAS Content Collection.

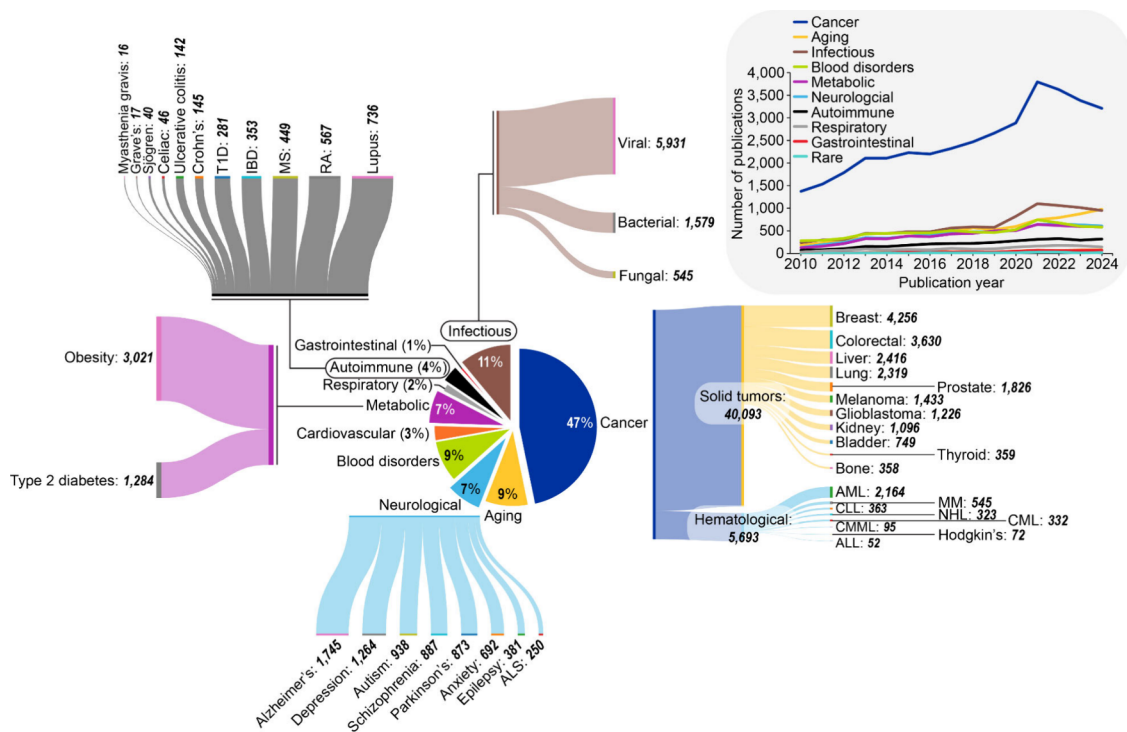


Figure 5. Relative distribution of epigenetic-related publications among various disease types depicted in the pie chart at the center. The individual Sankey charts show the distribution of publications among subtypes of diseases. Inset line graph at the top right shows publication trends of the broader disease categories. Data includes journal articles and patents extracted from the CAS Content Collection for the period 2000–2024.

means that it remains an area of active research. Understanding transgenerational effects can inform policies to reduce exposure to harmful environmental factors, particularly during critical windows of development (e.g., pregnancy).^{291–293} However, challenges persist – difficulty in separating the contributions of genetic and epigenetic factors to inheritance as well as in successfully replicating experiments highlighting the need for rigorous experimental design.

■ EPIGENETICS IN HEALTH AND DISEASE

Epigenetic modifications guide embryonic development and cell fate decisions by activating or silencing specific gene sets and epigenetic changes accumulate with age, influencing longevity and age-related diseases.^{294,295} Dysregulation of these processes

underlies diverse pathologies including cancer,^{16,296} cardiovascular disorders,²⁹⁷ neurodegenerative diseases,^{298–300} and metabolic syndromes.^{301,302} Epigenetic diseases are conditions caused by chemical modifications on DNA and its associated proteins (histones) without altering the underlying DNA sequence. These modifications, called epigenetic marks, can affect gene expression and play a role in various biological processes.^{4,303,304} Summarized in Table 1 are the core epigenetic mechanisms and related biomarkers co-occurring with different diseases as well as information about U.S. FDA approved epigenetic drug availability and ongoing clinical trials.

Cancer: Epigenetic Dysregulation in Tumorigenesis

While genetic mutations have long been recognized as drivers of cancer, epigenetic alterations are now understood to play an

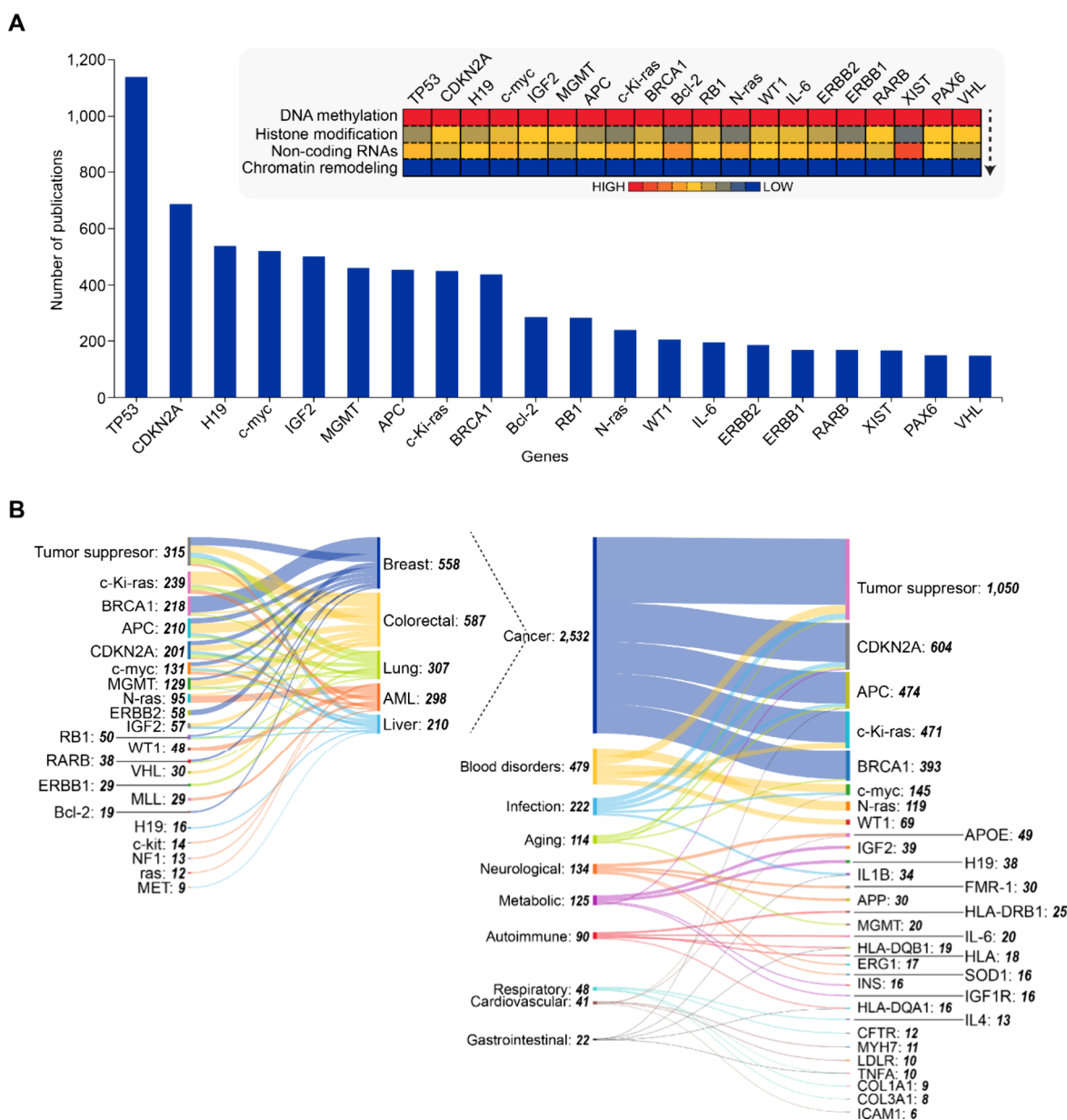


Figure 6. (A) Leading genes associated with epigenetic publications based on CAS indexing. Inset heat map table shows co-occurrence of epigenetic mechanisms with individual genes. (B) Sankey graphs depicting disease and gene co-occurrences in the epigenetics data set (restricted to top 5 genes for each disease indication). Data includes journal and patent publications from the CAS Content Collection for the period 2000–2024.

equally critical role in tumorigenesis.³⁰⁵ The epigenetic disease research landscape (Figure 5, pie chart) reveals **cancer's** dominance with 47% of publications, reflecting both the field's maturity and clinical translation success with temporal analysis demonstrating cancer research's sustained growth (Figure 5, inset). Solid tumors account for an overwhelming majority of publications with breast, colorectal, liver, and lung cancer leading in terms of research activity (Figure 5, sankey). Among hematological malignancies, acute myeloid leukemia (AML) dominates, followed by chronic lymphocytic leukemia (CLL) and multiple myeloma (MM). This distribution observed based on publication data from the CAS Content Collection aligns with U.S. FDA-approved epigenetic therapies targeting these malignancies, including azacitidine^{306,307} and decitabine^{308,309} used for treatment of AML and vorinostat for cutaneous T-cell lymphoma.³¹⁰

Epigenetic regulation in cancer involves three primary mechanisms: (i) DNA methylation³¹¹ – hypermethylation of promoter regions often leads to the silencing of tumor suppressor genes (e.g., p16,^{312,313} BRCA1^{314,315}); global hypomethylation, on the other hand, can activate oncogenes and promote genomic instability;⁶⁴

(ii) histone modifications⁷⁷ – alterations in histone acetylation, methylation, and phosphorylation can change chromatin structure and gene expression – for example, loss of histone acetylation is associated with the repression of tumor suppressor genes, while gain of repressive histone marks (e.g., H3K27me3^{316,317}) can silence critical regulatory genes;

(iii) ncRNAs³¹⁸ – dysregulation of miRNAs (e.g., miR-21 overexpression) can promote cancer progression by targeting tumor suppressors or oncogenes.^{16,319,320}

The gene-epigenetic mechanism co-occurrence heatmap (Figure 6A, inset) reveals distinct patterns: DNA methylation's dominance reflecting its role as an important epigenetic mechanism; chromatin remodeling shows minimal co-occurrence across most genes, suggesting either its role in broader architectural changes rather than gene-specific regulation; while publications associated with ncRNA and histone modifications lie between the two extremes – DNA methylation and chromatin remodeling. Our analysis of gene and disease co-occurrences (Figure 6B) illustrates cancer's central position, with tumor suppressor genes showing highest connectivity. Other genes with high co-occurrences across multiple cancers include CDKN2A,^{321,322} followed by APC^{323,324} and c-Ki-ras (KRAS³²⁵). This pattern reflects these genes' roles as epigenetic hubs subject to methylation-mediated silencing across diverse malignancies.

Epigenetic Clocks: Quantifying Biological Aging

Epigenetic clocks are a groundbreaking concept in epigenetics implicating computational models that predict biological age based on DNA methylation patterns,^{326,327} emerging as powerful tools for studying aging,³²⁸ longevity, and age-related diseases.³²⁹ They can reveal discrepancies between an individual's biological age and chronological age, providing insights into the aging process and the impact of lifestyle and environmental factors. These models exploit predictable age-related changes in DNA methylation, where specific genomic regions become hypermethylated or hypomethylated in reproducible patterns.^{326,330,331}

Research publication trends (Figure 5) demonstrate that aging represents 9% of epigenetic research explicitly mentioning disease and aging, significantly smaller than cancer research (47%), yet showing consistent growth from 2010 to 2024 (Figure 5, inset). This proportion reflects aging research's specialized but expanding role within the broader epigenetic landscape dominated by cancer applications.

The first-generation epigenetic clock, Horvath's clock, was developed by Steve Horvath in 2013 and utilizes DNA methylation data from 353 CpG sites across multiple tissues and cell types to estimate biological age,³³² while Hannum's clock, developed in the same year,³³³ employs 71 CpG sites, primarily in blood. More recent clocks like GrimAge³³⁴ and PhenoAge³³⁵ incorporate additional biomarkers (e.g., plasma proteins) to improve accuracy³³⁶ and predict mortality and disease risk.³³⁷

Epigenetic clocks use machine learning algorithms to analyze DNA methylation data and help in prediction of biological age including age acceleration (if epigenetic age > chronological age) or deceleration (if epigenetic age < chronological age), with acceleration associated with increased age-related disease and mortality risk.^{327,338}

Applications span intervention studies evaluating antiaging treatments (caloric restriction, exercise, pharmacological agents),^{295,339,340} longevity research enabling researchers to study the effects of genetics, lifestyle, and environmental factors on aging, and age-related disease prediction. In longevity research centenarians and individuals with exceptional longevity often exhibit slower epigenetic aging, providing clues to the genetic and environmental factors that promote healthy aging.^{341–343} Current research explores reversibility through senolytics and epigenetic regulator-targeting compounds.³³⁹

Neurological Disorders: Epigenetic Mechanisms in Neurodegeneration

Epigenetic modifications have been linked to neurodegenerative diseases such as Alzheimer's,^{262,344} Parkinson's,³⁴⁵ and ALS,^{346–348} as well as mental health disorders such as depression,³⁴⁹ and schizophrenia.³⁵⁰ The substantial research output reflects growing recognition of epigenetic mechanisms in neuropsychiatric and neurodegenerative pathologies (Figure 5). Epigenetic alterations affect genes involved in amyloid-beta production (APP;¹⁸² Figure 6) and tau phosphorylation in Alzheimer's disease, with dysregulation of miRNAs,³⁵¹ such as miR-29^{352,353} and miR-34³⁵⁴ being associated with cognitive decline. Parkinson's disease shows epigenetic changes in mitochondrial function genes (e.g., PINK1³⁵⁵) and α -synuclein aggregation pathways.³⁵⁶ Histone acetylation^{78,357} and DNA methylation³⁵⁸ patterns are disrupted in Parkinson's disease models, affecting neuronal survival.

Autism spectrum disorders³⁵⁹ exhibit dysregulation of synaptic genes (e.g., SHANK3³⁶⁰) through aberrant DNA methylation^{361,362} and histone modifications,^{83,363,364} with environmental factors such as prenatal exposure to valproic acid shown to induce epigenetic changes in animal models.³⁶⁵ In epilepsy, epigenetic mechanisms regulate ion channel genes and synaptic plasticity genes, contributing to seizure susceptibility, with histone modifications³⁶⁶ and miRNA dysregulation³⁶⁷ being observed in epilepsy models. Neurodevelopmental disorders exemplify critical epigenetic regulation as exhibited by mutations in the MECP2 gene, encoding a methyl-CpG-binding protein, causing Rett syndrome³⁶⁸ and FMR1 hypermethylation in Fragile X syndrome.^{369,370}

Epigenetic alterations are increasingly recognized as biomarkers for neurological disorders.³⁷¹ Methylation patterns of genes such as BDNF^{372,373} and COMT³⁷⁴ are associated with cognitive function and psychiatric disorders, while specific histone marks (e.g., H3K27me3) are linked to gene regulatory networks in neurodegeneration.⁷⁸ Circulating miRNAs,³⁷⁵ particularly miR-132^{376,377} and miR-124³⁷⁸ have shown potential as noninvasive biomarkers for diagnosis of Alzheimer's and Parkinson's disease, respectively, offering potential for early detection and therapeutic monitoring.

Cardiovascular: Environmental-Epigenetic Interactions

Epigenetic mechanisms contribute to the pathogenesis of various cardiovascular diseases^{297,379–381} though our analysis of publication data suggests smaller fraction of research interest when compared to cancer (Figure 5 and Figure 6). For instance, in atherosclerosis, DNA methylation⁵⁷ and histone modifications³⁸² regulate genes involved in inflammation, lipid metabolism, and endothelial function;³⁸³ hypomethylation of pro-inflammatory genes (e.g., IL-6³⁸⁴) and hypermethylation of anti-inflammatory genes (e.g., PPAR γ ³⁸⁵) promote plaque formation. Epigenetic changes in genes regulating vascular tone (e.g., ACE,³⁸⁶ eNOS³⁸⁷) and sodium homeostasis contribute to hypertension; environmental factors such as high-salt diet^{388–390} and stress can induce these epigenetic alterations. Epigenetic regulation of ion channel genes (e.g., SCN5A,³⁹¹ KCNQ1³⁹²) can predispose individuals to arrhythmias. Similarly, epigenetic modifications drive pathological cardiac remodeling, including hypertrophy and fibrosis;³⁸¹ DNA methylation,³⁹³ histone modifications³⁹⁴ and miRNAs (e.g., miR-208³⁹⁵) are key regulators of these processes. In ischemic heart disease and stroke, DNA methylation³⁹⁶ and histone

modifications³⁹⁷ alter the expression of genes that govern cell survival, stress responses, and inflammatory pathways.

Epigenetic alterations are increasingly documented as biomarkers for cardiovascular disease diagnosis, prognosis, and risk stratification.^{398,399} Methylation patterns of genes such as F2RL3⁴⁰⁰ and AHR⁴⁰¹ are associated with cardiovascular risk and outcomes. Specific histone marks (e.g., H3K27ac^{402,403}) are linked to gene regulatory networks in heart failure and atherosclerosis. Circulating miRNAs (e.g., miR-126,⁴⁰⁴ miR-499⁴⁰⁵) are used as biomarkers for acute myocardial infarction and heart failure.^{398,399,406}

Metabolic Disorders

Epigenetic modifications regulate genes controlling insulin sensitivity, fat storage, and inflammation in metabolic diseases.^{17,407,408} Environmental factors including diet and exercise induce persistent epigenetic changes contributing to insulin resistance and metabolic dysfunction.⁴⁰⁹ Metabolic diseases represent a sizable fraction of epigenetic research (Figure 5), with obesity studies and type 2 diabetes dominating this field. The substantial publication volume reflects the global epidemic of metabolic diseases⁴¹⁰ and growing understanding of epigenetic contributions to metabolic dysfunction. The steady but modest growth in metabolic epigenetic research (Figure 5, inset) contrasts with the exponential increase in cancer studies, suggesting significant untapped potential in this area. Recent studies reveal epigenetic transgenerational inheritance of metabolic phenotypes^{292,293,411} with these findings have profound implications for public health interventions targeting metabolic disease prevention across generations.

- Obesity: DNA methylation and histone modifications regulating adipogenesis, appetite control, and energy expenditure genes, with dysregulated miRNAs (e.g., miR-103, miR-143) being associated with obesity and adipose tissue dysfunction.⁴¹²
- Type 2 diabetes (T2D): Epigenetic changes in pancreatic β cells, liver, and skeletal muscle affecting insulin production and sensitivity, including DNA methylation of PPARGC1A and IRS1 genes.^{17,413}
- Nonalcoholic fatty liver disease (NAFLD): DNA methylation and histone modifications regulate hepatic lipid metabolism and inflammation, with miRNAs (e.g., miR-34a, miR-122) contributing to disease progression.^{414,415}
- Metabolic syndrome: Epigenetic changes in glucose metabolism, lipid metabolism, and blood pressure regulation genes. Metabolic syndrome models characterized by histone modifications and miRNA dysregulation.^{11,416,417}
- Cardiometabolic diseases: Epigenetic mechanisms link metabolic dysregulation to cardiovascular complications through DNA methylation of genes including FTO⁴¹⁸ and ABCA1,⁴¹⁹ associated with atherosclerosis and hypertension risk.^{11,297} These findings demonstrate epigenetic mechanisms as critical links between environmental exposures and metabolic disease development.

Epigenetic biomarkers for metabolic diseases include: methylation patterns of TXNIP^{420,421} and SREBF1 genes associated with T2D and NAFLD;^{422,423} specific histone marks (e.g., H3K9Ac⁴²⁴) linked to metabolic gene regulatory networks; and circulating miRNAs (e.g., miR-375,⁴²⁵ miR-21⁴²⁶) serving as biomarkers for T2D and obesity. These

biomarkers⁴²⁷ enable early detection, risk stratification, and therapeutic monitoring in metabolic disorders.

Autoimmune Diseases: Immune Dysregulation through Epigenetic Alterations

Aberrant methylation of immune tolerance genes can activate self-reactive T cells, leading to autoimmune conditions. Despite their clinical importance, autoimmune diseases represent a modest portion of epigenetic research (Figure 5), with steady but limited growth from 2010 to 2024 compared to the exponential increase in cancer studies (Figure 5, inset). This research gap suggests significant potential for expanded investigation in autoimmune epigenetics. Within autoimmune diseases, systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), multiple sclerosis (MS), inflammatory bowel disease (IBD), and type 1 diabetes (T1D)⁴²⁸ have emerged as primary research areas.

- Systemic lupus erythematosus (SLE) pathogenesis associated with dysregulated miRNAs (miR-21, miR-148a).⁴²⁹
- Rheumatoid arthritis (RA) involves DNA methylation changes in synovial fibroblasts and immune cells promoting joint inflammation, while histone modifications (H3K27ac) regulate pro-inflammatory cytokine production (TNF- α , IL-6).⁴³⁰
- Multiple sclerosis (MS) exhibits epigenetic changes in T and B cells affecting immune regulation and myelin destruction genes, with miRNA dysregulation (e.g., miR-326,⁴³¹ miR-155⁴³²) linked to disease progression.
- Type 1 diabetes (T1D) involves DNA methylation and histone modifications in pancreatic β cells and immune cells contributing to autoimmune destruction of insulin-producing cells, with miRNAs (e.g., miR-21, miR-34a) implicated in pathogenesis.⁴³³
- Inflammatory bowel disease (IBD) such as Crohn's and ulcerative colitis exhibit epigenetic changes in intestinal epithelial and immune cells driving chronic inflammation along with association with miRNA dysregulation (e.g., miR-21, miR-155).^{434–436}

Epigenetics in Personalized Medicine

Integration of epigenetic data enables personalized treatment strategies based on individual epigenetic profiles, aiding development of highly precise diagnostic tools, prognostic markers, and therapeutic approaches particularly for complex diseases including cancer, neurological disorders, and metabolic conditions.^{437,438} Epigenetic modifications can serve as tissue-specific biomarkers⁴³⁹ providing insights into disease mechanisms and progression.

Examples of disease-specific epigenetic patterns include: cancer (tumor suppressor gene hypermethylation and global hypomethylation), neurological disorders (aberrant methylation and histone modifications in Alzheimer's, Parkinson's, and autism spectrum disorders), and metabolic diseases (epigenetic changes in glucose and lipid metabolism genes contributing to diabetes and obesity).

Epigenetic biomarkers enable early disease detection and diagnosis, for example through liquid biopsies utilized for detecting cancer-specific DNA methylation patterns in blood or other bodily fluids for noninvasive diagnosis and monitoring,⁴⁴⁰ and prenatal testing using epigenetic markers in maternal blood to assess fetal health.^{441,442} Prognostically epigenetic markers can predict disease outcomes and response to therapy. For

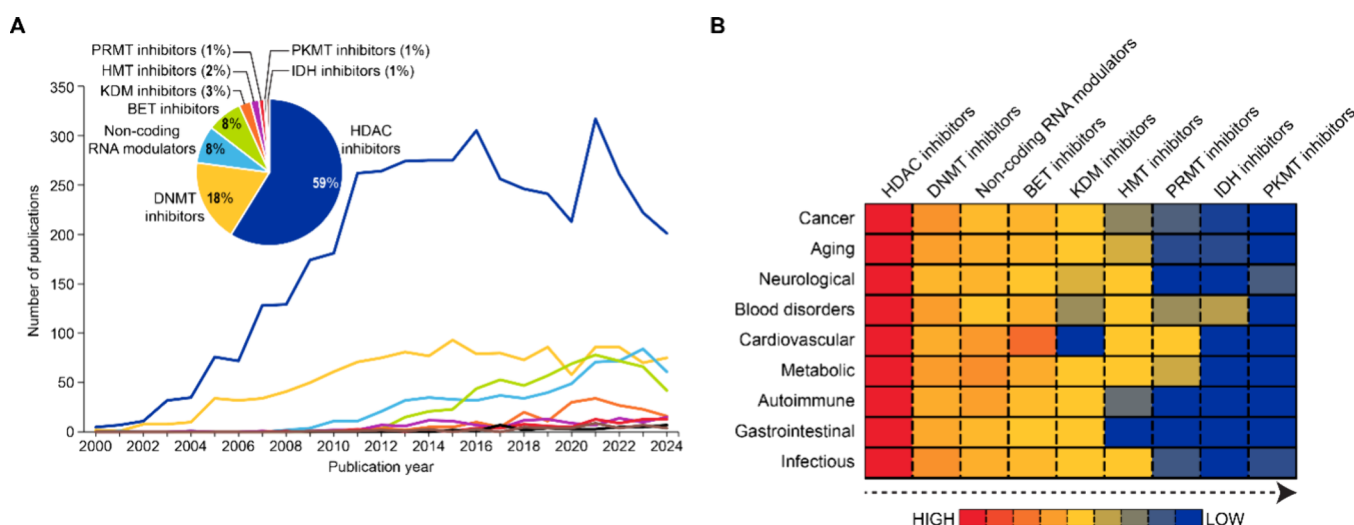


Figure 7. (A) Distribution and publication trends of epi-drug classes in the epigenetics data set. (B) Heat map showing co-occurrence of epi-drug classes with diseases. Data includes journal and patent publications from the CAS Content Collection for the period 2000–2024. Abbreviations used: HDAC, histone deacetylase; DNMT, DNA methyltransferase; BET, bromodomain and extra-terminal domain; KDM, histone lysine demethylase; HMT, histone methyltransferase; PRMT, arginine methyltransferase; PKMT, lysine methyltransferase; IDH, isocitrate dehydrogenase.

example, methylation status predicts outcomes and metastasis likelihood in some cancers,^{443,444} while epigenetic changes in psychiatric disorders may predict medication response.⁴⁴⁵

Epigenetic drugs targeting DNA methyltransferases (DNMTs) and histone deacetylases (HDACs) are clinically established and continue to be developed. DNMT inhibitors (e.g., azacitidine, decitabine) treat myelodysplastic syndromes and AML, while HDAC inhibitors address certain lymphomas and multiple myeloma.^{26,446} Combination therapies integrating epigenetic drugs with chemotherapy and immunotherapy enhance treatment efficacy, representing a key advancement in personalized therapeutic approaches.^{26,446}

Epigenetic Therapeutics: From Mechanisms to Clinical Applications of Epi-Drugs

Epigenetic drugs (epi-drugs) reverse aberrant epigenetic modifications to restore normal gene expression in diseases characterized by epigenetic dysregulation.^{26,305,447} The therapeutic landscape (Figure 7) reveals HDAC inhibitors dominating with 59% of publications, followed by DNMT inhibitors (18%), and RNA modulators (8% for ncRNA and general RNA modulators). Table 2 summarizes mechanisms of action of epi-drugs, with these mechanisms highlighting how epi-drugs target specific components of the epigenetic machinery to restore normal gene expression patterns, making them promising tools for treating cancers and other diseases driven by epigenetic dysregulation. Table S2 lists exemplary epi-drugs approved for clinical application.

- **Histone deacetylase (HDAC) inhibitors** function as broad-spectrum lysine deacylases that target both histone and nonhistone substrates while removing diverse acyl modifications beyond acetylation.^{448,449} HDACs were originally known for preventing the removal of acetyl groups from histones, which relaxes chromatin and increases gene expression, including tumor suppressor genes. However, researchers now recognize that HDACs have much broader substrate specificity.^{450,451} These enzymes deacetylate numerous nonhistone proteins including p53, NF- κ B, HSP90, α -tubulin, and STAT3, thereby regulating protein stability, subcellular localization, and

protein–protein interactions critical for cellular homeostasis.⁴⁵² Beyond acetylation, HDACs remove various acyl modifications including propionylation, butyrylation, crotonylation, and succinylation from lysine residues, expanding their regulatory impact on cellular metabolism and gene expression.⁴⁵³ This multifaceted biology explains the pleiotropic effects of HDAC inhibitors in cancer therapy, including induction of apoptosis, cell cycle arrest, and immune modulation. HDAC inhibitors also affect nonhistone proteins, such as transcription factors⁴⁵⁴ and chaperone proteins,⁴⁵⁵ further contributing to their anticancer effects. The dramatic surge in publications from early 2000s to early 2010s followed by stabilization (Figure 7A) indicates transition from discovery to clinical implementation. HDAC inhibitors currently approved for clinical use include vorinostat (Zolinza), Romidepsin (Istodax), belinostat (Beleodaq), panobinostat (Farydak) (canceled by U.S. FDA in 2022 but still approved by European Medicines Agency (EMA)), givinostat (Duvyzat) and chidamide (Tucidinostat) (approved by National Medical Products Administration (NMPA) in China and Pharmaceuticals and Medical Devices Agency (PMDA) in Japan) (Table S2).

- **DNA methyltransferases (DNMT) inhibitors**, representing the second-largest drug class in terms of associated research publications (Figure 7A, inset pie chart), block DNA methyltransferases (DNMTs) aiding reactivation of silenced tumor suppressor genes in cancer.^{456–458} DNMT inhibitors currently approved for clinical use are azacitidine (Vidaza) and decitabine (Dacogen) (Table S2), nucleoside analogs that incorporate into DNA during replication, irreversibly bind DNMTs and preventing methylation. This leads to hypomethylation of DNA and reactivation of silenced tumor suppressor genes and other genes involved in differentiation and apoptosis. Temporal analysis shows steady growth in DNMT inhibitor research reflecting continued optimization of these first-generation epigenetic drugs (Figure 7A).

Table 2. Summary of Mechanisms of Action of Known Classes of Epi-Drugs

Epi-drug class	Target	Mechanism	Outcome
HDAC inhibitors ^{448–453}	Histone deacetylases (HDACs)	Inhibit removal of acetyl and other acyl groups (propionyl, butyryl, crotonyl, succinyl) from histone and nonhistone proteins (p53, NF-κB, HSP90, α-tubulin, STAT3)	Chromatin relaxation, reactivation of silenced genes, modulation of protein stability/localization, induction of apoptosis, cell cycle arrest, immune activation
DNMT inhibitors ^{456–458}	DNA methyltransferases (DNMTs)	Inhibit DNA methylation, leading to hypomethylation	Reactivation of silenced tumor suppressor genes
BET inhibitors ^{459–462}	BET proteins (BRD2, BRD3, BRD4)	Block binding to acetylated histones, disrupting transcription	Downregulation of oncogenes (e.g., MYC)
KDM inhibitors ^{463–467}	Histone lysine demethylases (KDMs)	Prevents histone demethylation	Downregulation of oncogenes
HMT inhibitors ^{468–470}	EZH2 (e.g., PRC2 complex)	Inhibit histone methylation (e.g., H3K27me3)	Reactivation of silenced tumor suppressor genes
PRMT inhibitors ^{471,472}	Protein arginine methyltransferases (PRMTs)	Decreases histone methylation	Helps overcome drug resistance (e.g., by regulating the Wnt/β-catenin signaling pathway)
PKMT inhibitors ^{473–475}	Protein lysine methyltransferases (PKMTs)	Decreases histone methylation	Helps overcome drug resistance
EZH2 inhibitors ^{476–478}	EZH2 (PRC2 complex)	Inhibit H3K27 methylation	Reactivation of silenced genes, restoration of differentiation
IDH inhibitors ^{479–482}	Mutant IDH1/IDH2 enzymes	Reduce 2-HG levels, restoring normal epigenetic regulation	Restoration of cellular differentiation, inhibition of tumor growth

- **ncRNA modulators** target miRNAs or lncRNAs to influence gene regulation. RNA modulators appear to be a fast-growing category, especially evident after 2018 (Figure 7A), reflecting recognition of miRNAs and lncRNAs as druggable targets for precision medicine.^{116,318,483,484}
- **Bromodomain and extra-terminal domain (BET) inhibitors** block the binding of BET proteins to acetylated histones, disrupting the transcriptional activation of oncogenes. BET proteins (e.g., BRD2, BRD3, BRD4) are involved in recognition and binding of acetylated lysines on histones and acting as “readers” of epigenetic marks.^{459–462} BET inhibitors (OTX015, CPI-0610, JQ1, I-BET762 etc.; Table S2) remain in clinical trials, with consistent publication growth since 2012 indicating sustained development efforts (Figure 7A).
- **Histone lysine demethylases (KDMs) inhibitors** work by inhibiting enzymes belonging to lysine demethylases (LSDs) or JmjC family N-methyl lysine demethylases (JmjC) family of enzymes.⁴⁶⁷ No approved KDM inhibitors so far, however, a KDM4 inhibitor is currently in clinical trial (zavondemstat, NCT05076552⁴⁸⁵) with ongoing research directed toward developing more KDM4 inhibitors.⁴⁸⁶
- **Histone methyltransferase (HMT) inhibitors** target enzymes that add methyl groups to specific lysine or arginine residues on histone proteins, which can either activate or repress gene expression depending on the specific histone and the location of the methylation. Inhibiting these enzymes can alter gene expression profiles in cancer cells.^{468–470}
- **Protein arginine methyltransferase (PRMT) inhibitors** target protein methyltransferases, enzymes responsible for adding methyl groups to proteins and impacting gene expression and cellular processes.^{487–489} HMT inhibitors (2%) and protein methyltransferase inhibitors such as PRMT and PKMT inhibitors (1% each) remain in early development, while KDM inhibitors (3%) show emerging therapeutic potential (Figure 7A, inset pie chart).
- **Isocitrate dehydrogenase (IDH) inhibitors** target mutant forms of IDH enzymes, such as IDH1 and IDH2, that produce the oncometabolite 2-hydroxyglutamate, causing DNA hypermethylation and block cellular differentiation by accumulating in cells and inhibiting enzymes involved in epigenetic regulation, such as TET proteins and histone demethylases.^{479–482} Mutant IDH enzymes are common in certain cancers such as AML, gliomas, etc.⁴⁹⁰ and inhibition of mutant IDHs can restore normal cellular differentiation.^{491,492} Ivosidenib (Tibsovo) and enasidenib (Idhifa) are examples of IDH inhibitors approved for clinical use (Table S2).
- **Enhancer of zeste homologue 2 (EZH2) inhibitors** function as specific HMT inhibitors, reducing H3K27me3 levels and reactivating silenced tumor suppressor genes which can restore normal cellular differentiation and inhibit tumor growth, particularly in cancers with EZH2 mutations or overexpression. EZH2 is the catalytic subunit of the polycomb repressive complex 2 (PRC2) and mediates the methylation of histone H3 at lysine 27 (H3K27me3), a mark associated with gene silencing.^{476–478} In cancer, EZH2 is often overactive, leading to the silencing of tumor suppressor genes. Approved EZH2 inhibitors are tazemetostat (Tazverik) and

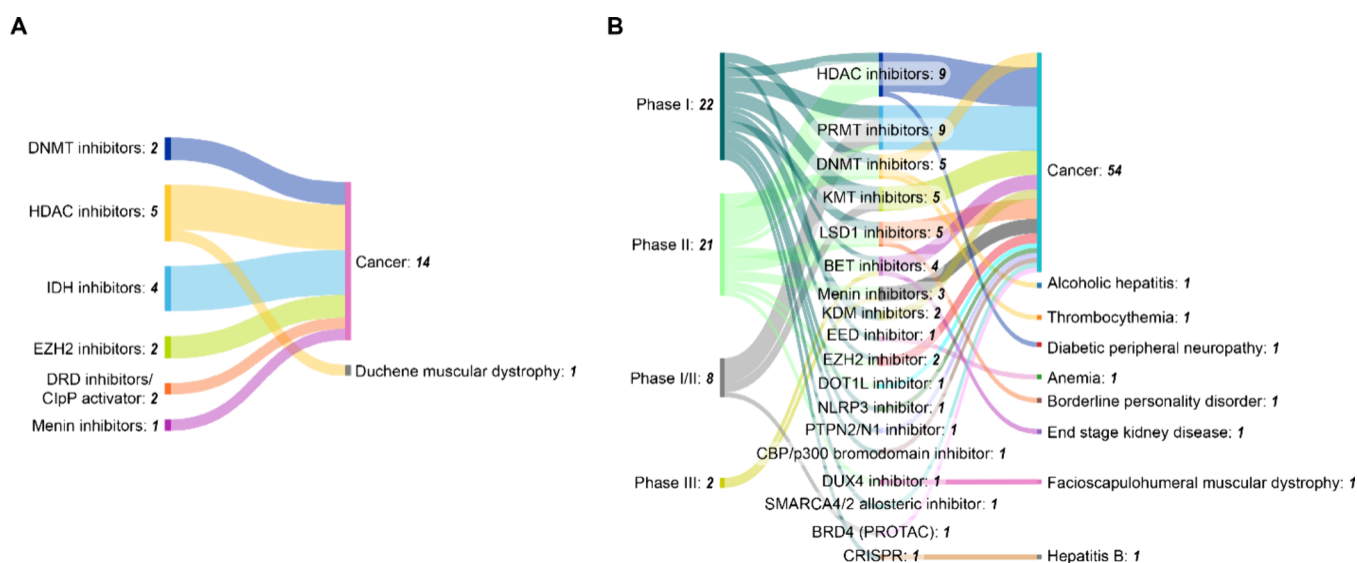


Figure 8. Distribution of (A) FDA approved epigenetic therapeutics and (B) those currently in clinical trials across disease conditions. Does not include combination therapies currently in clinical trials. Each drug/therapy has been counted once in the highest phase trial.

valemetostat tosilate (Ezharmia) (only approved by PMDA in Japan) (Table S2).

- **Dual-action or multipigenetic modulators** combine mechanisms targeting multiple epigenetic pathways. These drugs may enhance therapeutic efficacy in complex diseases.^{493–495}
- Combination strategies integrating epi-drugs with chemotherapy, immunotherapy, and targeted agents show promise for enhanced efficacy.⁴⁹⁶

Co-occurrence analysis of diseases with specific epi-drugs shown as a heat map in Figure 7B, reveals disease-specific research patterns across drug classes. For all of the broad disease categories considered, HDAC inhibitors show the highest co-occurrence reflecting their broad therapeutic potential. Similar patterns were observed for DNMT inhibitors and ncRNA modulators. Cardiovascular and metabolic diseases demonstrate moderate research activity, primarily focusing on HDAC and DNMT inhibitors for reversing pathological cardiac remodeling and metabolic dysfunction, though BET inhibitors had a slightly higher co-occurrence while KDM inhibitors had little to no co-occurrence with cardiovascular diseases.

Key challenges include improving target specificity to reduce off-target toxicity, overcoming resistance mechanisms, identifying predictive biomarkers for patient selection, and enhancing drug delivery to solid tumors.²⁶ The expanding application beyond oncology, particularly in neurological, cardiovascular, and autoimmune diseases, combined with emerging drug classes targeting specific epigenetic mechanisms, positions epigenetic therapeutics at the forefront of precision medicine approaches for complex diseases.

The clinical landscape of epigenetic therapeutics demonstrates regulatory success, with 13 U.S. FDA-approved drugs targeting key epigenetic regulators (Figure 8A). HDAC inhibitors dominate with 6 approvals and hematological malignancies represent the primary therapeutic application. This concentration reflects the particular sensitivity of blood cancers to epigenetic dysregulation and the accessibility of hematologic targets compared to solid tumors.⁴⁹⁷

DNMT inhibitors azacitidine and decitabine were among the first approved epigenetic drugs, establishing proof-of-concept

for targeting DNA methylation in myelodysplastic syndromes and AML. HDAC inhibitors including vorinostat, romidepsin, and belinostat have shown efficacy across multiple hematologic malignancies, validating histone modification as a therapeutic target.

Other Therapeutic Modalities and Strategies in Epigenetic Drug Discovery

Discussed briefly are a few therapeutic approaches beyond the existing/well-established epigenetic modulators.²⁶

- Dual-action or multipigenetic modulators and combination strategies

Combine mechanisms targeting multiple epigenetic pathways. These drugs may enhance therapeutic efficacy in complex diseases.^{493–495} This strategy either utilizes multiple targets from same epigenetic mechanisms (e.g., dual inhibitors targeting histone erasers, readers and writers),⁴⁹⁸ multiple targets from two different epigenetic mechanisms (e.g., CM-272),^{499,500} or a combination of targets from epigenetic and nonepigenetic mechanisms⁴⁹³ (e.g., CUDC-101).⁵⁰¹ A recently published article⁵⁰² details the use of computer-aided drug design to identify small molecule inhibitors of multiple methyltransferase-like protein (METTLs, m⁶A methylation writers).

CM-272, a small molecule inhibitor with a quinolone moiety acts as a fairly potent dual reversible inhibitor of DNMTs (DNMT1, DNMT3A) and euchromatic histone-lysine N-methyltransferase 2 (EHMT2, also referred to as G9a) with possible use in the treatment of AML, acute lymphoblastic leukemia (ALL), and diffuse large B-cell lymphoma (DLBCL)^{499,500} and eventually expanded beyond to hepatocellular carcinoma (HCC)⁵⁰³ and multiple myeloma (MM)⁵⁰⁴ serving as an example of dual-targeting epigenetic modulators utilizing targets from two different epigenetic mechanisms. Other examples include a DNMT/HDAC inhibitor capable of eliciting a viral mimicry response,⁵⁰⁵ defined as “a cellular state in which the reactivation of silenced transposable elements (TEs) leads to the accumulation of immunogenic nucleic acids, triggering innate immune pathways that resemble responses mounted against viral pathogens”⁵⁰⁶ and therefore likely to be useful in combination with immune checkpoint inhibitors.

Examples of dual epigenetic and nonepigenetic targeting agents include CUDC-101, a small molecule inhibitor of HDAC, EGFR, and HER2,⁵⁰¹ and HH-2853, a small molecule inhibitor of EZH1/2, BRD4, PARP1, and EHMT2⁵⁰⁷ among many others.

Integrating epi-drugs with chemotherapy, immunotherapy, and targeted agents show promise for enhanced efficacy.⁴⁹⁶ Some of the most explored combination strategies involve combining U.S. FDA approved DNMT inhibitors azacitidine or decitabine with U.S. FDA approved HDAC inhibitors such as vorinostat and romidepsin or with other investigational HDAC inhibitors such as pracinostat and mocetinostat.⁵⁰⁸ Other combination strategies include designing hybrid molecules containing pharmacophores for more than one epigenetic target.⁵⁰⁹

• CRISPR-based epigenetic therapies

Various CRISPR-based epigenetic therapeutic approaches are being explored including in the treatment of sickle cell anemia,⁵¹⁰ cancer,⁵¹¹ and hepatitis B.⁵¹² Among these, Tune-401 has reached clinical trials with Phase I study initiated in 2024 (NCT06671093²²³).

• PROTACs and protein degraders

The development of PROTACs as a therapeutic modality appears to be occurring at a brisk and continuous pace with several epigenetic PROTACs being pursued. These include PROTACs targeting HDAC6,⁵¹³ bromodomain-containing protein 9 (BRD9)^{514,515} as well as embryonic ectoderm repressive complex 2 (PRC2).⁵¹⁶ Besides these, other epigenetic PROTACs under development include SMARCA2/4 targeting PROTACs (ABCI1⁵¹⁷) as well those targeting other members of the HDAC family such as HDAC3,⁵¹⁸ HDAC4,⁵¹⁹ HDAC6⁵²⁰ and many others. Most epigenetic PROTACs appear to utilize either cereblon targeting ligands such as thalidomide, pomalidomide and their variations or Von-Hippel-Lindau targeting ligands such as VHL1. For instance, compound 17c, a HDAC6 PROTAC utilizes 6-fluorothalidomide to recruit cereblon while the two BRD9 PROTACs CFT8634 and FHD-609 utilize anilino glutarimide and thalidomide, respectively, to also recruit cereblon. The EED PROTAC, ABI1, on the other hand utilized a VHL targeting ligand, VHL1 (Figure S5).

Out of these PROTACs, both BRD9 targeted ones (CFT8634 and FHD-609) reached clinical trials (NCT05355753,⁵²¹ NCT04965753⁵²²). However, both trials were terminated, the former (CFT8634, NCT05355753) due to lack of BRD9 degradation resulting in insufficient efficacy for treating patients with synovial sarcoma⁵²³ while the latter (FHD-609, NCT04965753) appears to a sponsor decision due to safety concerns (related to a grade 4 QTc prolongation event).⁵²⁴ Efforts have also been made toward designing regulated induced proximity targeting chimeras (RIPTACs), a type of heterobifunctional molecule designed for cancer therapy that leads to formation of a ternary complex achieved by targeting two proteins one of which is exclusively expressed in cancer cells while the other is essential for cell survival.^{525,526} RNK-05047 (Ranok Therapeutics (Hangzhou) Co., Ltd.) is a RIPTAC that has entered Phase I/II trials and is actively recruiting individuals (NCT05487170,⁵²⁷ Table S3). While the termination of the two clinical trials is disappointing, the development of PROTACs and other protein degraders is only likely to continue and accelerate.⁵²⁸

• Antisense oligonucleotides (ASOs)

Antisense oligonucleotides (ASOs) are short single strand RNA or DNA fragments usually comprising of 15–21 nucleotides and synthesized to bind to mRNA to achieve change in protein expression by inhibition of or increasing translation, by causing RNA knockdown by causing degradation of RNA.^{529,530} While ASOs have been around for a while, it is only in the past 10 or so years that development of ASOs as therapeutics has increased with multiple ASOs entering clinical trials.^{529–531} In the area of epigenetics, ASOs are being pursued to target lncRNAs such as Chaserr potentially useful in treating neurological disorders such as epilepsy⁵³² or methylation writers such as EZH2⁵³³ in combination with androgen receptors to effectively inhibit growth of castration-resistant prostate cancer cells.⁵³³

• miRNA mimics

miRNA mimicry involves the use of synthetic double-stranded microRNAs that imitate endogenous miRNA⁵³⁴ allowing regulation of gene transcription.⁵³⁵ The first miRNA mimic entered clinical trials in the early 2010s⁵³⁶ and while there has been continued interest in the development of miRNA mimics, persistent challenges remain.⁵³⁷ In the epigenetic landscape, the use of miRNA mimics has been mostly been explored in the treatment of cancer (MRX34, miR-34 mimic, NCT01829971,⁵³⁸ NCT02862145;⁵³⁹ MesomiR-1, miR-16 mimic, NCT02369198;⁵⁴⁰ INT-1B3, miR-193a mimic, NCT04675996⁵⁴¹) and keloid (MRG-201, miR-29 mimic, NCT03601052⁵⁴²). The miR-34 clinical trials had to be terminated or withdrawn due to severe immune related adverse events, while the miR-193a trial was terminated due to insufficient funding. MesomiR-1, a type of TargomiR described as nonliving bacterial minicells packaged with miRNA mimic, showed promising results with reasonable safety profile.^{543,544} In 2022, the development of MRG-229, another miR-29 mimic based on MRG-201, was reported with potential use in treating idiopathic pulmonary fibrosis.⁵⁴⁵ While clinical development of miRNA mimics has a long way to go, they nonetheless remain an important avenue for continued exploration.

■ EPIGENETIC DRUGS IN CLINICAL TRIALS

The epigenetic drug clinical trial landscape has expanded dramatically over 25 years, with nearly 2,200 trials registered on clinicaltrials.gov.⁵⁴⁶ Figure 9 shows sustained increase from a

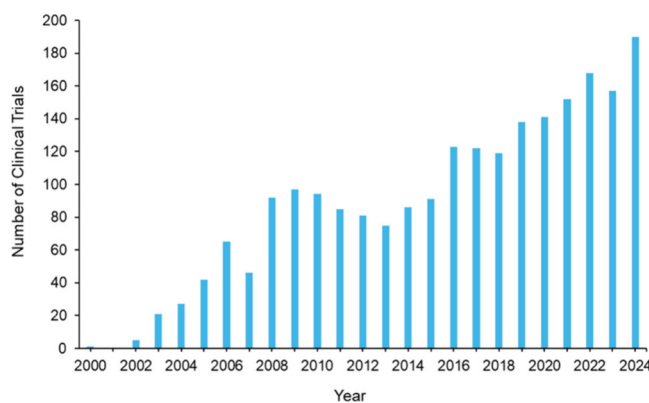


Figure 9. Number of therapeutic epigenetic drug clinical trials by year. Source: CAS, publicly available information from clinicaltrials.gov.⁵⁴⁶ Clinical trials are characterized by first posted date.

single trial in 2000 to just under 200 clinical trials a year in 2024, with notable oscillations reflecting regulatory milestones and market dynamics. Following azacitidine's U.S. FDA approval in 2004, trial activity demonstrated waxing and waning patterns with overall upward trajectory. This growth pattern indicates sustained pharmaceutical investment despite periodic consolidation phases.

Analysis of therapeutic epigenetic clinical trials with respect to phase distribution reveals that Phase II trials dominate at 57%, followed by Phase I (32%) and Phase III (9%) (Figure 10A).

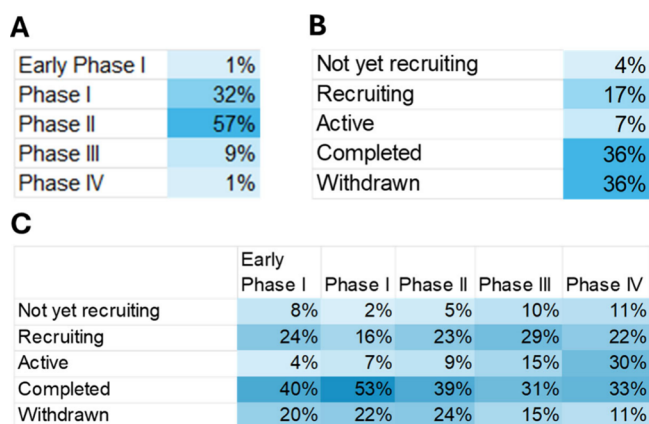


Figure 10. Distribution of epigenetic drug clinical trials in terms of (A) overall clinical trial phase development, (B) overall clinical trial statuses, (C) clinical trial phases compared against statuses. Phase I/II and Phase II/III studies are classified under Phase II and Phase III studies, respectively. Source: CAS, publicly available information from clinicaltrials.gov.⁵⁴⁶

This distribution is typical of drug development and reflects the high attrition rate in epigenetic drug development and the exploratory nature of many combination strategies. Similar analysis in terms of trial status (Figure 10B) shows that 36% of all clinical trials over the past 25 years have been completed, while 29% of trials remain in early development (not yet recruiting, recruiting, and active status). The phase-specific status breakdown (Figure 10C) shows that Phase I trials have the highest percentage of completed trials (53%) while Phase III trials demonstrate highest active recruitment (29%). For active trials we see the highest percentages for Phase IV and Phase III trials at 30% and 15%, respectively.

Notably, 62% of trials involve U.S. FDA-approved drugs (Supplementary Figure S6), suggesting extensive label expansion efforts and combination therapy exploration beyond initial indications. The remaining 38% of clinical trials are for newer/novel unapproved drugs. Exemplary nonregulatory approved epigenetic drugs active in the clinical trial pipeline are summarized in Supplementary Table S3.

The clinical development pipeline reveals robust activity with more than 50 ongoing trials across three phases. Phase I and II trials dominate (22 and 21 trials) with DNMT and HDAC inhibitors maintaining a strong clinical presence with 5 and 9 candidates in trials, respectively. However, the clinical landscape shows diversification beyond DNMT and HDAC inhibitors, with LSD1, BET and menin inhibitors being increasingly explored. Additionally, a BRD4 PROTAC and CRISPR based epigenetic therapy are also currently in Phase I (Figure 8B and Table S3). This distribution demonstrates both continued

optimization of established targets and expansion into novel epigenetic regulators.

Cancer applications continue to predominate with more than 50 trials, though data suggests encouraging diversification into nononcologic conditions including thrombocythemia, alcoholic hepatitis, anemia, and diabetic peripheral neuropathy. This expansion reflects growing recognition of epigenetic contributions to diverse pathologies beyond cancer.

Promising Epigenetic Agents in Clinical Development

Oncology Agents. Pelabresib (CPI-0610), an advanced BET inhibitor, reached Phase III trials for myelofibrosis in the MANIFEST-2 study (NCT04603495⁵⁴⁷). The U.S. FDA gave Fast Track Designation to pelabresib for myelofibrosis in December 2019.⁵⁴⁸ The progression to Phase III indicates strong Phase II efficacy data with a 35% spleen volume reduction rate when combined with ruxolitinib.⁵⁴⁹ Its mechanism of action involves disrupting the interaction between BET proteins and acetylated chromatin, leading to downregulation of oncogenic transcription.⁵⁵⁰

Ziftomenib (KO-539), a selective small-molecule inhibitor of the menin-KMT2A protein–protein interaction,⁵⁵¹ received U.S. FDA Breakthrough Therapy designation in March 2024 for relapsed or refractory NPM1 mutant AML.⁵⁵² In the ongoing KOMET-001 Phase I/II trial (NCT04067336⁵⁵³), ziftomenib has demonstrated meaningful clinical activity in both NPM1 mutant AML (30% of cases) and KMT2A-rearranged AML (10% of adult cases).⁵⁵⁴ This targeted therapy works by disrupting menin-KMT2A interactions to downregulate specific oncogenes and induce myeloid differentiation.⁵⁵⁵

Nononcology Agents. *Cardiovascular and Metabolic Indications.* Apabetalone (RVX-208), a selective BET inhibitor targeting BD2 domains, is currently in Phase I/II for end-stage kidney disease (NCT03160430⁵⁵⁶). The U.S. FDA previously granted Breakthrough Therapy Designation in 2020 for major adverse cardiovascular events (MACE) reduction in high-risk type 2 diabetes patients with coronary artery disease.⁵⁵⁷ Unlike oncology-focused BET inhibitors, apabetalone modulates cardiovascular and renal disease pathways through selective inhibition of BRD4.⁵⁵⁸ This represents a new approach to treating both kidney and cardiovascular diseases through epigenetic modulation, potentially establishing BET inhibition as a new therapeutic approach for chronic metabolic and inflammatory conditions.

Larsucosterol (DUR-928), an endogenous sulfated oxysterol and first-in-class epigenetic regulator, is in Phase II trials for alcoholic hepatitis (NCT04563026⁵⁵⁹). The U.S. FDA granted Breakthrough Therapy Designation for alcoholic hepatitis in 2024.⁵⁶⁰ Larsucosterol functions via DNMT inhibition and modulating lipid metabolism, inflammatory responses, and cell survival pathways.⁵⁶¹ Phase II results showed a 74% survival rate at day 28 compared to 53% for standard of care in severe alcoholic hepatitis patients.⁵⁶² Larsucosterol was also explored in a Phase 1b clinical study in nonalcoholic steatohepatitis (NASH) patients where it showed a good safety profile and improvement in insulin resistance, liver stiffness, liver enzyme, and biomarkers for liver health.⁵⁶¹

Neuropsychiatric Applications. Vafidemstat (ORY-2001) is a selective LSD1 inhibitor that is currently being evaluated in Phase II clinical trials for borderline personality disorder (NCT04932291⁵⁶³). This represents a novel therapeutic approach to treating psychiatric conditions through epigenetic modulation, specifically targeting lysine-specific

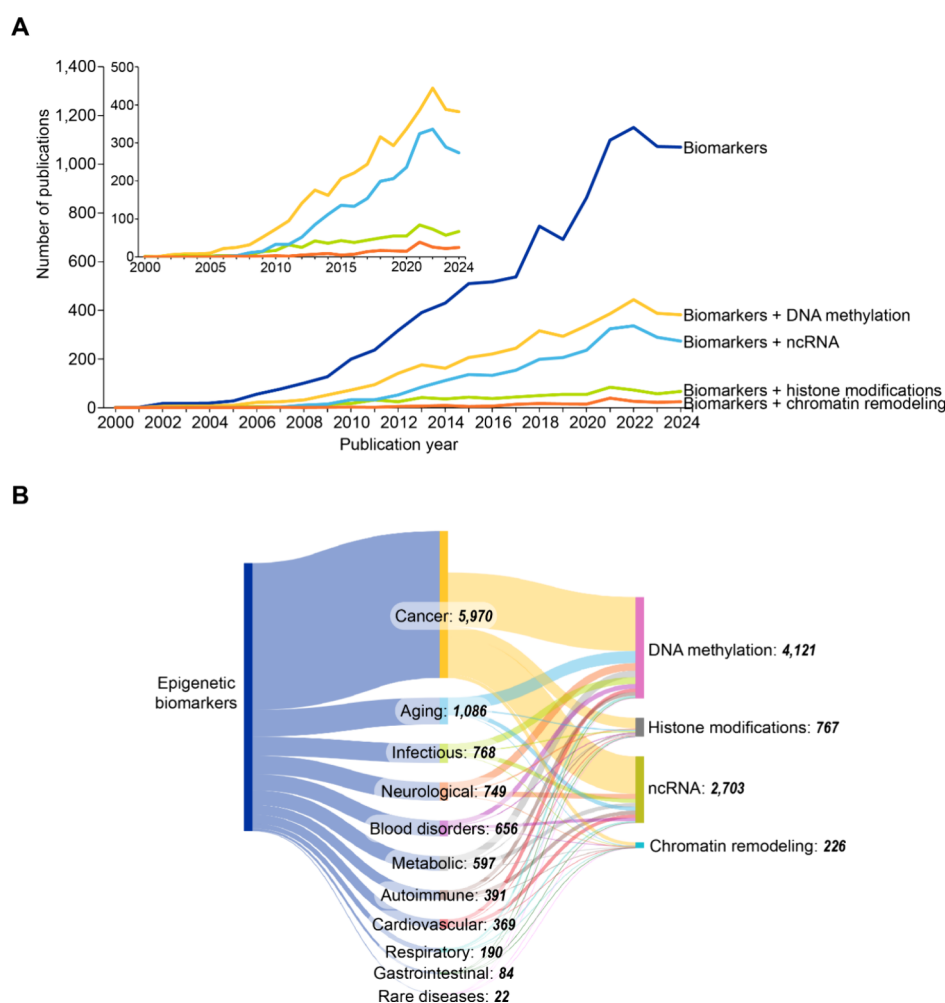


Figure 11. (A) Publication trends for publications related to biomarkers and specific epigenetic mechanisms and (B) their co-occurrences with specific disease types and epigenetic mechanism. Data includes journal articles and patents extracted from the CAS Content Collection for the period 2000–2024.

demethylase1 (LSD1) to potentially address the underlying neurobiological mechanisms of personality disorders.⁵⁶⁴ The development of vafidemstat for borderline personality disorder highlights the expanding application of epigenetic therapies beyond oncology into complex psychiatric and neurological conditions.

Recently Approved Agents: Validating Novel Mechanisms. First-in-Class Approvals. Modeyso (Dordaviprone) received U.S. FDA accelerated approval in August 2025 for H3K27 M mutant diffuse midline gliomas in patients over the age of one.⁵⁶⁵ This first in class drug represents a novel epigenetic approach through dopamine receptor antagonism, leading to integrated stress response activation and epigenetic modulation.⁵⁶⁶ It addresses a critical unmet need in neuro-oncology with a 5-year survival rate of 1%.⁵⁶⁷ Previous/Completed clinical trials have shown that Modeyso had successful tumor uptake with therapeutic intratumoral concentrations.⁵⁶⁸ The ongoing Phase III ACTION study (NCT05580562⁵⁶⁹) continues efficacy and safety evaluation.

Revuforj (Revumenib) gained U.S. FDA approval in November 2024 for relapsed/refractory KMT2A-rearranged AML in adults and pediatric patients.⁵⁷⁰ The AUGMENT-101 trial (NCT04065399⁵⁷¹) demonstrated a 63% overall response rate in KMT2Ar acute leukemia,⁵⁷² representing first U.S. FDA-approved menin inhibitor for cancer treatment.⁵⁷³ Ongoing

trials for AML (NCT06652438⁵⁷⁴) and colorectal carcinoma (NCT05731947⁵⁷⁵) explore broader applicability of menin inhibition.

Future Perspectives. The clinical pipeline demonstrates maturation of epigenetic therapeutics beyond first-generation DNMT and HDAC inhibitors toward precision-targeted agents. The expansion into nononcology indications such as cardiovascular, metabolic, and neuropsychiatric diseases, validates epigenetic modulation as a broadly applicable therapeutic strategy. With multiple Phase III programs and recent approvals validating novel mechanisms, the field approaches an inflection point where epigenetic drugs may transform treatment paradigms across diverse pathologies.

■ EPIGENETIC BIOMARKERS: CLINICAL APPLICATIONS AND FUTURE PERSPECTIVES

Epigenetic biomarkers are molecular signatures derived from epigenetic modifications that reflect changes in gene expression patterns without altering the DNA sequence. These biomarkers are increasingly recognized for their potential in disease diagnosis, prognosis, and therapeutic decision-making. Their reversible nature and responsiveness to environmental factors make them particularly valuable for understanding complex diseases such as cancer,⁵⁷⁶ neurological disorders,⁵⁷⁷ and

cardiovascular diseases.³⁹⁸ Temporal analysis of publication indicate a sharp increase starting around 2008–2009 and marked by a few periods of plateauing (2015–2017 and 2023–2024) (Figure 11A). Among the four main epigenetic mechanisms, DNA methylation and ncRNA appear to be the most well-studied consistently, while histone modifications and chromatin remodeling remain comparatively underexplored in this context. (Figure 11A, inset). The sankey diagram (Figure 11B) illustrates the distribution of epigenetic biomarker research across disease categories and mechanism types. Cancer appears to co-occur most extensively with publications related to epigenetic biomarkers by a long margin, reflecting the well-established role of epigenetic dysregulation in oncogenesis and the clinical utility of methylation markers such as MGMT⁵⁷⁸ and VIM⁵⁷⁹ for diagnosis and prognosis. The substantial representation of aging research aligns with recent advances in epigenetic clock development and their applications in biological age assessment.

Epigenetic biomarkers offer distinct advantages:

- Dynamic responsiveness to environmental factors enables real-time disease monitoring;
- Tissue specificity provides organ-specific disease insights;
- Early detection capability precedes clinical manifestation allowing early intervention;
- Noninvasive sampling through blood, urine, and saliva facilitates routine monitoring and improving patient compliance.

Epigenetic biomarkers are broadly categorized based on the type of epigenetic modification they represent.

DNA Methylation Biomarkers

Aberrant DNA methylation patterns serve as disease hallmarks, with hypermethylation of tumor suppressors (e.g., BRCA1,⁵⁸⁰ p16³¹³) being a common biomarker of various cancers, while altered BDNF methylation has been linked to depression and schizophrenia.⁵⁸¹

SEPT9 methylation enables noninvasive colorectal cancer screening through blood-based detection (Epi proColon⁵⁸²), offering an alternative to colonoscopy. **MGMT methylation** predicts Temozolomide response in glioblastoma, with hypermethylation being associated with better outcomes.⁵⁸³ **BRCA1 methylation** serves as diagnostic and prognostic marker associated with increased risk of breast⁵⁸⁴ and ovarian cancer.⁵⁸⁵ Notably, DNA methylation co-occurs most predominantly with epigenetic biomarker research, likely due to its chemical stability, established detection methodologies, and direct clinical translation through U.S. FDA-approved tests.

Histone Modification Biomarkers

Histone modifications influence chromatin structure and gene expression and specific histone marks are associated with disease states. The relatively modest co-occurrence of histone modifications (Figure 11B) suggests untapped potential, particularly given their dynamic nature and responsiveness to therapeutic interventions. However, challenges remain such as the highly labile nature of histone modifications and their susceptibility to enzymatic degradation causing rapid signal loss within minutes of sample collection. Current protocols require specialized preservation buffers containing protease, deacetylase, and demethylase inhibitors, complicating routine clinical implementation.

Loss of histone H4 acetylation predicts poor prognosis in certain cancers,⁵⁸⁶ whereas **increased acetylation at lysine 12**

(H4K12) is a novel biomarker for Alzheimer's.⁵⁸⁷ Beyond acylation, other post-translational modifications on histones offers additional biomarker potential. H3 lysine 27 trimethylation (**H3K27me3**) reflects PRC2 dysregulation and correlates with poor prognosis in prostate⁵⁸⁸ and bladder cancers.⁵⁸⁹ **Histone H3/H4 citrullination** is linked to the production of anticitrullinated protein antibodies (ACPAs), a hallmark of rheumatoid arthritis enabling rheumatoid arthritis diagnosis and monitoring.^{590,591} **Crotonylation of H2BK12 (H2BK12cr)** in peripheral blood mononuclear cells (PBMCs) is significantly elevated in colorectal cancer patients,⁵⁹² while serum **histone succinylation** shows strong correlations with tumor type, stage, and prognosis, enabling rapid pan-cancer screening through a single-tube blood test.⁵⁹³ Additionally, increased histone **lactylation at H4K5 (H4K5lac)** in PBMCs and breast cancer tissues correlates with serum lactate and CEA levels, suggesting its potential role in diagnosis and therapeutic strategies.⁵⁹⁴

ncRNA Biomarkers

ncRNAs, including miRNAs like miR-21⁵⁹⁵ and miR-155⁵⁹⁶ are overexpressed in various cancers and serve as diagnostic and prognostic biomarkers, often provide accessible disease markers through bodily fluids.⁵⁹⁷ The substantial ncRNA co-occurrence with epigenetic biomarker research reflects growing interest in circulating miRNAs and lncRNAs as minimally invasive biomarkers (Figure 11B).

miR-21 overexpression is associated with tumor growth, invasion, metastasis, and chemotherapy resistance across multiple cancers including breast, lung, and colorectal cancer. **Elevated miR-208a levels** serves as a biomarker for acute myocardial infarction (heart attack).⁵⁹⁸ **HOTAIR lncRNA overexpression** is associated with metastasis and poor prognosis in breast,⁵⁹⁹ colorectal,⁶⁰⁰ and pancreatic cancers.⁶⁰¹

Chromatin Remodeling Biomarkers

Chromatin remodeling patterns predict therapeutic resistance, with open chromatin regions in drug-resistant cancer cells being used to predict treatment failure.⁶⁰² Similar to histone modifications, the relatively low co-occurrence of chromatin remodeling suggests untapped potential.

The future of epigenetic biomarkers lies in the development of robust, high-throughput technologies and integrative approaches. Multiomic integration combining epigenetic, genomic, transcriptomic, and proteomic data will enhance biomarker discovery and validation. Noninvasive detection of epigenetic biomarkers in bodily fluids will revolutionize early diagnosis and monitoring. Advanced computational tools are expected to aid in identifying complex epigenetic signatures for patient stratification. CRISPR-based epigenome editing may enable manipulation of disease-associated epigenetic marks for therapeutic purposes, potentially transitioning biomarkers from diagnostic to therapeutic tools.

While challenges remain including standardization and validation, technological advances and clinical implementation strategies are establishing epigenetic biomarkers as cornerstone components of precision medicine, offering unprecedented opportunities for early detection, prognostic assessment, and therapeutic guidance across diverse pathologies.

Emerging Technologies in Epigenetic Research

Technological advances are revolutionizing epigenetic research, enabling precise, scalable investigations into epigenetic modification mechanisms and therapeutic applications. These innovations drive discoveries in gene regulation, disease

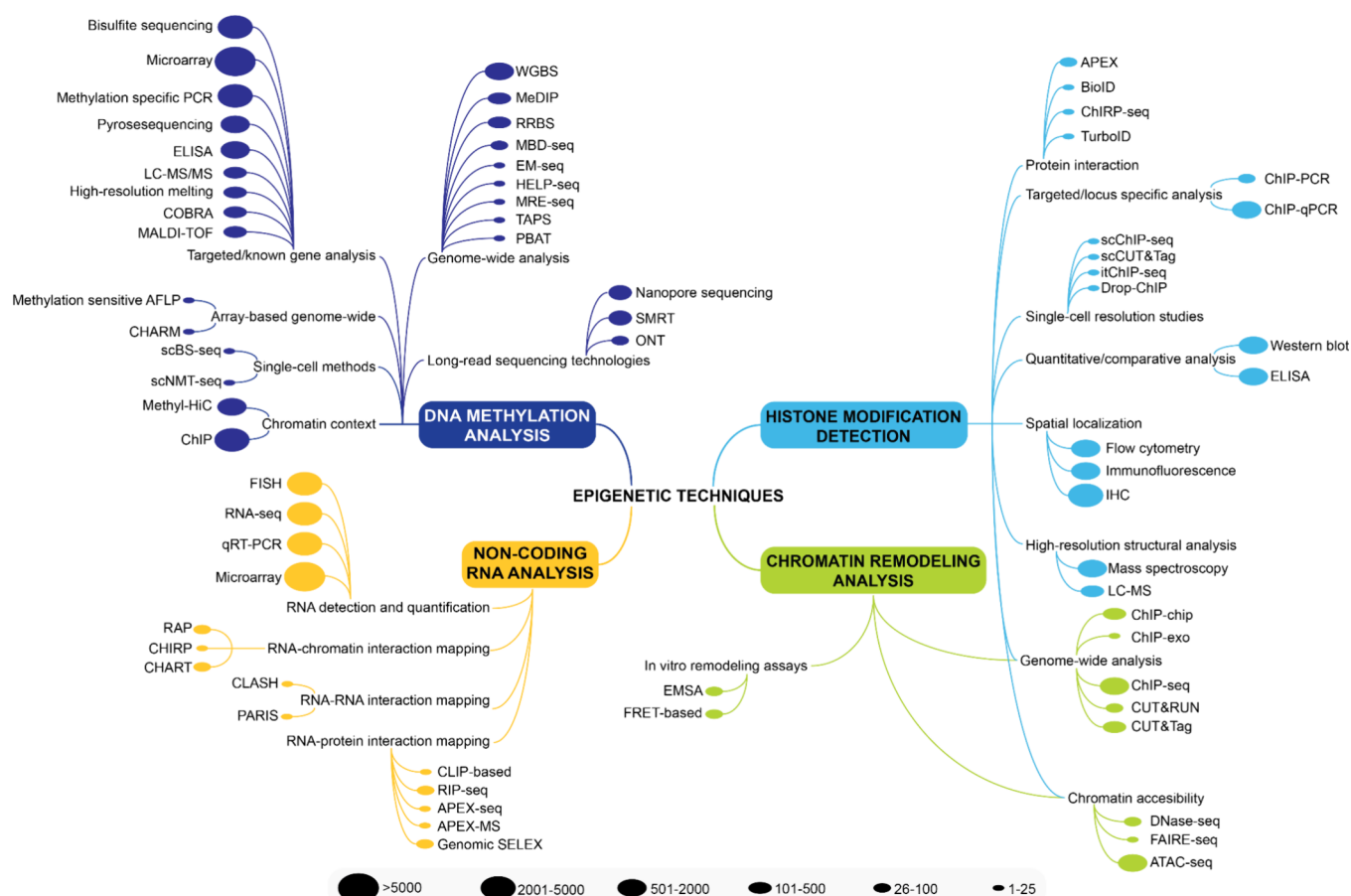


Figure 12. Comprehensive overview of epigenetic publications organized by key epigenetic analytical techniques. The size of the ellipse corresponds to number of publications (journal and patents) extracted from the CAS Content Collection for the period 2000–2024.

pathogenesis, and drug development across multiple research domains. In Table S4 we have summarized some of the most exciting and transformative technologies in epigenetic research, highlighting single-cell resolution methods, multiomics integration, and precision editing tools that complement established techniques shown in Figure 12.

The current epigenetic research landscape appears to be focused on four major analytical domains (Figure 12): DNA methylation analysis, histone modification detection, ncRNA analysis, and chromatin remodeling analysis. Our analysis indicates that DNA methylation analysis methods are led by whole-genome bisulfite sequencing (WGBS) and targeted approaches like methylated immunoprecipitation (MeDIP)⁶⁰³ and reduced representation bisulfite sequencing (RRBS),⁶⁰⁴ reflecting the maturity of methylation research. Histone modification detection shows strong representation of ChIP-seq variants, with emerging single-cell methods (scChIP-seq, scCUT&Tag) indicating technological evolution toward higher resolution analysis, revealing cell-to-cell variability in cancer, immune responses, and development.^{147,166,168,225,605–607} Single-cell epigenomics addresses limitations of bulk methods that average signals across heterogeneous cell populations.

Multiomics integration combines epigenomic data with transcriptomics, proteomics, and metabolomics using technologies like CUT&RUN⁶⁰⁸ and Hi-C integration,⁶⁰⁹ providing holistic cellular function views.^{610,611} These approaches enable comprehensive pathway analysis and biomarker discovery in complex diseases.

Epigenome editing tools, particularly CRISPR/dCas9-based systems fused with epigenetic effectors (DNMTs, TETs, HDACs), enable targeted modification investigation without DNA sequence alteration.^{612,613} Alternative platforms include TALE⁶¹⁴ and zinc-finger proteins for locus-specific targeting.^{615,616}

Advanced imaging techniques including super-resolution microscopy (STORM,⁶¹⁷ PALM⁶¹⁸) and live-cell fluorescent probes enable real-time chromatin dynamics observation,^{619,620} complementing sequencing-based approaches¹⁵⁹ shown in Figure 11. Long-read sequencing technologies (PacBio, Oxford Nanopore) provide direct modification detection without chemical conversion, addressing limitations in repetitive genomic regions.^{621,622}

The technological evolution from traditional methods to emerging approaches demonstrates the field's progression toward higher resolution, greater precision, and enhanced clinical relevance, positioning epigenetic research for significant therapeutic breakthroughs.

■ ETHICAL, LEGAL, AND SOCIAL IMPLICATIONS

Epigenetics has the potential to transform our understanding of health and disease, but it also raises significant ethical, legal, and social challenges.^{623–628} Addressing these challenges requires a proactive and collaborative approach that balances the benefits of epigenetic research with the need to protect individual rights and promote social justice. By developing robust ethical guidelines, legal protections, and public engagement strategies,

we can harness the power of epigenetics to improve health and well-being while minimizing potential harms.^{623–628}

Ethical Considerations

Privacy and Discrimination. Epigenetic information, like genetic data, is highly personal and can reveal sensitive information about an individual's health, lifestyle choices, and environmental exposures requiring robust privacy protections. For instance, epigenetic markers predicting disease risk could enable discrimination by health insurers through premium increases or coverage denial. Similarly, the ability to detect epigenetic modifications from past behaviors raises concerns about retroactive health assessments and employment discrimination.

Individual Responsibility and Transgenerational Impact. Epigenetic research highlights how lifestyle choices (e.g., smoking, diet) can influence gene expression. This could lead to blaming individuals for their health conditions, ignoring broader social and environmental determinants of health. Epigenetic changes induced by environmental factors can sometimes be passed to future generations. This raises ethical concerns about the long-term consequences of interventions that alter epigenetic markers.

Legal Framework Requirements

Regulatory Oversight. The proliferation of direct-to-consumer epigenetic testing kits raises concerns about accuracy, interpretation, and regulation. Epigenetic biomarkers are increasingly used in diagnostics and personalized medicine. Legal frameworks must ensure that these applications are evidence-based and ethically sound while preventing premature commercialization of unproven technologies.

Liability and Intellectual Property. Epigenetic modifications linked to environmental exposure raise complex liability questions regarding responsibility, for instance should toxin-induced harm be the responsibility of employers, manufacturers, or governments? Potential legal accountability for epigenetic harm to fetuses through parental or third-party negligence or environmental exposures would require careful consideration. The commercialization of epigenetic research, such as biomarkers or therapies, raises questions about patenting and ownership, with balancing innovation with public access to epigenetic technologies being a key legal challenge.

Social Implications

Public Understanding and Health Equity. Limited public understanding of epigenetics enables misconceptions and exploitation through unproven therapies including overhyped claims marketed as "epigenetic diets". Comprehensive education and robust scientific communication are essential to address this issue and to improve informed decision-making.

Epigenetic research linking environmental exposures to disease may inadvertently contribute to stigmatization of certain communities or populations, particularly those exposed to high levels of pollution or toxins. Ensuring equitable access to epigenetic therapies and preventing exacerbation of health disparities requires deliberate policy interventions addressing socioeconomic barriers.

Implementation Strategies. Effective governance requires clear ethical guidelines for epigenetic data collection, use, and sharing; updated legal frameworks addressing unique challenges of epigenetic information; public engagement fostering trust and informed consent; and international cooperation ensuring consistent standards across jurisdictions. By developing robust

protections while promoting responsible innovation, society can harness epigenetics' transformative potential while safeguarding individual rights and promoting health equity.^{623–628}

FUTURE DIRECTIONS AND CHALLENGES

Epigenetics has transformed our understanding of gene-environment interactions and their roles in disease pathogenesis, including cancer, neurodegenerative diseases, and metabolic disorders and has opened new avenues for research and therapy including exploring their potential in personalized medicine. However, the field remains in relative infancy with fundamental questions unanswered. Future progress depends on technological innovation, clinical translation, and addressing complex ethical and methodological challenges.

Technological Advances Driving Discovery

- Single-cell technologies are revolutionizing the study of epigenetic heterogeneity within tissues and cell populations allowing researchers to map epigenetic landscapes at unprecedented resolution, revealing cell-specific regulatory mechanisms. Spatial epigenomics further preserves tissue architecture, providing context-dependent regulatory insights essential for understanding development and disease progression.
- Advances in next-generation sequencing (NGS) and third-generation sequencing (e.g., nanopore sequencing) are enabling genome-wide profiling of DNA methylation, histone modifications, and chromatin accessibility. These technologies will facilitate the identification of novel epigenetic biomarkers and therapeutic targets.
- CRISPR-Cas9 and related technologies are being adapted for precise editing of epigenetic marks, such as DNA methylation and histone modifications. These tools hold promise for functional studies and therapeutic applications, such as reactivating silenced tumor suppressor genes and modulation of disease-associated regulatory elements.

Clinical Translation and Precision Medicine

- Epigenetic modifications, such as DNA methylation patterns and miRNA profiles, are being developed as biomarkers for early disease detection, prognosis, and treatment response. For example, epigenetic biomarkers are being explored for cancer screening and monitoring.
- Beyond first-generation epigenetic drugs such as DNMT inhibitors and HDAC inhibitors, already in use for certain cancers, next-generation therapies are likely to include targeted epigenome editing, miRNA-based treatments, ASOs, PROTACs and other protein degraders, and combination strategies.
- Epigenetic profiling can help tailor treatments to individual patients by leveraging their unique epigenetic signatures allowing optimization of treatment selection and dosing and improving efficacy while minimizing adverse effects.

Mechanistic Understanding and Functional Validation

- Despite remarkable progress in cataloging epigenetic modifications across diverse biological contexts, significant gaps remain in our mechanistic understanding of how these modifications are established, maintained, and erased. Future research must prioritize the functional

characterization of these complexes and their context-dependent roles in gene regulation.

Epitranscriptomics and ncRNA Regulation

- Recent studies have demonstrated that lncRNAs are extensively modified by m⁶A, with these modifications influencing their stability, localization, and protein interactions and potentially play a role in diseases such as cancer.^{629–631} The role of RNA modifications in miRNA biogenesis and function has revealed additional layers of regulatory complexity⁶³² with modifications in other ncRNAs suggesting that epitranscriptomic regulation extends across the entire spectrum of regulatory RNAs.
- The identification of disease-associated changes in RNA modification patterns may provide new diagnostic biomarkers and therapeutic targets and understanding the crosstalk between DNA modification and RNA modifications will be essential for developing comprehensive models of gene regulation.

Targeting the epitranscriptomic machinery using small molecule inhibitors⁶³³ and incorporating epitranscriptomic modifications to enhance stability, specificity, and efficacy of RNA-based therapeutics are viable strategies being pursued.

Interdisciplinary Integration and Data Management

- Epigenetics intersects with fields such as genetics, bioinformatics, environmental science, and social sciences. Understanding gene-environment interactions and their impact on health and disease requires collaboration across these domains, integrating molecular mechanisms with population-level health outcomes.
- Integrating epigenomic data with transcriptomic, proteomic, and metabolomic data will provide a more comprehensive understanding of gene regulation and cellular function. Machine learning and artificial intelligence (AI) will play a key role in analyzing and interpreting these complex data sets, identifying potential patterns invisible to traditional analyses and predicting therapeutic responses but will likely require careful validation and interpretation.
- The generation of large-scale epigenomic data sets demand robust computational infrastructure and presents challenges in data storage, analysis, interpretation, and sharing. Developing standardized protocols and computational tools including interoperable databases, and cloud-based platforms will be critical for reproducible research and meta-analyses across studies contributing to advancing the field.

Persistent Challenges

Despite its potential, epigenetics faces several challenges that must be overcome. The dynamic and complex interplay between genetic, epigenetic, and environmental factors complicates the identification of causal mechanisms. Understanding context-specific effects and tissue-specific regulation requiring sophisticated experimental designs and analytical approaches to disentangle remains a major challenge. Current technologies for mapping epigenetic modifications are not yet fully comprehensive or cost-effective, limiting accessibility. Single-cell methods suffer from technical noise and dropout, while spatial techniques sacrifice resolution for coverage. Developing more robust, scalable, and cost-effective methods remains a

priority. Variability in experimental protocols, data processing pipelines and analysis methods can lead to inconsistent results. Establishing standardized guidelines and best practices are critical to improve reproducibility and reliability.

CONCLUSIONS

The landscape of epigenetic research has undergone a remarkable transformation over the past decade, evolving from a specialized area of molecular biology into a mainstream biomedical discipline with profound implications for human health and therapeutic intervention. This remarkable expansion has been driven by a complex interplay and convergence of scientific breakthroughs, technological innovations, regulatory evolution and economic incentives that have transformed our understanding of gene regulation and its implications for human health. The potential pleiotropic effects of epigenetic therapies⁶³⁴ with applications across multiple diseases have altered cost-benefit calculations and require novel approaches to value assessment unlike most conventional drugs. Technological innovations have served as critical enablers of epigenetic research growth, with NGS technologies revolutionizing the scale and precision of epigenomic analysis. The development of ChIP-seq and bisulfite sequencing technologies has allowed comprehensive analysis of histone modifications and DNA methylation patterns, respectively. Recent and ongoing cutting-edge innovations such as single-cell epigenomics and spatial epigenomics continue to push the field forward.

The clinical translation of epigenetic research has achieved remarkable success, with 13 U.S. FDA-approved drugs validating epigenetic targets as therapeutically viable. The more than 35 ongoing clinical trials across multiple development phases indicate sustained investment and confidence in epigenetic therapeutics, with encouraging diversification beyond oncology, with trials spanning metabolic, neurological, inflammatory, and rare diseases. The potential for epigenetic age clocks and other aging biomarkers to find applications in clinical practice and consumer health markets has generated additional commercial interest. The surge in interest in CRISPR-based therapies has meant the concomitant development of epigenome editing technologies, including dCas9-based systems and base editors, all of which are poised to open new possibilities for therapeutic intervention and research applications.

The continued emergence of environmental epigenetic research reflects growing recognition that environmental factors, such as diet, stress, toxins, and lifestyle choices can induce heritable epigenetic changes with transgenerational implications. The integration of environmental exposure data with individual epigenetic profiles enables personalized risk assessment and intervention strategies, representing a fundamental shift toward preventive, precision medicine approaches.

The future of epigenetics is bright, with immense potential for advancing our understanding of biology and improving human health. However, realizing this potential will require overcoming significant challenges, including technological limitations, ethical concerns, and the complexity of epigenetic regulation. By fostering interdisciplinary collaboration, investing in innovative technologies, and addressing ethical and social implications, the field of epigenetics can continue to make groundbreaking discoveries and translate them into meaningful clinical and societal benefits.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acsptscl.5c00729>.

Detailed description of methodology, figures resulting from data analysis of the CAS Content Collection data (Figures S1–S4), structures of epigenetic PROTACs (Figure S5), and clinical trial data (Figure S6), table listing notable patents related to epigenetics (Table S1), table listing approved epigenetic drugs (Table S2), table listing information related to epigenetic drugs currently in clinical trials (Table S3), table summarizing emerging technologies in epigenetics (Table S4) (PDF)

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Notes

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■ ABBREVIATIONS

ABCA1	ATP binding cassette subfamily A member 1
ACE	angiotensin I converting enzyme
ACPA	anticitrullinated protein antibody
AHRR	aryl hydrocarbon receptor repressor
AI	artificial intelligence
ALKBH5	alkB homologue 5, RNA demethylase
ALL	acute lymphoblastic leukemia
AML	acute myeloid leukemia

APC	APC regulator of Wnt signaling pathway
ARID1A	AT-rich interaction domain 1A
ATAC-seq	assay for transposase-accessible chromatin using sequencing
BDNF	brain derived neurotrophic factor
BET	Bromodomain and extra-terminal domain
BPA	bisphenol A
BRCA1	BRCA1 DNA repair associated
CDKN2A	cyclin dependent kinase inhibitor 2A
CHD	chromodomain helicase DNA
ChIP-seq	chromatin immunoprecipitation sequencing
CLL	chronic lymphocytic leukemia
COMT	catechol-O-methyltransferase
CRISPR	clustered regularly interspaced short palindromic repeats
CUT&RUN	cleavage under targets and release using nuclease
dCas9	direct catalytically inactive Cas9
DLBCL	diffuse large B-cell lymphoma
DNMT	DNA methyltransferases
eNOS	nitric oxide synthase 3
F2RL3	F2R like thrombin or trypsin receptor 3
FDA	Food and Drug Administration
FMR1	fragile X messenger ribonucleoprotein 1
FTO	fat mass- and obesity-associated protein
gRNA	guide RNA
H3K27me3	histone H3 lysine 27 trimethylation
HAT	histone acetyltransferases
HCC	hepatocellular carcinoma
HDAC	histone deacetylases
HDM	histone demethylases
HMT	histone methyltransferases
HOTAIR	HOX transcript antisense intergenic RNA
IBD	inflammatory bowel disease
IDH	isocitrate dehydrogenase
ISWI	imitation switch
KCNQ1	potassium voltage-gated channel subfamily Q member 1
KDM	histone lysine demethylase
KRAS	c- <i>K_r</i> -Ras
lncRNA	long noncoding RNA
m ⁶ A	N ⁶ -methyladenosine
MACE	major adverse cardiovascular event
MECP2	methyl-CpG binding protein 2
MeDIP	methylated immunoprecipitation
MeRIP-seq	m ⁶ A specific RNA immunoprecipitation sequencing
MGMT	O ⁶ -methylguanine-DNA methyltransferase
miCLI	m ⁶ A individual-nucleotide resolution cross-linking and immunoprecipitation
miRNA	microRNA
MM	multiple myeloma
MS	multiple sclerosis
NAFLD	nonalcoholic fatty liver disease
NASH	nonalcoholic steatohepatitis
ncRNA	noncoding RNA
NGS	next-generation sequencing
NMPA	National Medical Products Administration
PALM	photoactivated localization microscopy
PBMC	peripheral blood mononuclear cell
PINK1	PTEN-induced kinase 1
piRNA	PIWI-interacting RNA
PKMT	protein lysine methyltransferase
PMDA	Pharmaceuticals and Medical Devices Agency

PPARGC1A	PPARG coactivator 1 alpha
PPAR γ	peroxisome proliferator activated receptor γ
PRC2	polycomb repressive complex 2
PRMT	protein arginine methyltransferase
PROTAC	proteolysis targeting chimera
RA	rheumatoid arthritis
RNA	ribonucleic acid
RRBS	reduced representation bisulfite sequencing
SCN5A	sodium voltage-gated channel alpha subunit 5
SEPT9	septin-9
SHANK3	SH3 and multiple ankyrin repeat domains 3
siRNA	small interfering RNA
SLE	systemic lupus erythematosus
snoRNA	small nucleolar RNA
snRNA	small nuclear RNA
SREBF1	sterol regulatory element binding transcription factor 1
STORM	stochastic optical reconstruction microscopy
SWI/SNF	switch/sucrose nonfermentable
T1D	type 1 diabetes
T2D	type 2 diabetes
TALE	transcription activator-like effector
TET	ten-11 translocation
TXNIP	thioredoxin interacting protein
USD	United States Dollar
UTR	untranslated regions
VIM	vimentin
WGBS	whole-genome bisulfite sequencing
XIST	X inactive specific transcript

REFERENCES

- (1) Anjaria, P.; Asediya, V.; Nayak, J.; Koringa, P. The Epigenetic Landscape: How Environmental Cues Shape Gene Expression. *Epigenomics* **2023**, *15* (5), 267–270.
- (2) Nicoglou, A.; Merlin, F. Epigenetics: A Way to Bridge the Gap Between Biological Fields. *Studies in History and Philosophy of Science Part C-Studies in History and Philosophy of Biological and Biomedical Sciences* **2017**, *66*, 73–82.
- (3) Conway, S. J.; Woster, P. M.; Shen, J. K.; Georg, G.; Wang, S. Epigenetics: Novel Therapeutics Targeting Epigenetics. *J. Med. Chem.* **2015**, *58* (2), 523–824.
- (4) Simmons, D. Epigenetic Influences and Disease. *Nature Education* **2008**, *1* (1), 6.
- (5) Deichmann, U. Epigenetics: The Origins and Evolution of a Fashionable Topic. *Dev. Biol.* **2016**, *416* (1), 249–254.
- (6) Gibney, E. R.; Nolan, C. M. Epigenetics and Gene Expression. *Heredity* **2010**, *105* (1), 4–13.
- (7) Phillips, T. The Role of Methylation in Gene Expression. *Nature Education* **2008**, *1*, 116.
- (8) Moore, L. D.; Le, T.; Fan, G. DNA Methylation and Its Basic Function. *Neuropsychopharmacology* **2013**, *38* (1), 23–38.
- (9) Kaikkonen, M. U.; Lam, M. T. Y.; Glass, C. K. Non-coding RNAs as Regulators of Gene Expression and Epigenetics. *Cardiovasc. Res.* **2011**, *90* (3), 430–440.
- (10) Pauli, A.; Rinn, J. L.; Schier, A. F. Non-coding RNAs as Regulators of Embryogenesis. *Nat. Rev. Genet.* **2011**, *12* (2), 136–149.
- (11) Wu, Y.-L.; Lin, Z.-J.; Li, C.-C.; Lin, X.; Shan, S.-K.; Guo, B.; Zheng, M.-H.; Li, F.; Yuan, L.-Q.; Li, Z.-h. Epigenetic Regulation in Metabolic Diseases: Mechanisms and Advances in Clinical Study. *Signal Transduction and Targeted Therapy* **2023**, *8* (1), 98.
- (12) Fessele, K. L.; Wright, F. Primer in Genetics and Genomics, Article 6: Basics of Epigenetic Control. *Biological Research for Nursing* **2018**, *20* (1), 103–110.
- (13) Farhana, A.; Yusuf, N.; Rasheed, Z. Editorial: Cancer Genetics and Epigenetics: Theranostic Targets and Mechanisms. *Frontiers in Genetics* **2024**, *15*. DOI: 10.3389/fgene.2024.1446474.
- (14) Skinner, M. K. Role of Epigenetics in Developmental Biology and Transgenerational Inheritance. *Birth Defects Research Part C: Embryo Today* **2011**, *93* (1), 51–55.
- (15) Wilkinson, A. L.; Zorzan, I.; Rugg-Gunn, P. J. Epigenetic Regulation of Early Human Embryo Development. *Cell Stem Cell* **2023**, *30* (12), 1569–1584.
- (16) Yu, X.; Zhao, H.; Wang, R.; Chen, Y.; Ouyang, X.; Li, W.; Sun, Y.; Peng, A. Cancer Epigenetics: From Laboratory Studies and Clinical Trials to Precision Medicine. *Cell Death Discovery* **2024**, *10* (1), 28.
- (17) Ling, C.; Rönn, T. Epigenetics in Human Obesity and Type 2 Diabetes. *Cell Metabolism* **2019**, *29* (5), 1028–1044.
- (18) Hwang, J. Y.; Aromolaran, K. A.; Zukin, R. S. The Emerging Field of Epigenetics in Neurodegeneration and Neuroprotection. *Nat. Rev. Neurosci.* **2017**, *18* (6), 347–361.
- (19) van Zundert, B.; Montecino, M. Epigenetics in Neurodegenerative Diseases. *Subcellular Biochemistry* **2025**, *108*, 73–109.
- (20) Mazzone, R.; Zwergel, C.; Artico, M.; Taurone, S.; Ralli, M.; Greco, A.; Mai, A. The Emerging Role of Epigenetics in Human Autoimmune Disorders. *Clinical Epigenetics* **2019**, *11* (1), 34.
- (21) Surace, A. E. A.; Hedrich, C. M. The Role of Epigenetics in Autoimmune/Inflammatory Disease. *Frontiers in Immunology* **2019**, *10*, 1525.
- (22) Mu, S.; Wang, W.; Liu, Q.; Ke, N.; Li, H.; Sun, F.; Zhang, J.; Zhu, Z. Autoimmune Disease: A View of Epigenetics and Therapeutic Targeting. *Frontiers in Immunology* **2024**, *15*, 1482728.
- (23) Caporali, S.; Russo, S.; Leist, M.; Wirtz, P. H.; Amelio, I. Interplay Between Genes and Social Environment: From Epigenetics to Precision Medicine. *Cell Death Discovery* **2025**, *11* (1), 293.
- (24) Heard, E.; Martienssen, R. A. Transgenerational Epigenetic Inheritance: Myths and Mechanisms. *Cell* **2014**, *157* (1), 95–109.
- (25) Fitz-James, M. H.; Cavalli, G. Molecular Mechanisms of Transgenerational Epigenetic Inheritance. *Nat. Rev. Genet.* **2022**, *23* (6), 325–341.
- (26) Dai, W.; Qiao, X.; Fang, Y.; Guo, R.; Bai, P.; Liu, S.; Li, T.; Jiang, Y.; Wei, S.; Na, Z.; Xiao, X.; Li, D. Epigenetics-Targeted Drugs: Current Paradigms and Future Challenges. *Signal Transduction and Targeted Therapy* **2024**, *9* (1), 332.
- (27) Epigenetics Market Size, Share, and Trends 2025 to 2034. 2025. <https://www.precedenceresearch.com/epigenetics-market>.
- (28) Ganesan, A.; Arimondo, P. B.; Rots, M. G.; Jeronimo, C.; Berdasco, M. The Timeline of Epigenetic Drug Discovery: From Reality to Dreams. *Clinical Epigenetics* **2019**, *11* (1), 174.
- (29) Garde, D. Merck Bets up to \$515M on an Epigenetic Project for Cancer and Blood Disease. 2016. <https://www.fiercebiotech.com/partnering/merck-bets-up-to-515m-on-an-epigenetic-project-for-cancer-and-blood-disease> (accessed 2025 Oct 20, 2025).
- (30) Hale, C. Celgene Inks \$1B Deal for a Preclinical Epigenetic Blood Cancer Drug from Canada. 2019. <https://www.fiercebiotech.com/biotech/celgene-inks-1b-deal-for-a-preclinical-epigenetic-blood-cancer-drug-from-canada> (accessed 2025 Oct 20, 2025).
- (31) CAS Content Collection. <https://www.cas.org/cas-data> (accessed 2024 March 31).
- (32) Cerneckis, J.; Ming, G. L.; Song, H.; He, C.; Shi, Y. The Rise of Epitranscriptomics: Recent Developments and Future Directions. *Trends Pharmacol. Sci.* **2024**, *45* (1), 24–38.
- (33) RePORTER. <https://reporter.nih.gov/> (accessed 2025 October 20).
- (34) Lavoro, A.; Ricci, D.; Gattuso, G.; Longo, F.; Spoto, G.; Vitale, A. C. V.; Giuliana, M. C.; Falzone, L.; Libra, M.; Candido, S. Recent Advances on Gene-related DNA Methylation in Cancer Diagnosis, Prognosis, and Treatment: A Clinical Perspective. *Clinical Epigenetics* **2025**, *17* (1), 76.
- (35) Marei, H. E. Epigenetic Regulators in Cancer Therapy and Progression. *npj Precision Oncology* **2025**, *9* (1), 206.
- (36) Draskovic, T.; Hauptman, N. Discovery of Novel DNA Methylation Biomarker Panels for the Diagnosis and Differentiation Between Common Adenocarcinomas and Their Liver Metastases. *Sci. Rep.* **2024**, *14* (1), 3095.

- (37) Li, N.; Song, K.; Chen, H.; Dai, M. Advance and Challenge of DNA Methylation as Cancer Biomarkers for Risk Stratification, Screening and Early Detection. *Journal of the National Cancer Center* **2025**, *5* (2), 108–112.
- (38) Hu, C.; Liu, X.; Zeng, Y.; Liu, J.; Wu, F. DNA Methyltransferase Inhibitors Combination Therapy for the Treatment of Solid Tumor: Mechanism and Clinical Application. *Clinical Epigenetics* **2021**, *13* (1), 166.
- (39) Yang, J.; Meng, X.; Pan, J.; Jiang, N.; Zhou, C.; Wu, Z.; Gong, Z. CRISPR/Cas9-mediated Noncoding RNA Editing in Human Cancers. *RNA Biology* **2018**, *15* (1), 35–43.
- (40) M, S. Z.; Hartford, C. C. R.; Lal, A. Interrogating lncRNA Functions Via CRISPR/Cas Systems. *RNA Biology* **2021**, *18* (12), 2097–2106.
- (41) Xu, D.; Cai, Y.; Tang, L.; Han, X.; Gao, F.; Cao, H.; Qi, F.; Kapranov, P. A CRISPR/Cas13-based Approach Demonstrates Biological Relevance of VlinC Class of Long non-coding RNAs in Anticancer Drug Response. *Sci. Rep.* **2020**, *10* (1), 1794.
- (42) Lyu, Q. R.; Zhang, S.; Zhang, Z.; Tang, Z. Functional Knockout of Long Non-coding RNAs with Genome Editing. *Frontiers in Genetics* **2023**, *14*, 1242129.
- (43) Liang, W. W.; Muller, S.; Hart, S. K.; Wessels, H. H.; Mendez-Mancilla, A.; Sookdeo, A.; Choi, O.; Caragine, C. M.; Corman, A.; Lu, L.; Kolumba, O.; Williams, B.; Sanjana, N. E. Transcriptome-scale RNA-targeting CRISPR Screens Reveal Essential lncRNAs in Human Cells. *Cell* **2024**, *187* (26), 7637–7654.e29.
- (44) Cotton, A. M.; Price, E. M.; Jones, M. J.; Balaton, B. P.; Kobor, M. S.; Brown, C. J. Landscape of DNA Methylation on the X Chromosome Reflects CpG Density, Functional Chromatin State and X-Chromosome Inactivation. *Hum. Mol. Genet.* **2015**, *24* (6), 1528–1539.
- (45) Elhamamsy, A. R. Role of DNA Methylation in Imprinting Disorders: An Updated Review. *Journal of Assisted Reproduction and Genetics* **2017**, *34* (5), 549–562.
- (46) Walter, M.; Teissandier, A.; Perez-Palacios, R.; Bourc'h, D. An Epigenetic Switch Ensures Transposon Repression Upon Dynamic Loss of DNA Methylation in Embryonic Stem Cells. *Elife* **2016**, *5*. DOI: 10.7554/eLife.11418.
- (47) Zhou, W.; Liang, G.; Molloy, P. L.; Jones, P. A. DNA Methylation Enables Transposable Element-driven Genome Expansion. *Proc. Natl. Acad. Sci. U.S.A.* **2020**, *117* (32), 19359–19366.
- (48) Robert, M. F.; Morin, S.; Beaulieu, N.; Gauthier, F.; Chute, I. C.; Barsalou, A.; MacLeod, A. R. DNMT1 is Required to Maintain CpG Methylation and Aberrant Gene Silencing in Human Cancer Cells. *Nat. Genet.* **2003**, *33* (1), 61–65.
- (49) Song, J.; Teplova, M.; Ishibe-Murakami, S.; Patel, D. J. Structure-based Mechanistic Insights into DNMT1-mediated Maintenance DNA Methylation. *Science* **2012**, *335* (6069), 709–712.
- (50) Okano, M.; Bell, D. W.; Haber, D. A.; Li, E. DNA Methyltransferases Dnmt3a and Dnmt3b are Essential for De novo Methylation and Mammalian Development. *Cell* **1999**, *99* (3), 247–257.
- (51) Liao, H. F.; Tai, K. Y.; Chen, W. S.; Cheng, L. C.; Ho, H. N.; Lin, S. P. Functions of DNA Methyltransferase 3-like in Germ Cells and Beyond. *Biology of the Cell* **2012**, *104* (10), 571–587.
- (52) Chedin, F.; Lieber, M. R.; Hsieh, C. L. The DNA Methyltransferase-Like Protein DNMT3L Stimulates De novo Methylation by Dnmt3a. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99* (26), 16916–16921.
- (53) Hervouet, E.; Peixoto, P.; Delage-Mourroux, R.; Boyer-Guittaut, M.; Cartron, P. F. Specific or Not Specific Recruitment of DNMTs for DNA Methylation, an Epigenetic Dilemma. *Clinical Epigenetics* **2018**, *10*, 17.
- (54) Jin, B.; Robertson, K. D. DNA Methyltransferases, DNA Damage Repair, and Cancer. *Advances in Experimental Medicine and Biology* **2013**, *754*, 3–29.
- (55) Geissler, F.; Nesic, K.; Kondrashova, O.; Dobrovic, A.; Swisher, E. M.; Scott, C. L.; M, J. W. The Role of Aberrant DNA Methylation in Cancer Initiation and Clinical Impacts. *Therapeutic Advances in Medical Oncology* **2024**, *16*, 17588359231220511.
- (56) Navas-Acien, A.; Domingo-Relloso, A.; Subedi, P.; Riffio-Campos, A. L.; Xia, R.; Gomez, L.; Haack, K.; Goldsmith, J.; Howard, B. V.; Best, L. G.; Devereux, R.; Tauqueer, A.; Zhang, Y.; Fretts, A. M.; Pichler, G.; Levy, D.; Vasan, R. S.; Baccarelli, A. A.; Herreros-Martinez, M.; Tang, W. Y.; Bressler, J.; Fornage, M.; Umans, J. G.; Tellez-Plaza, M.; Fallin, M. D.; Zhao, J.; Cole, S. A. Blood DNA Methylation and Incident Coronary Heart Disease: Evidence From the Strong Heart Study. *JAMA Cardiology* **2021**, *6* (11), 1237–1246.
- (57) Dai, Y.; Chen, D.; Xu, T. DNA Methylation Aberrant in Atherosclerosis. *Frontiers in Pharmacology* **2022**, *13*, 815977.
- (58) Singh, M.; Saxena, S.; Mohan, K. N. DNMT1 Downregulation as Well as Its Overexpression Distinctly Affect Mostly Overlapping Genes Implicated in Schizophrenia, Autism Spectrum, Epilepsy, and Bipolar Disorders. *Frontiers in Molecular Neuroscience* **2023**, *16*, 1275697.
- (59) Poon, C. H.; Tse, L. S. R.; Lim, L. W. DNA Methylation in the Pathology of Alzheimer's Disease: From Gene to Cognition. *Ann. N.Y. Acad. Sci.* **2020**, *1475* (1), 15–33.
- (60) Iwata, K.; Nakabayashi, K.; Ishiwata, K.; Nakamura, K.; Kamen, Y.; Hata, K.; Matsuzaki, H. Genome-wide DNA Methylation Profiles in the Raphe Nuclei of Patients with Autism Spectrum Disorder. *Psychiatry and Clinical Neurosciences* **2025**, *79* (7), 415–424.
- (61) Baek, S. J.; Ban, H. J.; Park, S. M.; Kim, S. Y.; Lee, S.; Jin, H. J. Genome-Wide DNA Methylation Profiling Reveals Candidate Biomarkers and Probable Molecular Mechanism of Metabolic Syndrome. *Genes & Diseases* **2022**, *9* (4), 833–836.
- (62) Womersley, J. S.; Nothling, J.; Toikumo, S.; Malan-Muller, S.; van den Heuvel, L. L.; McGregor, N. W.; Seedat, S.; Hemmings, S. M. J. Childhood Trauma, the Stress Response and Metabolic Syndrome: A Focus on DNA Methylation. *European Journal of Neuroscience* **2022**, *55* (9–10), 2253–2296.
- (63) Glodzik, D.; Bosch, A.; Hartman, J.; Aine, M.; Vallon-Christersson, J.; Reuterswärd, C.; Karlsson, A.; Mitra, S.; Nimeus, E.; Holm, K.; Hakkinen, J.; Hegardt, C.; Saal, L. H.; Larsson, C.; Malmberg, M.; Ryden, L.; Ehinger, A.; Loman, N.; Kvist, A.; Ehrencrona, H.; Nik-Zainal, S.; Borg, A.; Staaf, J. Comprehensive Molecular Comparison of BRCA1 Hypermethylated and BRCA1 Mutated Triple Negative Breast Cancers. *Nat. Commun.* **2020**, *11* (1), 3747.
- (64) Sheaffer, K. L.; Elliott, E. N.; Kaestner, K. H. DNA Hypomethylation Contributes to Genomic Instability and Intestinal Cancer Initiation. *Cancer Prevention Research* **2016**, *9* (7), 534–546.
- (65) Besselink, N.; Keijer, J.; Vermeulen, C.; Boymans, S.; de Ridder, J.; van Hoeck, A.; Cuppen, E.; Kuijk, E. The Genome-Wide Mutational Consequences of DNA Hypomethylation. *Sci. Rep.* **2023**, *13* (1), 6874.
- (66) Jung, M.; Pfeifer, G. P. Aging and DNA Methylation. *BMC Biology* **2015**, *13*, 7.
- (67) Ashapkin, V. V.; Kutueva, L. I.; Vanyushin, B. F. Aging Epigenetics: Accumulation of Errors or Realization of a Specific Program? *Biochemistry (Mosc)* **2015**, *80* (11), 1406–1417.
- (68) Vaidya, H.; Jelinek, J.; Issa, J. J. DNA Methylation, Aging, and Cancer. *Epigenomes* **2025**, *9* (2), 18.
- (69) Handy, D. E.; Castro, R.; Loscalzo, J. Epigenetic Modifications: Basic Mechanisms and Role in Cardiovascular Disease. *Circulation* **2011**, *123* (19), 2145–2156.
- (70) Fenley, A. T.; Anandakrishnan, R.; Kidane, Y. H.; Onufriev, A. V. Modulation of Nucleosomal DNA Accessibility Via Charge-altering Post-translational Modifications in Histone Core. *Epigenetics Chromatin* **2018**, *11* (1), 11.
- (71) Liu, R.; Wu, J.; Guo, H.; Yao, W.; Li, S.; Lu, Y.; Jia, Y.; Liang, X.; Tang, J.; Zhang, H. Post-translational Modifications of Histones: Mechanisms, Biological Functions, and Therapeutic Targets. *MedComm* (2020) **2023**, *4* (3), No. e292.
- (72) Sadakierska-Chudy, A.; Filip, M. A Comprehensive View of the Epigenetic Landscape. Part II: Histone Post-translational Modification, Nucleosome Level, and Chromatin Regulation by ncRNAs. *Neurotoxicity Research* **2015**, *27* (2), 172–197.
- (73) Kimura, H. Histone Modifications for Human Epigenome Analysis. *Journal of Human Genetics* **2013**, *58* (7), 439–445.

- (74) Kouzarides, T. Chromatin Modifications and Their Function. *Cell* **2007**, 128 (4), 693–705.
- (75) Marino-Ramirez, L.; Kann, M. G.; Shoemaker, B. A.; Landsman, D. Histone Structure and Nucleosome Stability. *Expert Review of Proteomics* **2005**, 2 (5), 719–729.
- (76) Kitazawa, R.; Haraguchi, R.; Kitazawa, S. Histone Modification in Histochemistry and Cytochemistry. *Acta Histochemica et Cytochemica* **2023**, 56 (3), 41–47.
- (77) Yang, Y.; Zhang, M.; Wang, Y. The Roles of Histone Modifications in Tumorigenesis and Associated Inhibitors in Cancer Therapy. *Journal of the National Cancer Center* **2022**, 2 (4), 277–290.
- (78) Park, J.; Lee, K.; Kim, K.; Yi, S. J. The Role of Histone Modifications: From Neurodevelopment to Neurodegeneration. *Signal Transduction and Targeted Therapy* **2022**, 7 (1), 217.
- (79) Gao, Z.; Fu, H. J.; Zhao, L. B.; Sun, Z. Y.; Yang, Y. F.; Zhu, H. Y. Aberrant DNA Methylation Associated with Alzheimer's Disease in the Superior Temporal Gyrus. *Experimental and Therapeutic Medicine* **2017**, 15 (1), 103–108.
- (80) Cao, Q.; Wang, W.; Williams, J. B.; Yang, F.; Wang, Z. J.; Yan, Z. Targeting Histone K4 Trimethylation for Treatment of Cognitive and Synaptic Deficits in Mouse Models of Alzheimer's Disease. *Science Advances* **2020**, 6 (50). DOI: 10.1126/sciadv.abc8096.
- (81) Sadri-Vakili, G.; Cha, J. H. Mechanisms of Disease: Histone Modifications in Huntington's Disease. *Nature Clinical Practice Neurology* **2006**, 2 (6), 330–338.
- (82) Francelle, L.; Lotz, C.; Outeiro, T.; Brouillet, E.; Merienne, K. Contribution of Neuroepigenetics to Huntington's Disease. *Frontiers in Human Neuroscience* **2017**, 11, 17.
- (83) Fard, Y. A.; Sadeghi, E. N.; Pajooheh, Z.; Gharehdaghi, Z.; Khatibi, D. M.; Khosravifar, S.; Pishkari, Y.; Nozari, S.; Hijazi, A.; Pakmehr, S.; Shayan, S. K. Epigenetic Underpinnings of the Autistic Mind: Histone Modifications and Prefrontal Excitation/Inhibition Imbalance. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics* **2024**, 195 (8), No. e32986.
- (84) Cheung, P.; Vallania, F.; Warsinske, H. C.; Donato, M.; Schaffert, S.; Chang, S. E.; Dvorak, M.; Dekker, C. L.; Davis, M. M.; Utz, P. J.; Khatri, P.; Kuo, A. J. Single-Cell Chromatin Modification Profiling Reveals Increased Epigenetic Variations with Aging. *Cell* **2018**, 173 (6), 1385–1397.e14. From NLM Medline
- (85) Araki, Y.; Mimura, T. The Histone Modification Code in the Pathogenesis of Autoimmune Diseases. *Mediators of Inflammation* **2017**, 2017, 2608605.
- (86) Yao, W.; Hu, X.; Wang, X. Crossing Epigenetic Frontiers: The Intersection of Novel Histone Modifications and Diseases. *Signal Transduction and Targeted Therapy* **2024**, 9 (1), 232.
- (87) Jenuwein, T.; Allis, C. D. Translating the Histone Code. *Science* **2001**, 293 (5532), 1074–1080.
- (88) Turner, B. M. Cellular Memory and the Histone Code. *Cell* **2002**, 111 (3), 285–291.
- (89) Falkenberg, K. J.; Johnstone, R. W. Histone Deacetylases and Their Inhibitors in Cancer, Neurological Diseases and Immune Disorders. *Nat. Rev. Drug Discovery* **2014**, 13 (9), 673–691.
- (90) Shi, M. Q.; Xu, Y.; Fu, X.; Pan, D. S.; Lu, X. P.; Xiao, Y.; Jiang, Y. Z. Advances in Targeting Histone Deacetylase for Treatment of Solid Tumors. *Journal of Hematology & Oncology* **2024**, 17 (1), 37.
- (91) Hedayat, F.; Faghfuri, E. Harnessing Histone Deacetylase Inhibitors for Enhanced Cancer Immunotherapy. *Eur. J. Pharmacol.* **2025**, 997, 177620.
- (92) Uppaluri, K. R.; Challa, H. J.; Gaur, A.; Jain, R.; Krishna Vardhani, K.; Geddam, A.; Natya, K.; Aswini, K.; Palasamudram, K.; K. S. M. Unlocking the Potential of Non-coding RNAs in Cancer Research and Therapy. *Translational Oncology* **2023**, 35, 101730.
- (93) Esteller, M. Non-Coding RNAs in Human Disease. *Nat. Rev. Genet.* **2011**, 12 (12), 861–874.
- (94) Mattick, J. S.; Makunin, I. V. Non-coding RNA. *Hum. Mol. Genet.* **2006**, 15 (Spec No 1), R17–29.
- (95) Brillante, S.; Volpe, M.; Indrieri, A. Advances in MicroRNA Therapeutics: From Preclinical to Clinical Studies. *Hum. Gene Ther.* **2024**, 35 (17–18), 628–648.
- (96) Martino, M. T. D.; Tagliaferri, P.; Tassone, P. MicroRNA in Cancer Therapy: Breakthroughs and Challenges in Early Clinical Applications. *Journal of Experimental & Clinical Cancer Research* **2025**, 44 (1), 126.
- (97) Mattick, J. S.; Amaral, P. P.; Carninci, P.; Carpenter, S.; Chang, H. Y.; Chen, L. L.; Chen, R.; Dean, C.; Dinger, M. E.; Fitzgerald, K. A.; Gingeras, T. R.; Guttman, M.; Hirose, T.; Huarte, M.; Johnson, R.; Kanduri, C.; Kapranov, P.; Lawrence, J. B.; Lee, J. T.; Mendell, J. T.; Mercer, T. R.; Moore, K. J.; Nakagawa, S.; Rinn, J. L.; Spector, D. L.; Ulitsky, I.; Wan, Y.; Wilusz, J. E.; Wu, M. Long Non-coding RNAs: Definitions, Functions, Challenges and Recommendations. *Nat. Rev. Mol. Cell Biol.* **2023**, 24 (6), 430–447.
- (98) Liao, J.; Wang, J.; Liu, Y.; Li, J.; Duan, L. Transcriptome Sequencing of lncRNA, miRNA, mRNA and Interaction Network Constructing in Coronary Heart Disease. *BMC Medical Genomics* **2019**, 12 (1), 124.
- (99) Chowdhary, A.; Satagopam, V.; Schneider, R. Long Non-coding RNAs: Mechanisms, Experimental, and Computational Approaches in Identification, Characterization, and Their Biomarker Potential in Cancer. *Frontiers in Genetics* **2021**, 12, 649619.
- (100) Hu, B.; Zhong, L.; Weng, Y.; Peng, L.; Huang, Y.; Zhao, Y.; Liang, X. J. Therapeutic siRNA: State of the Art. *Signal Transduction and Targeted Therapy* **2020**, 5 (1), 101.
- (101) Alshaer, W.; Zureigat, H.; Al Karaki, A.; Al-Kadash, A.; Gharaibeh, L.; Hatmal, M. M.; Aljabali, A. A. A.; Awidi, A. siRNA: Mechanism of Action, Challenges, and Therapeutic Approaches. *Eur. J. Pharmacol.* **2021**, 905, 174178.
- (102) Toth, K. F.; Pezic, D.; Stuwe, E.; Webster, A. The piRNA Pathway Guards the Germline Genome Against Transposable Elements. *Advances in Experimental Medicine and Biology* **2016**, 886, 51–77.
- (103) Zhang, H.; Zhang, F.; Chen, Q.; Li, M.; Lv, X.; Xiao, Y.; Zhang, Z.; Hou, L.; Lai, Y.; Zhang, Y.; Zhang, A.; Gao, S.; Fu, H.; Xiao, W.; Zhou, J.; Diao, F.; Shi, A.; Su, Y. Q.; Zeng, W.; Wu, L.; Li, J. The piRNA Pathway is Essential for Generating Functional Oocytes in Golden Hamsters. *Nat. Cell Biol.* **2021**, 23 (9), 1013–1022.
- (104) Loubalova, Z.; Fulka, H.; Horvat, F.; Pasulka, J.; Malik, R.; Hirose, M.; Ogura, A.; Svoboda, P. Formation of Spermatogonia and Fertile Oocytes in Golden Hamsters Requires piRNAs. *Nat. Cell Biol.* **2021**, 23 (9), 992–1001.
- (105) Hasuwa, H.; Iwasaki, Y. W.; Au Yeung, W. K.; Ishino, K.; Masuda, H.; Sasaki, H.; Siomi, H. Production of Functional Oocytes Requires Maternally Expressed PIWI Genes and piRNAs in Golden Hamsters. *Nat. Cell Biol.* **2021**, 23 (9), 1002–1012.
- (106) Ding, D.; Chen, C. Cracking the Egg: A Breakthrough in piRNA Function in Mammalian Oocytes and Embryos. *Biol. Reprod.* **2022**, 106 (1), 6–8.
- (107) Claro-Linares, F.; Rojas-Rios, P. PIWI Proteins and piRNAs: Key Regulators of Stem Cell Biology. *Frontiers in Cell Developmental Biology* **2025**, 13, 1540313.
- (108) Landry, C. D.; Kandel, E. R.; Rajasethupathy, P. New Mechanisms in Memory Storage: piRNAs and Epigenetics. *Trends in Neurosciences* **2013**, 36 (9), 535–542.
- (109) Sato, K.; Takayama, K. I.; Inoue, S. Role of piRNA Biogenesis and Its Neuronal Function in the Development of Neurodegenerative Diseases. *Frontiers in Aging Neuroscience* **2023**, 15, 1157818.
- (110) Beatini, S.; La Rosa, L.; Giantomasi, L.; De Pietri Tonelli, D. PIWI-interacting RNAs in Neurological and Neuropsychiatric Disorders and Potential for Clinical Development. *Brain Organoid & Systems Neuroscience Journal* **2025**, 3, 154–169.
- (111) Liu, Y.; Dou, M.; Song, X.; Dong, Y.; Liu, S.; Liu, H.; Tao, J.; Li, W.; Yin, X.; Xu, W. The Emerging Role of the piRNA/piwi Complex in Cancer. *Molecular Cancer* **2019**, 18 (1), 123.
- (112) Garcia-Borja, E.; Siegl, F.; Mateu, R.; Slaby, O.; Sedo, A.; Busek, P.; Sana, J. Critical Appraisal of the piRNA-PIWI Axis in Cancer and Cancer Stem Cells. *Biomark Research* **2024**, 12 (1), 15.
- (113) Hanusek, K.; Poletajew, S.; Kryst, P.; Piekliko-Witkowska, A.; Boguslawska, J. piRNAs and PIWI Proteins as Diagnostic and

Prognostic Markers of Genitourinary Cancers. *Biomolecules* **2022**, *12* (2), 186.

(114) Ahmadi Asouri, S.; Aghadavood, E.; Mirzaei, H.; Abaspour, A.; Esmail Shahaboddin, M. PIWI-interacting RNAs (PiRNAs) as Emerging Biomarkers and Therapeutic Targets in Biliary Tract Cancers: A Comprehensive Review. *Heliyon* **2024**, *10* (13), No. e33767.

(115) Kumar, S.; Gonzalez, E. A.; Rameshwar, P.; Etchegaray, J. P. Non-Coding RNAs as Mediators of Epigenetic Changes in Malignancies. *Cancers (Basel)* **2020**, *12* (12), 3657.

(116) Peschansky, V. J.; Wahlestedt, C. Non-coding RNAs as Direct and Indirect Modulators of Epigenetic Regulation. *Epigenetics* **2014**, *9* (1), 3–12.

(117) Statello, L.; Guo, C. J.; Chen, L. L.; Huarte, M. Gene Regulation by Long Non-coding RNAs and Its Biological Functions. *Nat. Rev. Mol. Cell Biol.* **2021**, *22* (2), 96–118.

(118) Wang, K. C.; Chang, H. Y. Molecular Mechanisms of Long Noncoding RNAs. *Mol. Cell* **2011**, *43* (6), 904–914.

(119) Mangiavacchi, A.; Morelli, G.; Orlando, V. Behind the Scenes: How RNA Orchestrates the Epigenetic Regulation of Gene Expression. *Frontiers in Cell and Developmental Biology* **2023**, *11*, 1123975.

(120) Irwin, A. B.; Bahabry, R.; Lubin, F. D. A Putative Role for lncRNAs in Epigenetic Regulation of Memory. *Neurochem. Int.* **2021**, *150*, 105184.

(121) Much, C.; Lasda, E. L.; Pereira, I. T.; Vallery, T. K.; Ramirez, D.; Lewandowski, J. P.; Dowell, R. D.; Smallegan, M. J.; Rinn, J. L. The Temporal Dynamics of lncRNA Firre-mediated Epigenetic and Transcriptional Regulation. *Nat. Commun.* **2024**, *15* (1), 6821.

(122) Phillips, T.; Shaw, K. Chromatin Remodeling in Eukaryotes. *Nature Education* **2008**, *1* (1), 209.

(123) Saha, D.; Animireddy, S.; Bartholomew, B. The SWI/SNF ATP-dependent Chromatin Remodeling Complex in Cell Lineage Priming and Early Development. *Biochem. Soc. Trans.* **2024**, *52* (2), 603–616.

(124) Goldman, J. A.; Garlick, J. D.; Kingston, R. E. Chromatin Remodeling by Imitation Switch (ISWI) Class ATP-dependent Remodelers is Stimulated by Histone Variant H2A.Z. *J. Biol. Chem.* **2010**, *285* (7), 4645–4651.

(125) Li, Y.; Gong, H.; Wang, P.; Zhu, Y.; Peng, H.; Cui, Y.; Li, H.; Liu, J.; Wang, Z. The Emerging Role of ISWI Chromatin Remodeling Complexes in Cancer. *Journal of Experimental & Clinical Cancer Research* **2021**, *40* (1), 346.

(126) Murawska, M.; Brehm, A. CHD Chromatin Remodelers and the Transcription Cycle. *Transcription* **2011**, *2* (6), 244–253.

(127) Morrison, A. J.; Shen, X. Chromatin Remodelling Beyond Transcription: The INO80 and SWR1 Complexes. *Nat. Rev. Mol. Cell Biol.* **2009**, *10* (6), 373–384.

(128) Hargreaves, D. C.; Crabtree, G. R. ATP-dependent Chromatin Remodeling: Genetics, Genomics and Mechanisms. *Cell Research* **2011**, *21* (3), 396–420.

(129) Sahu, R. K.; Singh, S.; Tomar, R. S. The Mechanisms of Action of Chromatin Remodelers and Implications in Development and Disease. *Biochem. Pharmacol.* **2020**, *180*, 114200.

(130) Vignali, M.; Hassan, A. H.; Neely, K. E.; Workman, J. L. ATP-dependent Chromatin-remodeling Complexes. *Mol. Cell. Biol.* **2000**, *20* (6), 1899–1910.

(131) Bowman, G. D. Mechanisms of ATP-Dependent Nucleosome Sliding. *Curr. Opin. Struct. Biol.* **2010**, *20* (1), 73–81.

(132) Park, S.; Brandani, G. B.; Ha, T.; Bowman, G. D. Bi-directional Nucleosome Sliding by the Chd1 Chromatin Remodeler Integrates Intrinsic Sequence-dependent and ATP-dependent Nucleosome Positioning. *Nucleic Acids Res.* **2023**, *51* (19), 10326–10343.

(133) Clapier, C. R.; Iwasa, J.; Cairns, B. R.; Peterson, C. L. Mechanisms of Action and Regulation of ATP-Dependent Chromatin-Remodelling Complexes. *Nat. Rev. Mol. Cell Biol.* **2017**, *18* (7), 407–422.

(134) Geiman, T. M.; Robertson, K. D. Chromatin Remodeling, Histone Modifications, and DNA Methylation-How Does It All Fit Together? *Journal of Cellular Biochemistry* **2002**, *87* (2), 117–125.

(135) Choudhuri, S. Chapter 1 - Fundamentals of Genes and Genomes. In *Bioinformatics for Beginners*; Choudhuri, S., Ed.; Academic Press, 2014; pp 1–25.

(136) Martire, S.; Banaszynski, L. A. The Roles of Histone Variants in Fine-tuning Chromatin Organization and Function. *Nat. Rev. Mol. Cell Biol.* **2020**, *21* (9), 522–541.

(137) Kumar, R.; Li, D. Q.; Muller, S.; Knapp, S. Epigenomic Regulation of Oncogenesis by Chromatin Remodeling. *Oncogene* **2016**, *35* (34), 4423–4436.

(138) Sehgal, P.; Chaturvedi, P. Chromatin and Cancer: Implications of Disrupted Chromatin Organization in Tumorigenesis and Its Diversification. *Cancers (Basel)* **2023**, *15* (2), 466.

(139) Bastle, R. M.; Maze, I. Chromatin Regulation in Complex Brain Disorders. *Current Opinion in Behavioural Sciences* **2019**, *25*, 57–65.

(140) Larizza, L.; Finelli, P. Developmental Disorders with Intellectual Disability Driven by Chromatin Dysregulation: Clinical Overlaps and Molecular Mechanisms. *Clinical Genetics* **2019**, *95* (2), 231–240.

(141) Mossink, B.; Negwer, M.; Schubert, D.; Nadif Kasri, N. The Emerging Role of Chromatin Remodelers in Neurodevelopmental Disorders: A Developmental Perspective. *Cell. Mol. Life Sci.* **2021**, *78* (6), 2517–2563.

(142) Takeda, T.; Banno, K.; Okawa, R.; Yanokura, M.; Iijima, M.; Irie-Kunitomi, H.; Nakamura, K.; Iida, M.; Adachi, M.; Umene, K.; Nogami, Y.; Masuda, K.; Kobayashi, Y.; Tominaga, E.; Aoki, D. ARID1A Gene Mutation in Ovarian and Endometrial Cancers. *Oncol. Rep.* **2016**, *35* (2), 607–613.

(143) Tokunaga, R.; Xiu, J.; Goldberg, R. M.; Philip, P. A.; Seeber, A.; Battaglin, F.; Arai, H.; Lo, J. H.; Naseem, M.; Puccini, A.; Berger, M. D.; Soni, S.; Zhang, W.; Chen, S.; Hwang, J. J.; Shields, A. F.; Marshall, J. L.; Baba, H.; Korn, W. M.; Lenz, H. J. The Impact of ARID1A Mutation on Molecular Characteristics in Colorectal Cancer. *Eur. J. Cancer* **2020**, *140*, 119–129.

(144) Malone, H. A.; Roberts, C. W. M. Chromatin Remodellers as Therapeutic Targets. *Nat. Rev. Drug Discovery* **2024**, *23* (9), 661–681.

(145) Lim, J.; Park, C.; Kim, M.; Kim, H.; Kim, J.; Lee, D. S. Advances in Single-cell Omics and Multiomics for High-resolution Molecular Profiling. *Experimental & Molecular Medicine* **2024**, *56* (3), 515–526.

(146) Kang, B.; Lee, H.; Roh, T. Y. Deciphering Single-cell Genomic Architecture: Insights into Cellular Heterogeneity and Regulatory Dynamics. *Genomics & Informatics* **2025**, *23* (1), 5.

(147) Hu, Y.; Shen, F.; Yang, X.; Han, T.; Long, Z.; Wen, J.; Huang, J.; Shen, J.; Guo, Q. Single-cell Sequencing Technology Applied to Epigenetics for the Study of Tumor Heterogeneity. *Clinical Epigenetics* **2023**, *15* (1), 161.

(148) Fang, L.; Wuptra, K.; Chen, D.; Li, H.; Huang, S. K.; Jin, C.; Yokoyama, K. K. Environmental-stress-induced Chromatin Regulation and its Heritability. *Journal of Carcinogenesis & Mutagenesis* **2014**, *5* (1), 22058.

(149) Suter, M. A.; Aagaard-Tillery, K. M. Environmental Influences on Epigenetic Profiles. *Seminars in Reproductive Medicine* **2009**, *27* (5), 380–390.

(150) Breton, C. V.; Landon, R.; Kahn, L. G.; Enlow, M. B.; Peterson, A. K.; Bastain, T.; Braun, J.; Comstock, S. S.; Duarte, C. S.; Hipwell, A.; Ji, H.; LaSalle, J. M.; Miller, R. L.; Musci, R.; Posner, J.; Schmidt, R.; Suglia, S. F.; Tung, I.; Weisenberger, D.; Zhu, Y.; Fry, R. Exploring the Evidence for Epigenetic Regulation of Environmental Influences on Child Health Across Generations. *Communications Biology* **2021**, *4* (1), 769.

(151) Kumaraswamy, A.; Welker Leng, K. R.; Westbrook, T. C.; Yates, J. A.; Zhao, S. G.; Evans, C. P.; Feng, F. Y.; Morgan, T. M.; Alumkal, J. J. Recent Advances in Epigenetic Biomarkers and Epigenetic Targeting in Prostate Cancer. *European Urology* **2021**, *80* (1), 71–81.

(152) Jin, N.; George, T. L.; Otterson, G. A.; Verschraegen, C.; Wen, H.; Carbone, D.; Herman, J.; Bertino, E. M.; He, K. Advances in Epigenetic Therapeutics with Focus on Solid Tumors. *Clinical Epigenetics* **2021**, *13* (1), 83.

(153) Kanwal, R.; Gupta, S. Epigenetic Modifications in Cancer. *Clinical Genetics* **2012**, *81* (4), 303–311.

- (154) Sharma, S.; Kelly, T. K.; Jones, P. A. Epigenetics in Cancer. *Carcinogenesis* **2010**, *31* (1), 27–36.
- (155) Ren, L.; Yang, Y.; Li, W.; Yang, H.; Zhang, Y.; Ge, B.; Zhang, S.; Du, G.; Wang, J. Recent Advances in Epigenetic Anticancer Therapeutics and Future Perspectives. *Frontiers in Genetics* **2023**, *13*, 1085391.
- (156) Majchrzak-Celinska, A.; Warych, A.; Szoszkiewicz, M. Novel Approaches to Epigenetic Therapies: From Drug Combinations to Epigenetic Editing. *Genes (Basel)* **2021**, *12* (2), 208.
- (157) Yao, Q.; Chen, Y.; Zhou, X. The Roles of microRNAs in Epigenetic Regulation. *Curr. Opin. Chem. Biol.* **2019**, *51*, 11–17.
- (158) Lee, J. T. Epigenetic Regulation by Long Noncoding RNAs. *Science* **2012**, *338* (6113), 1435–1439.
- (159) Mehrmohamadi, M.; Sepehri, M. H.; Nazer, N.; Norouzi, M. R. A Comparative Overview of Epigenomic Profiling Methods. *Frontiers in Cell and Developmental Biology* **2021**, *9*, 714687.
- (160) Meers, M. P.; Llagas, G.; Janssens, D. H.; Codomo, C. A.; Henikoff, S. Multifactorial Profiling of Epigenetic Landscapes at Single-cell Resolution Using MulTI-Tag. *Nat. Biotechnol.* **2023**, *41* (5), 708–716.
- (161) Heyn, H.; Mendez-Gonzalez, J.; Esteller, M. Epigenetic Profiling Joins Personalized Cancer Medicine. *Expert Review of Molecular Diagnostics* **2013**, *13* (5), 473–479.
- (162) Rasool, M.; Malik, A.; Naseer, M. I.; Manan, A.; Ansari, S.; Begum, I.; Qazi, M. H.; Pushparaj, P.; Abuzenadah, A. M.; Al-Qahtani, M. H.; Kamal, M. A.; Gan, S. The Role of Epigenetics in Personalized Medicine: Challenges and Opportunities. *BMC Medical Genomics* **2015**, *8* (Suppl 1), S5.
- (163) Guo, X.; Feng, C.; Xing, J.; Cao, Y.; Liu, T.; Yang, W.; Mu, R.; Wang, T. Epigenetic Profiling for Prognostic Stratification and Personalized Therapy in Breast Cancer. *Frontiers in Immunology* **2025**, *15*, 1510829.
- (164) McCutcheon, S. R.; Rohm, D.; Iglesias, N.; Gersbach, C. A. Epigenome Editing Technologies for Discovery and Medicine. *Nat. Biotechnol.* **2024**, *42* (8), 1199–1217.
- (165) Xu, D.; Besselink, S.; Ramadoss, G. N.; Dierks, P. H.; Lubin, J. P.; Pattali, R. K.; Brim, J. I.; Christenson, A. E.; Colias, P. J.; Ornelas, I. J.; Nguyen, C. D.; Chasins, S. E.; Conklin, B. R.; Nunez, J. K. Programmable Epigenome Editing by Transient Delivery of CRISPR Epigenome Editor Ribonucleoproteins. *Nat. Commun.* **2025**, *16* (1), 7948.
- (166) Clark, S. J.; Lee, H. J.; Smallwood, S. A.; Kelsey, G.; Reik, W. Single-Cell Epigenomics: Powerful New Methods for Understanding Gene Regulation and Cell Identity. *Genome Biology* **2016**, *17*, 72.
- (167) Bird, A. Transgenerational Epigenetic Inheritance: A Critical Perspective. *Frontiers in Epigenetics and Epigenomics* **2024**, *2*, 1434253.
- (168) Zhang, D.; Deng, Y.; Kukanja, P.; Agirre, E.; Bartosovic, M.; Dong, M.; Ma, C.; Ma, S.; Su, G.; Bao, S.; Liu, Y.; Xiao, Y.; Rosoklija, G. B.; Dwork, A. J.; Mann, J. J.; Leong, K. W.; Boldrini, M.; Wang, L.; Haeussler, M.; Raphael, B. J.; Kluger, Y.; Castelo-Branco, G.; Fan, R. Spatial Epigenome-transcriptome Co-profiling of Mammalian Tissues. *Nature* **2023**, *616* (7955), 113–122.
- (169) Gionco, J. T.; Bernstein, A. I. Emerging Role of Environmental Epitranscriptomics and RNA Modifications in Parkinson's Disease. *Journal of Parkinson's Disease* **2024**, *14* (4), 643–656.
- (170) Angelova, M. T.; Dimitrova, D. G.; Dinges, N.; Lence, T.; Worpenberg, L.; Carre, C.; Roignant, J. Y. The Emerging Field of Epitranscriptomics in Neurodevelopmental and Neuronal Disorders. *Frontiers in Bioengineering and Biotechnology* **2018**, *6*, 46.
- (171) Cross, R. Epitranscriptomics: The New RNA Code and the Race to Drug It. *Chemical & Engineering News, C&EN* **2019**, *97*, 34–39.
- (172) Boo, S. H.; Kim, Y. K. The Emerging Role of RNA Modifications in the Regulation of mRNA Stability. *Experimental & Molecular Medicine* **2020**, *52* (3), 400–408.
- (173) Roundtree, I. A.; Evans, M. E.; Pan, T.; He, C. Dynamic RNA Modifications in Gene Expression Regulation. *Cell* **2017**, *169* (7), 1187–1200.
- (174) Ontiveros, R. J.; Stoute, J.; Liu, K. F. The Chemical Diversity of RNA Modifications. *Biochem. J.* **2019**, *476* (8), 1227–1245.
- (175) Esteve-Puig, R.; Bueno-Costa, A.; Esteller, M. Writers, Readers and Erasers of RNA Modifications in Cancer. *Cancer Letters* **2020**, *474*, 127–137.
- (176) Flamand, M. N.; Tegowski, M.; Meyer, K. D. The Proteins of mRNA Modification: Writers, Readers, and Erasers. *Annu. Rev. Biochem.* **2023**, *92*, 145–173.
- (177) Chen, D.; Gu, X.; Nurzat, Y.; Xu, L.; Li, X.; Wu, L.; Jiao, H.; Gao, P.; Zhu, X.; Yan, D.; Li, S.; Xue, C. Writers, Readers, and Erasers RNA Modifications and Drug Resistance in Cancer. *Molecular Cancer* **2024**, *23* (1), 178.
- (178) Fang, Z.; Mei, W.; Qu, C.; Lu, J.; Shang, L.; Cao, F.; Li, F. Role of m6A Writers, Erasers and Readers in Cancer. *Experimental Hematology & Oncology* **2022**, *11* (1), 45.
- (179) Deng, X.; Qing, Y.; Horne, D.; Huang, H.; Chen, J. The Roles and Implications of RNA m(6A) Modification in Cancer. *Nature Reviews Clinical Oncology* **2023**, *20* (8), S07–S26.
- (180) Yin, H.; Zhang, X.; Yang, P.; Zhang, X.; Peng, Y.; Li, D.; Yu, Y.; Wu, Y.; Wang, Y.; Zhang, J.; Ding, X.; Wang, X.; Yang, A.; Zhang, R. RNA m6A Methylation Orchestrates Cancer Growth and Metastasis via Macrophage Reprogramming. *Nat. Commun.* **2021**, *12* (1), 1394.
- (181) Chen, C.; Guo, Y.; Guo, Y.; Wu, X.; Si, C.; Xu, Y.; Kang, Q.; Sun, Z. m6A Modification in Non-Coding RNA: The Role in Cancer Drug Resistance. *Frontiers in Oncology* **2021**, *11*, 746789.
- (182) Han, M.; Liu, Z.; Xu, Y.; Liu, X.; Wang, D.; Li, F.; Wang, Y.; Bi, J. Abnormality of m6A mRNA Methylation Is Involved in Alzheimer's Disease. *Frontiers in Neuroscience* **2020**, *14*, 98.
- (183) Xia, L.; Zhang, F.; Li, Y.; Mo, Y.; Zhang, L.; Li, Q.; Luo, M.; Hou, X.; Du, Z.; Deng, J.; Hao, E. A New Perspective on Alzheimer's Disease: m6A Modification. *Frontiers in Genetics* **2023**, *14*, 1166831.
- (184) Yu, Z.; Huang, L.; Xia, Y.; Cheng, S.; Yang, C.; Chen, C.; Zou, Z.; Wang, X.; Tian, X.; Jiang, X.; Zhou, L. Analysis of m6A Modification Regulators in the Substantia Nigra and Striatum of MPTP-induced Parkinson's Disease Mice. *Neurosci. Lett.* **2022**, *791*, 136907.
- (185) He, H.; Zhang, Q.; Liao, J.; Lei, J.; Luo, M.; Huang, J.; Chen, M.; Shen, Y.; Wang, J.; Xu, P.; Xiao, Y. METTL14 is Decreased and Regulates m(6)A Modification of Alpha-synuclein in Parkinson's Disease. *Journal of Neurochemistry* **2023**, *166* (3), 609–622.
- (186) Zhou, J.; Han, Y.; Hou, R. Potential Role of N6-methyladenosine Modification in the Development of Parkinson's Disease. *Frontiers in Cell and Developmental Biology* **2023**, *11*, 1321995.
- (187) Li, Y.; Dou, X.; Liu, J.; Xiao, Y.; Zhang, Z.; Hayes, L.; Wu, R.; Fu, X.; Ye, Y.; Yang, B.; Ostrow, L. W.; He, C.; Sun, S. Globally Reduced N(6)-methyladenosine (m(6)A) in C9ORF72-ALS/FTD Dysregulates RNA Metabolism and Contributes to Neurodegeneration. *Nature Neuroscience* **2023**, *26* (8), 1328–1338.
- (188) Locatelli, M.; Faure-Dupuy, S. Virus Hijacking of Host Epigenetic Machinery to Impair Immune Response. *Journal of Virology* **2023**, *97* (9), No. e0065823. From NLM Medline
- (189) Yang, Y.; Lu, Y.; Wang, Y.; Wen, X.; Qi, C.; Piao, W.; Jin, H. Current Progress in Strategies to Profile Transcriptomic m(6)A Modifications. *Frontiers in Cell and Developmental Biology* **2024**, *12*, 1392159.
- (190) Zhang, Y.; Lu, L.; Li, X. Detection Technologies for RNA Modifications. *Experimental & Molecular Medicine* **2022**, *54* (10), 1601–1616.
- (191) *Charting a Future for Sequencing RNA and Its Modifications: A New Era for Biology and Medicine*; National Academies Press (US), 2024.
- (192) Li, Y.; Su, R.; Deng, X.; Chen, Y.; Chen, J. FTO in Cancer: Functions, Molecular Mechanisms, and Therapeutic Implications. *Trends in Cancer* **2022**, *8* (7), 598–614.
- (193) Qiao, Y.; Su, M.; Zhao, H.; Liu, H.; Wang, C.; Dai, X.; Liu, L.; Liu, G.; Sun, H.; Sun, M.; Wang, J.; Li, Z.; Fan, J.; Zhang, Q.; Li, C.; Situ, F.; Xue, J.; Jia, Z.; Zhang, C.; Zhang, S.; Shan, C. Targeting FTO Induces Colorectal Cancer Ferroptotic Cell Death by Decreasing SLC7A11/GPX4 Expression. *Journal of Experimental & Clinical Cancer Research* **2024**, *43* (1), 108.
- (194) Liang, X.; Huang, Y.; Ren, H.; Liu, Q.; Chen, L.; Zhao, J.; Gao, X.; Lu, J.; Yang, C. G.; Liu, H. Discovery of Novel RNA Demethylase

FTO Inhibitors Featuring an Acylhydrazone Scaffold with Potent Antileukemia Activity. *J. Med. Chem.* **2025**, *68* (3), 2742–2763.

(195) Kim, S. C.; Sekhon, S. S.; Shin, W. R.; Ahn, G.; Cho, B. K.; Ahn, J. Y.; Kim, Y. H. Modifications of mRNA Vaccine Structural Elements for Improving mRNA Stability and Translation Efficiency. *Molecular & Cellular Toxicology* **2022**, *18* (1), 1–8.

(196) Liu, A.; Wang, X. The Pivotal Role of Chemical Modifications in mRNA Therapeutics. *Frontiers in Cell and Developmental Biology* **2022**, *10*, 901510.

(197) Kadumuri, R. V.; Janga, S. C. Epitranscriptomic Code and Its Alterations in Human Disease. *Trends in Molecular Medicine* **2018**, *24* (10), 886–903.

(198) Chang, H. Y.; Qi, L. S. Reversing the Central Dogma: RNA-guided Control of DNA in Epigenetics and Genome Editing. *Mol. Cell* **2023**, *83* (3), 442–451.

(199) Crespo-Garcia, E.; Bueno-Costa, A.; Esteller, M. Single-Cell Analysis of the Epitranscriptome: RNA Modifications Under the Microscope. *RNA Biology* **2024**, *21* (1), 304–311.

(200) Wang, H.; Wang, Y.; Zhou, J.; Song, B.; Tu, G.; Nguyen, A.; Su, J.; Coenen, F.; Wei, Z.; Rigden, D. J.; Meng, J. Statistical Modeling of Single-cell Epitranscriptomics Enabled Trajectory and Regulatory Inference of RNA Methylation. *Cell Genomics* **2025**, *5* (1), 100702.

(201) Fustin, J. M.; Doi, M.; Yamaguchi, Y.; Hida, H.; Nishimura, S.; Yoshida, M.; Isagawa, T.; Morioka, M. S.; Kakeya, H.; Manabe, I.; Okamura, H. RNA-methylation-dependent RNA Processing Controls the Speed of the Circadian Clock. *Cell* **2013**, *155* (4), 793–806.

(202) Lokody, I. Gene Regulation: RNA Methylation Regulates the Circadian Clock. *Nat. Rev. Genet.* **2014**, *15* (1), 3.

(203) Dedon, P. C.; Begley, T. J. A System of RNA Modifications and Biased Codon Use Controls Cellular Stress Response at the Level of Translation. *Chem. Res. Toxicol.* **2014**, *27* (3), 330–337.

(204) Dannfald, A.; Favory, J. J.; Deragon, J. M. Variations in Transfer and Ribosomal RNA Epitranscriptomic Status can Adapt Eukaryote Translation to Changing Physiological and Environmental Conditions. *RNA Biology* **2021**, *18* (sup1), 4–18.

(205) Alum, E. U.; Ejemot-Nwadiaro, R. I.; Basajja, M.; Uti, D. E.; Ugwu, O. P.; Aja, P. M. Epitranscriptomic Alterations Induced by Environmental Toxins: Implications for RNA Modifications and Disease. *Genes and Environment* **2025**, *47* (1), 14.

(206) Ilyas, M.; Shah, Q.; Gul, A.; Ibrahim, H.; Fatima, R.; Babar, M. M.; Rajadas, J. Advances in CRISPR-Cas Systems for Epigenetics. *Progress in Molecular Biology and Translational Science* **2024**, *208*, 185–209.

(207) Nakamura, M.; Gao, Y.; Dominguez, A. A.; Qi, L. S. CRISPR Technologies for Precise Epigenome Editing. *Nat. Cell Biol.* **2021**, *23* (1), 11–22.

(208) Fadul, S. M.; Arshad, A.; Mehmood, R. CRISPR-Based Epigenome Editing: Mechanisms and Applications. *Epigenomics* **2023**, *15* (21), 1137–1155.

(209) Jeffries, M. A. Epigenetic Editing: How Cutting-edge Targeted Epigenetic Modification Might Provide Novel Avenues for Auto-immune Disease Therapy. *Clinical Immunology* **2018**, *196*, 49–58.

(210) O'Geen, H.; Bates, S. L.; Carter, S. S.; Nisson, K. A.; Halmaj, J.; Fink, K. D.; Rhie, S. K.; Farnham, P. J.; Segal, D. J. Ezh2-dCas9 and KRAB-dCas9 Enable Engineering of Epigenetic Memory in a Context-dependent Manner. *Epigenetics & Chromatin* **2019**, *12* (1), 26.

(211) Pulecio, J.; Verma, N.; Mejia-Ramirez, E.; Huangfu, D.; Raya, A. CRISPR/Cas9-Based Engineering of the Epigenome. *Cell Stem Cell* **2017**, *21* (4), 431–447.

(212) Kang, J. G.; Park, J. S.; Ko, J. H.; Kim, Y. S. Regulation of Gene Expression by Altered Promoter Methylation Using a CRISPR/Cas9-mediated Epigenetic Editing System. *Sci. Rep.* **2019**, *9* (1), 11960.

(213) Pacalin, N. M.; Steinhart, Z.; Shi, Q.; Belk, J. A.; Dorovskiy, D.; Kraft, K.; Parker, K. R.; Shy, B. R.; Marson, A.; Chang, H. Y. Bidirectional Epigenetic Editing Reveals Hierarchies in Gene Regulation. *Nat. Biotechnol.* **2025**, *43* (3), 355–368.

(214) Black, J. B.; Adler, A. F.; Wang, H. G.; D'Ippolito, A. M.; Hutchinson, H. A.; Reddy, T. E.; Pitt, G. S.; Leong, K. W.; Gersbach, C. A. Targeted Epigenetic Remodeling of Endogenous Loci by CRISPR/

Cas9-Based Transcriptional Activators Directly Converts Fibroblasts to Neuronal Cells. *Cell Stem Cell* **2016**, *19* (3), 406–414.

(215) Moses, C.; Hodgetts, S. I.; Nugent, F.; Ben-Ary, G.; Park, K. K.; Blancafort, P.; Harvey, A. R. Transcriptional Repression of PTEN in Neural Cells using CRISPR/dCas9 Epigenetic Editing. *Sci. Rep.* **2020**, *10* (1), 11393.

(216) Tejedor, J. R.; Penarroya, A.; Gancedo-Verdejo, J.; Santamarina-Ojeda, P.; Perez, R. F.; Lopez-Tamargo, S.; Diez-Borge, A.; Alba-Linares, J. J.; Gonzalez-Del-Rey, N.; Urdinguio, R. G.; Mangas, C.; Roberti, A.; Lopez, V.; Morales-Ruiz, T.; Ariza, R. R.; Roldan-Arjona, T.; Meijon, M.; Valledor, L.; Canal, M. J.; Fernandez-Martinez, D.; Fernandez-Hevia, M.; Jimenez-Fonseca, P.; Garcia-Florez, L. J.; Fernandez, A. F.; Fraga, M. F. CRISPR/dCAS9-mediated DNA Demethylation Screen Identifies Functional Epigenetic Determinants of Colorectal Cancer. *Clinical Epigenetics* **2023**, *15* (1), 133.

(217) Goell, J. H.; Hilton, I. B. CRISPR/Cas-Based Epigenome Editing: Advances, Applications, and Clinical Utility. *Trends Biotechnol.* **2021**, *39* (7), 678–691.

(218) Zahraei, M.; Azimi, Y.; Karimipour, M.; Rahimi-Jamnani, F.; Valizadeh, V.; Azizi, M. CRISPR/dCas9-TET1-mediated Epigenetic Editing Reactivates miR-200c in Breast Cancer Cells. *Sci. Rep.* **2025**, *15* (1), 31739.

(219) Tremblay, F.; Xiong, Q.; Shah, S. S.; Ko, C. W.; Kelly, K.; Morrison, M. S.; Giancarlo, C.; Ramirez, R. N.; Hildebrand, E. M.; Voytek, S. B.; El Sebae, G. K.; Wright, S. H.; Lofgren, L.; Clarkson, S.; Waters, C.; Linder, S. J.; Liu, S.; Eom, T.; Parikh, S.; Weber, Y.; Martinez, S.; Malyala, P.; Abubucker, S.; Friedland, A. E.; Maeder, M. L.; Lombardo, A.; Myer, V. E.; Jaffe, A. B. A. Potent Epigenetic Editor Targeting Human PCSK9 for Durable Reduction of Low-density Lipoprotein Cholesterol Levels. *Nature Medicine* **2025**, *31* (4), 1329–1338.

(220) Azeez, S. S.; Hamad, R. S.; Hamad, B. K.; Shekha, M. S.; Bergsten, P. Advances in CRISPR-Cas Technology and Its Applications: Revolutionising Precision Medicine. *Frontiers in Genome Editing* **2024**, *6*, 1509924.

(221) Cappelluti, M. A.; Mollica Poeta, V.; Valsoni, S.; Quarato, P.; Merlin, S.; Merelli, I.; Lombardo, A. Durable and Efficient Gene Silencing In vivo by Hit-and-Run Epigenome Editing. *Nature* **2024**, *627* (8003), 416–423.

(222) Cheng, H.; Jeong, E.; Cho, S. W. Applications of Multiplexed CRISPR-Cas for Genome Engineering. *Experimental & Molecular Medicine* **2025**, *57* (7), 1373–1380.

(223) Phase 1b, Open-label Study of Tune-401 to Assess Safety, PK and PD in Adults With Chronic Hepatitis B. <https://www.clinicaltrials.gov/study/NCT06671093> (accessed 2025 October 23).

(224) Ueda, J.; Yamazaki, T.; Funakoshi, H. Toward the Development of Epigenome Editing-Based Therapeutics: Potentials and Challenges. *International Journal of Molecular Sciences* **2023**, *24* (5), 4778.

(225) Mazan-Mamczarz, K.; Ha, J.; De, S.; Sen, P. Single-Cell Analysis of the Transcriptome and Epigenome. *Methods Mol. Biol.* **2022**, *2399*, 21–60.

(226) Wen, L.; Tang, F. Single-cell Omics Sequencing Technologies: The Long-read Generation. *Trends in Genetics* **2026**, *42*, 46.

(227) Bonev, B.; Castelo-Branco, G.; Chen, F.; Codeluppi, S.; Corces, M. R.; Fan, J.; Heiman, M.; Harris, K.; Inoue, F.; Kellis, M.; Levine, A.; Lotfollahi, M.; Luo, C.; Maynard, K. R.; Nitzan, M.; Ramani, V.; Satijia, R.; Schirmer, L.; Shen, Y.; Sun, N.; Green, G. S.; Theis, F.; Wang, X.; Welch, J. D.; Gokce, O.; Konopka, G.; Liddelow, S.; Macosko, E.; Ali Bayraktar, O.; Habib, N.; Nowakowski, T. J. Opportunities and Challenges of Single-Cell and Spatially Resolved Genomics Methods for Neuroscience Discovery. *Nature Neuroscience* **2024**, *27* (12), 2292–2309.

(228) Ay-Berthomieu, A.-S. Beginner's Guide to Understanding Single-Cell ATAC-Seq. 2020. <https://www.activemotif.com/blog-single-cell-atac-seq> (accessed Feb 16, 2025).

(229) Tjoonk, N. Single-Cell ATAC-seq: The Basics. 2023. <https://www.scdiscoversies.com/blog/knowledge/single-cell-atac-seq-the-basics/> (accessed Feb 16, 2025).

- (230) Xiong, L.; Xu, K.; Tian, K.; Shao, Y.; Tang, L.; Gao, G.; Zhang, M.; Jiang, T.; Zhang, Q. C. SCALE Method for Single-cell ATAC-seq Analysis Via Latent Feature Extraction. *Nat. Commun.* **2019**, *10* (1), 4576.
- (231) Grosselin, K.; Durand, A.; Marsolier, J.; Poitou, A.; Marangoni, E.; Nemati, F.; Dahmani, A.; Lameiras, S.; Rey, F.; Frenoy, O.; Pousse, Y.; Reichen, M.; Woolfe, A.; Brenan, C.; Griffiths, A. D.; Vallot, C.; Gerard, A. High-throughput Single-cell ChIP-seq Identifies Heterogeneity of Chromatin States in Breast Cancer. *Nat. Genet.* **2019**, *51* (6), 1060–1066.
- (232) Rotem, A.; Ram, O.; Shores, N.; Sperling, R. A.; Goren, A.; Weitz, D. A.; Bernstein, B. E. Single-cell ChIP-seq Reveals Cell Subpopulations Defined by Chromatin State. *Nat. Biotechnol.* **2015**, *33* (11), 1165–1172.
- (233) Yu, X.; Zheng, G.; Xu, L.; Guo, W.; Chen, G.; Zhu, Y.; Li, T.; Rao, M.; Wang, L.; Cong, R.; Pei, H. MobiChIP: A Compatible Library Construction Method of Single-cell ChIP-seq Based Droplets. *Molecular Omics* **2024**, *21* (1), 32–37.
- (234) Ma, S.; Zhang, Y. Profiling Chromatin Regulatory Landscape: Insights into the Development of ChIP-seq and ATAC-seq. *Molecular Biomedicine* **2020**, *1* (1), 9.
- (235) Zhu, P.; Guo, H.; Ren, Y.; Hou, Y.; Dong, J.; Li, R.; Lian, Y.; Fan, X.; Hu, B.; Gao, Y.; Wang, X.; Wei, Y.; Liu, P.; Yan, J.; Ren, X.; Yuan, P.; Yuan, Y.; Yan, Z.; Wen, L.; Yan, L.; Qiao, J.; Tang, F. Single-cell DNA Methylome Sequencing of Human Preimplantation Embryos. *Nat. Genet.* **2018**, *50* (1), 12–19.
- (236) Cusanovich, D. A.; Reddington, J. P.; Garfield, D. A.; Daza, R. M.; Aghamirzaie, D.; Marco-Ferreres, R.; Pliner, H. A.; Christiansen, L.; Qiu, X.; Steemers, F. J.; Trapnell, C.; Shendure, J.; Furlong, E. E. M. The Cis-Regulatory Dynamics of Embryonic Development at Single-Cell Resolution. *Nature* **2018**, *555* (7697), 538–542.
- (237) Khavari, D. A.; Sen, G. L.; Rinn, J. L. DNA Methylation and Epigenetic Control of Cellular Differentiation. *Cell Cycle* **2010**, *9* (19), 3880–3883.
- (238) Pace, L.; Amigorena, S. Epigenetics of T Cell Fate Decision. *Current Opinion in Immunology* **2020**, *63*, 43–50.
- (239) Papalex, E.; Satija, R. Single-cell RNA Sequencing to Explore Immune Cell Heterogeneity. *Nature Reviews Immunology* **2018**, *18* (1), 35–45.
- (240) LaFave, L. M.; Savage, R. E.; Buenrostro, J. D. Single-Cell Epigenomics Reveals Mechanisms of Cancer Progression. *Annual Review of Cancer Biology* **2022**, *6*, 167–185.
- (241) Casado-Pelaez, M.; Bueno-Costa, A.; Esteller, M. Single Cell Cancer Epigenetics. *Trends in Cancer* **2022**, *8* (10), 820–838.
- (242) Eyler, C. E.; Matsunaga, H.; Hovestadt, V.; Vantine, S. J.; van Galen, P.; Bernstein, B. E. Single-Cell Lineage Analysis Reveals Genetic and Epigenetic Interplay in Glioblastoma Drug Resistance. *Genome Biology* **2020**, *21* (1), 174.
- (243) Fan, H.; Li, J.; Manuel, A. M.; Zhao, Z. Enzalutamide-Induced Signatures Revealed by Epigenetic Plasticity Using Single-Cell Multi-Omics Sequencing in Prostate Cancer. *Molecular Therapy Nucleic Acids* **2023**, *31*, 648–661.
- (244) Zifra, R. S.; Kim, C. N.; Ross, J. M.; Wilfert, A.; Turner, T. N.; Haeussler, M.; Casella, A. M.; Przytycki, P. F.; Keough, K. C.; Shin, D.; Bogdanoff, D.; Kreimer, A.; Pollard, K. S.; Ament, S. A.; Eichler, E. E.; Ahituv, N.; Nowakowski, T. J. Single-cell Epigenomics Reveals Mechanisms of Human Cortical Development. *Nature* **2021**, *598* (7879), 205–213.
- (245) Corces, M. R.; Shcherbina, A.; Kundu, S.; Gloudemans, M. J.; Fresard, L.; Granja, J. M.; Louie, B. H.; Eulalio, T.; Shams, S.; Bagdatli, S. T.; Mumbach, M. R.; Liu, B.; Montine, K. S.; Greenleaf, W. J.; Kundaje, A.; Montgomery, S. B.; Chang, H. Y.; Montine, T. J. Single-Cell Epigenomic Analyses Implicate Candidate Causal Variants at Inherited Risk Loci for Alzheimer's and Parkinson's Diseases. *Nat. Genet.* **2020**, *52* (11), 1158–1168.
- (246) Chen, Y.; Zander, R.; Khatun, A.; Schauder, D. M.; Cui, W. Transcriptional and Epigenetic Regulation of Effector and Memory CD8 T Cell Differentiation. *Frontiers in Immunology* **2018**, *9*, 2826.
- (247) Gulati, G. S.; D'Silva, J. P.; Liu, Y.; Wang, L.; Newman, A. M. Profiling Cell Identity and Tissue Architecture with Single-cell and Spatial Transcriptomics. *Nat. Rev. Mol. Cell Biol.* **2025**, *26* (1), 11–31.
- (248) Ding, J.; Sharon, N.; Bar-Joseph, Z. Temporal Modelling Using Single-Cell Transcriptomics. *Nat. Rev. Genet.* **2022**, *23* (6), 355–368.
- (249) Bond, D. R.; Uddipto, K.; Enjeti, A. K.; Lee, H. J. Single-Cell Epigenomics in Cancer: Charting a Course to Clinical Impact. *Epigenomics* **2020**, *12* (13), 1139–1151.
- (250) Kong, S.; Li, R.; Tian, Y.; Zhang, Y.; Lu, Y.; Ou, Q.; Gao, P.; Li, K.; Zhang, Y. Single-cell Omics: A New Direction for Functional Genetic Research in Human Diseases and Animal Models. *Frontiers in Genetics* **2023**, *13*, 1100016.
- (251) Lu, T.; Ang, C. E.; Zhuang, X. Spatially Resolved Epigenomic Profiling of Single Cells in Complex Tissues. *Cell* **2022**, *185* (23), 4448–4464.e17.
- (252) Xu, J.; Liu, Y. A Guide to Visualizing the Spatial Epigenome with Super-resolution Microscopy. *FEBS Journal* **2019**, *286* (16), 3095–3109.
- (253) Milosavljevic, A. Emerging Patterns of Epigenomic Variation. *Trends in Genetics* **2011**, *27* (6), 242–250.
- (254) Chen, T. Y.; You, L.; Hardillo, J. A. U.; Chien, M. P. Spatial Transcriptomic Technologies. *Cells* **2023**, *12* (16), 2042.
- (255) Choi, S. W.; Friso, S. Epigenetics: A New Bridge between Nutrition and Health. *Advances in Nutrition* **2010**, *1* (1), 8–16.
- (256) Alegria-Torres, J. A.; Baccarelli, A.; Bollati, V. Epigenetics and lifestyle. *Epigenomics* **2011**, *3* (3), 267–277.
- (257) Tiffon, C. The Impact of Nutrition and Environmental Epigenetics on Human Health and Disease. *International Journal of Molecular Sciences* **2018**, *19* (11), 3425.
- (258) Klibaner-Schiff, E.; Simonin, E. M.; Akdis, C. A.; Cheong, A.; Johnson, M. M.; Karagas, M. R.; Kirsh, S.; Kline, O.; Mazumdar, M.; Oken, E.; Sampath, V.; Vogler, N.; Wang, X.; Nadeau, K. C. Environmental Exposures Influence Multigenerational Epigenetic Transmission. *Clinical Epigenetics* **2024**, *16* (1), 145.
- (259) Keenan, C. E.; Kelleher, R.; Gray, S. G. Chapter 9 - Environmental Pollution, Epigenetics, and Cancer. In *Epigenetic Cancer Therapy*, Second ed.; Gray, S. G., Ed.; Academic Press, 2023; pp 175–194.
- (260) Ghosh, S.; Dhar, S.; Bhattacharjee, S.; Bhattacharjee, P. Contribution of Environmental, Genetic and Epigenetic Factors to Obesity-related Metabolic Syndrome. *Nucleus* **2023**, *66*, 215–237.
- (261) Yu, G.; Su, Q.; Chen, Y.; Wu, L.; Wu, S.; Li, H. Epigenetics in Neurodegenerative Disorders Induced by Pesticides. *Genes and Environment* **2021**, *43* (1), 55.
- (262) Migliore, L.; Coppede, F. Gene-environment Interactions in Alzheimer Disease: The Emerging Role of Epigenetics. *Nature Reviews Neurology* **2022**, *18* (11), 643–660.
- (263) Perera, B. P. U.; Faulk, C.; Svoboda, L. K.; Goodrich, J. M.; Dolinoy, D. C. The Role of Environmental Exposures and the Epigenome in Health and Disease. *Environmental and Molecular Mutagenesis* **2020**, *61* (1), 176–192.
- (264) Gomez-Verjan, J. C.; Barrera-Vazquez, O. S.; Garcia-Velazquez, L.; Samper-Ternent, R.; Arroyo, P. Epigenetic Variations Due to Nutritional Status in Early-life and Its Later Impact on Aging and Disease. *Clinical Genetics* **2020**, *98* (4), 313–321.
- (265) Sharavanan, V. J.; Sivaramakrishnan, M.; Sivarajasekar, N.; Senthilrani, N.; Kothandan, R.; Dhakal, N.; Sivamani, S.; Show, P. L.; Awall, M. R.; Naushad, M. Pollutants Inducing Epigenetic Changes and Diseases. *Environmental Chemistry Letters* **2020**, *18*, 325–343.
- (266) Major, K. M.; DeCourten, B. M.; Li, J.; Britton, M.; Settles, M. L.; Mehinto, A. C.; Connon, R. E.; Brander, S. M. Early Life Exposure to Environmentally Relevant Levels of Endocrine Disruptors Drive Multigenerational and Transgenerational Epigenetic Changes in a Fish Model. *Frontiers in Marine Science* **2020**, *7*, 471.
- (267) Kim, S.; Thapar, I.; Brooks, B. W. Epigenetic Changes by Per- and Polyfluoroalkyl Substances (PFAS). *Environ. Pollut.* **2021**, *279*, 116929.

- (268) Ell, M. A.; Schiele, M. A.; Iovino, N.; Domschke, K. Epigenetics of Fear, Anxiety and Stress - Focus on Histone Modifications. *Current Neuropsycharmacology* **2024**, *22* (5), 843–865.
- (269) Gottschalk, M. G.; Domschke, K.; Schiele, M. A. Epigenetics Underlying Susceptibility and Resilience Relating to Daily Life Stress, Work Stress, and Socioeconomic Status. *Frontiers in Psychiatry* **2020**, *11*, 163.
- (270) Seiler, C. L.; Song, J. U. M.; Kotandeniya, D.; Chen, J.; Kono, T. J. Y.; Han, Q.; Colwell, M.; Auch, B.; Sarver, A. L.; Upadhyaya, P.; Ren, Y.; Faulk, C.; De Flora, S.; La Maestra, S.; Chen, Y.; Kassie, F.; Tretyakova, N. Y. Inhalation Exposure to Cigarette Smoke and Inflammatory Agents Induces Epigenetic Changes in the Lung. *Sci. Rep.* **2020**, *10* (1), 11290.
- (271) Luo, A.; Jung, J.; Longley, M.; Rosoff, D. B.; Charlet, K.; Muench, C.; Lee, J.; Hodgkinson, C. A.; Goldman, D.; Horvath, S.; Kaminsky, Z. A.; Lohoff, F. W. Epigenetic Aging is Accelerated in Alcohol Use Disorder and Regulated by Genetic Variation in APOL2. *Neuropsychopharmacology* **2020**, *45* (2), 327–336.
- (272) Fox, F. A. U.; Liu, D.; Breteler, M. M. B.; Aziz, N. A. Physical Activity is Associated with Slower Epigenetic Ageing-Findings from the Rhineland Study. *Aging Cell* **2023**, *22* (6), No. e13828.
- (273) Nilsson, E. E.; Ben Maamar, M.; Skinner, M. K. Role of Epigenetic Transgenerational Inheritance in Generational Toxicology. *Environmental Epigenetics* **2022**, *8* (1), dvac001.
- (274) Moelling, K. Epigenetics and Transgenerational Inheritance. *Journal of Physiology* **2024**, *602* (11), 2537–2545.
- (275) Cao, S.; Chen, Z. J. Transgenerational Epigenetic Inheritance During Plant Evolution and Breeding. *Trends in Plant Science* **2024**, *29* (11), 1203–1223.
- (276) Thompson, R. P.; Nilsson, E.; Skinner, M. K. Environmental Epigenetics and Epigenetic Inheritance in Domestic Farm Animals. *Animal Reproduction Science* **2020**, *220*, 106316.
- (277) Horsthemke, B. A Critical View on Transgenerational Epigenetic Inheritance in Humans. *Nat. Commun.* **2018**, *9* (1), 2973.
- (278) Tuscher, J. J.; Day, J. J. Multigenerational Epigenetic Inheritance: One Step Forward, Two Generations Back. *Neurobiology of Disease* **2019**, *132*, 104591.
- (279) Niu, Z.; Mohazzab-Hosseini, S.; Breton, C. V. Transgenerational Epigenetic Inheritance: Perspectives and Challenges. *Journal of Allergy and Clinical Immunology* **2023**, *151* (6), 1474–1476.
- (280) Fallet, M.; Blanc, M.; Di Criscio, M.; Antczak, P.; Engwall, M.; Guerrero Bosagna, C.; Ruegg, J.; Keiter, S. H. Present and Future Challenges for the Investigation of Transgenerational Epigenetic Inheritance. *Environ. Int.* **2023**, *172*, 107776. From
- (281) Ben Maamar, M.; Nilsson, E. E.; Skinner, M. K. Epigenetic Transgenerational Inheritance, Gametogenesis and Germline Development. *Biol. Reprod.* **2021**, *105* (3), 570–592.
- (282) Tisato, V.; D'Aversa, E.; Salvatori, F.; Sbracia, M.; Peluso, G.; Scarpellini, F.; Gemmati, D. Epigenetic Mechanisms in Maternal-fetal Crosstalk: Inter- and Trans-generational Inheritance. *Epigenomics* **2025**, *17*, 1303.
- (283) McSwiggin, H.; Magalhaes, R.; Nilsson, E. E.; Yan, W.; Skinner, M. K. Epigenetic Transgenerational Inheritance of Toxicant Exposure-specific Non-coding RNA in Sperm. *Environmental Epigenetics* **2024**, *10* (1), dvae014.
- (284) Zimmer, C. The Famine Ended 70 Years Ago, But Dutch Genes Still Bear Scars. *New York Times* (Manhattan, U.S.), 2018. <https://www.nytimes.com/2018/01/31/science/dutch-famine-genes.html>.
- (285) Conti, G.; Poupakis, S.; Ekamper, P.; Bijwaard, G. E.; Lumey, L. H. Severe Prenatal Shocks and Adolescent Health: Evidence from the Dutch Hunger Winter. *Economics & Human Biology* **2024**, *53*, 101372.
- (286) Anway, M. D.; Skinner, M. K. Epigenetic Transgenerational Actions of Endocrine Disruptors. *Endocrinology* **2006**, *147* (6), S43–S49.
- (287) Brehm, E.; Flaws, J. A. Transgenerational Effects of Endocrine-Disrupting Chemicals on Male and Female Reproduction. *Endocrinology* **2019**, *160* (6), 1421–1435.
- (288) Nilsson, E.; King, S. E.; McBirney, M.; Kubsad, D.; Pappalardo, M.; Beck, D.; Sadler-Riggleman, I.; Skinner, M. K. Vinclozolin Induced Epigenetic Transgenerational Inheritance of Pathologies and Sperm Epimutation Biomarkers for Specific Diseases. *PLoS One* **2018**, *13* (8), No. e0202662.
- (289) Kitaba, N. T.; Knudsen, G. T. M.; Johannessen, A.; Rezwani, F. I.; Malinovski, A.; Oudin, A.; Benediksdottir, B.; Martino, D.; Gonzalez, F. J. C.; Gomez, L. P.; Holm, M.; Jogi, N. O.; Dharmage, S. C.; Skulstad, S. M.; Watkins, S. H.; Suderman, M.; Gomez-Real, F.; Schlunssen, V.; Svanes, C.; Holloway, J. W. Fathers' Preconception Smoking and Offspring DNA Methylation. *Clinical Epigenetics* **2023**, *15* (1), 131.
- (290) Adeline Dorothy, P. D.; Rajan, K. E. Prenatal Maternal Life Adversity Impacts on Learning and Memory in Offspring: Implication to Transgenerational Epigenetic Inheritance. *Frontiers in Neuroscience* **2025**, *19*, 1518046.
- (291) Nilsson, E. E.; Sadler-Riggleman, I.; Skinner, M. K. Environmentally Induced Epigenetic Transgenerational Inheritance of Disease. *Environmental Epigenetics* **2018**, *4* (2), dvv016.
- (292) King, S. E.; Skinner, M. K. Epigenetic Transgenerational Inheritance of Obesity Susceptibility. *Trends in Endocrinology & Metabolism* **2020**, *31* (7), 478–494.
- (293) Panera, N.; Mandato, C.; Crudele, A.; Bertrando, S.; Vajro, P.; Alisi, A. Genetics, Epigenetics and Transgenerational Transmission of Obesity in Children. *Frontiers in Endocrinology* **2022**, *13*, 1006008.
- (294) Franzago, M.; Pilenzi, L.; Di Rado, S.; Vitacolonna, E.; Stuppia, L. The Epigenetic Aging, Obesity, and Lifestyle. *Frontiers in Cell and Developmental Biology* **2022**, *10*, 985274.
- (295) Wang, K.; Liu, H.; Hu, Q.; Wang, L.; Liu, J.; Zheng, Z.; Zhang, W.; Ren, J.; Zhu, F.; Liu, G. H. Epigenetic Regulation of Aging: Implications for Interventions of Aging and Diseases. *Signal Transduction and Targeted Therapy* **2022**, *7* (1), 374.
- (296) Feinberg, A. P.; Levchenko, A. Epigenetics as a Mediator of Plasticity in Cancer. *Science* **2023**, *379* (6632), No. eaaw3835.
- (297) Shi, Y.; Zhang, H.; Huang, S.; Yin, L.; Wang, F.; Luo, P.; Huang, H. Epigenetic Regulation in Cardiovascular Disease: Mechanisms and Advances in Clinical Trials. *Signal Transduction and Targeted Therapy* **2022**, *7* (1), 200.
- (298) Berson, A.; Nativio, R.; Berger, S. L.; Bonini, N. M. Epigenetic Regulation in Neurodegenerative Diseases. *Trends in Neurosciences* **2018**, *41* (9), 587–598.
- (299) Mohd Murshid, N.; Aminullah Lubis, F.; Makpol, S. Epigenetic Changes and Its Intervention in Age-Related Neurodegenerative Diseases. *Cellular and Molecular Neurobiology* **2022**, *42* (3), 577–595.
- (300) Zhang, L.; Liu, Y.; Lu, Y.; Wang, G. Targeting Epigenetics as a Promising Therapeutic Strategy for Treatment of Neurodegenerative Diseases. *Biochem. Pharmacol.* **2022**, *206*, 115295.
- (301) Lai, C. Q.; Parnell, L. D.; Smith, C. E.; Guo, T.; Sayols-Baixeras, S.; Aslibekyan, S.; Tiwari, H. K.; Irvin, M. R.; Bender, C.; Fei, D.; Hidalgo, B.; Hopkins, P. N.; Absher, D. M.; Province, M. A.; Elosua, R.; Arnett, D. K.; Ordovas, J. M. Carbohydrate and Fat Intake Associated with Risk of Metabolic Diseases Through Epigenetics of CPT1A. *American Journal of Clinical Nutrition* **2020**, *112* (5), 1200–1211.
- (302) Ling, C.; Bacos, K.; Ronn, T. Epigenetics of Type 2 Diabetes Mellitus and Weight Change - A Tool for Precision Medicine? *Nature Reviews Endocrinology* **2022**, *18* (7), 433–448.
- (303) Samanta, S.; Rajasingh, S.; Cao, T.; Dawn, B.; Rajasingh, J. Epigenetic Dysfunctional Diseases and Therapy for Infection and Inflammation. *Biochimica et Biophysica Acta - Molecular Basis of Disease* **2017**, *1863* (2), 518–528.
- (304) Farsetti, A.; Illi, B.; Gaetano, C. How Epigenetics Impacts on Human Diseases. *European Journal of Internal Medicine* **2023**, *114*, 15–22.
- (305) Lu, Y.; Chan, Y. T.; Tan, H. Y.; Li, S.; Wang, N.; Feng, Y. Epigenetic Regulation in Human Cancer: The Potential Role of Epigenetic Regulation in Cancer Therapy. *Molecular Cancer* **2020**, *19* (1), 79.
- (306) Schuh, A. C.; Dohner, H.; Pleyer, L.; Seymour, J. F.; Fenaux, P.; Dombret, H. Azacitidine in Adult Patients with Acute Myeloid Leukemia. *Critical Reviews in Oncology/Hematology* **2017**, *116*, 159–177.

- (307) DiNardo, C. D.; Jonas, B. A.; Pullarkat, V.; Thirman, M. J.; Garcia, J. S.; Wei, A. H.; Konopleva, M.; Dohner, H.; Letai, A.; Fenaux, P.; Koller, E.; Havelange, V.; Leber, B.; Esteve, J.; Wang, J.; Pejsa, V.; Hajek, R.; Porkka, K.; Illes, A.; Lavie, D.; Lemoli, R. M.; Yamamoto, K.; Yoon, S. S.; Jang, J. H.; Yeh, S. P.; Turgut, M.; Hong, W. J.; Zhou, Y.; Potluri, J.; Pratz, K. W. Azacitidine and Venetoclax in Previously Untreated Acute Myeloid Leukemia. *New England Journal of Medicine* **2020**, *383* (7), 617–629.
- (308) Malik, P.; Cashen, A. F. Decitabine in the Treatment of Acute Myeloid Leukemia in Elderly Patients. *Cancer Management and Research* **2014**, *6*, 53–61.
- (309) Efficace, F.; Kicinski, M.; Coens, C.; Suciu, S.; van der Velden, W.; Noppeney, R.; Chantepie, S.; Griskevicius, L.; Neubauer, A.; Audisio, E.; Luppi, M.; Fuhrmann, S.; Foa, R.; Crysandt, M.; Gaidano, G.; Vrhovac, R.; Venditti, A.; Posthuma, E. F. M.; Candoni, A.; Baron, F.; Legrand, O.; Mengarelli, A.; Fazi, P.; Vignetti, M.; Giraut, A.; Wijermans, P. W.; Huls, G.; Lubbert, M. Decitabine in Older Patients with AML: Quality of Life Results of the EORTC-GIMEMA-GMDS-SG Randomized Phase 3 Trial. *Blood* **2024**, *144* (5), 541–551.
- (310) Mann, B. S.; Johnson, J. R.; Cohen, M. H.; Justice, R.; Pazdur, R. FDA Approval Summary: Vorinostat for Treatment of Advanced Primary Cutaneous T-cell Lymphoma. *Oncologist* **2007**, *12* (10), 1247–1252.
- (311) Nishiyama, A.; Nakanishi, M. Navigating the DNA Methylation Landscape of Cancer. *Trends in Genetics* **2021**, *37* (11), 1012–1027.
- (312) Ko, E.; Kim, Y.; Kim, S. J.; Joh, J. W.; Song, S.; Park, C. K.; Park, J.; Kim, D. H. Promoter Hypermethylation of the p16 Gene is Associated with Poor Prognosis in Recurrent Early-stage Hepatocellular Carcinoma. *Cancer Epidemiology, Biomarkers & Prevention* **2008**, *17* (9), 2260–2267.
- (313) Ye, X.; Mo, M.; Xu, S.; Yang, Q.; Wu, M.; Zhang, J.; Chen, B.; Li, J.; Zhong, Y.; Huang, Q.; Cai, C. The Hypermethylation of p16 Gene Exon 1 and Exon 2: Potential Biomarkers for Colorectal Cancer and are Associated with Cancer Pathological Staging. *BMC Cancer* **2018**, *18* (1), 1023.
- (314) Esteller, M.; Silva, J. M.; Dominguez, G.; Bonilla, F.; Matias-Guiu, X.; Lerma, E.; Bussaglia, E.; Prat, J.; Harkes, I. C.; Repasky, E. A.; Gabrielson, E.; Schutte, M.; Baylin, S. B.; Herman, J. G. Promoter Hypermethylation and BRCA1 Inactivation in Sporadic Breast and Ovarian Tumors. *Journal of the National Cancer Institute* **2000**, *92* (7), 564–569.
- (315) Lonning, P. E.; Nikolaienko, O.; Pan, K.; Kurian, A. W.; Eikesdal, H. P.; Pettinger, M.; Anderson, G. L.; Prentice, R. L.; Chlebowski, R. T.; Knappekog, S. Constitutional BRCA1 Methylation and Risk of Incident Triple-Negative Breast Cancer and High-grade Serous Ovarian Cancer. *JAMA Oncology* **2022**, *8* (11), 1579–1587.
- (316) Cai, Y.; Zhang, Y.; Loh, Y. P.; Tng, J. Q.; Lim, M. C.; Cao, Z.; Raju, A.; Lieberman Aiden, E.; Li, S.; Manikandan, L.; Tergaonkar, V.; Tucker-Kellogg, G.; Fullwood, M. J. H3K27me3-Rich Genomic Regions can Function as Silencers to Repress Gene Expression Via Chromatin Interactions. *Nat. Commun.* **2021**, *12* (1), 719.
- (317) Liang, Y.; Liu, M.; Liu, B.; Ziman, B.; Peng, G.; Mao, Q.; Wang, X.; Jiang, L.; Lin, D. C.; Zheng, Y. Comprehensive Analysis of H3K27me3 LOCKs Under Different DNA Methylation Contexts Reveal Epigenetic Redistribution in Tumorigenesis. *Epigenetics Chromatin* **2025**, *18* (1), 6.
- (318) Wei, J. W.; Huang, K.; Yang, C.; Kang, C. S. Non-coding RNAs as Regulators in Epigenetics. *Oncol. Rep.* **2017**, *37* (1), 3–9.
- (319) Zhu, S.; Wu, H.; Wu, F.; Nie, D.; Sheng, S.; Mo, Y. Y. MicroRNA-21 Targets Tumor Suppressor Genes in Invasion and Metastasis. *Cell Research* **2008**, *18* (3), 350–359.
- (320) Qin, X.; Yan, L.; Zhao, X.; Li, C.; Fu, Y. microRNA-21 Overexpression Contributes to Cell Proliferation by Targeting PTEN in Endometrioid Endometrial Cancer. *Oncology Letters* **2012**, *4* (6), 1290–1296.
- (321) Nosaka, K.; Maeda, M.; Tamiya, S.; Sakai, T.; Mitsuya, H.; Matsuoka, M. Increasing Methylation of the CDKN2A Gene Is Associated with the Progression of Adult T-Cell Leukemia. *Cancer Res.* **2000**, *60*, 1043–1048.
- (322) Wijetunga, N. A.; Belbin, T. J.; Burk, R. D.; Whitney, K.; Abadi, M.; Grealley, J. M.; Einstein, M. H.; Schlecht, N. F. Novel Epigenetic Changes in CDKN2A are Associated with Progression of Cervical Intraepithelial Neoplasia. *Gynecologic Oncology* **2016**, *142* (3), 566–573.
- (323) Suehiro, Y.; Wong, C. W.; Chirieac, L. R.; Kondo, Y.; Shen, L.; Webb, C. R.; Chan, Y. W.; Chan, A. S.; Chan, T. L.; Wu, T. T.; Rashid, A.; Hamanaka, Y.; Hinoda, Y.; Shannon, R. L.; Wang, X.; Morris, J.; Issa, J. P.; Yuen, S. T.; Leung, S. Y.; Hamilton, S. R. Epigenetic-genetic Interactions in the APC/WNT, RAS/RAF, and P53 Pathways in Colorectal Carcinoma. *Clin. Cancer Res.* **2008**, *14* (9), 2560–2569.
- (324) Zhu, L.; Li, X.; Yuan, Y.; Dong, C.; Yang, M. APC Promoter Methylation in Gastrointestinal Cancer. *Frontiers in Oncology* **2021**, *11*, 653222.
- (325) Tew, B. Y.; Durand, J. K.; Bryant, K. L.; Hayes, T. K.; Peng, S.; Tran, N. L.; Gooden, G. C.; Buckley, D. N.; Der, C. J.; Baldwin, A. S.; Salhia, B. Genome-wide DNA Methylation Analysis of KRAS Mutant Cell Lines. *Sci. Rep.* **2020**, *10* (1), 10149.
- (326) Noroozi, R.; Ghafouri-Fard, S.; Pisarek, A.; Rudnicka, J.; Spolnicka, M.; Branicki, W.; Taheri, M.; Pospiech, E. DNA Methylation-based Age Clocks: From Age Prediction to Age Reversion. *Ageing Research Reviews* **2021**, *68*, 101314.
- (327) Varshavsky, M.; Harari, G.; Glaser, B.; Dor, Y.; Shemer, R.; Kaplan, T. Accurate Age Prediction from Blood Using a Small Set of DNA Methylation Sites and a Cohort-based Machine Learning Algorithm. *Cell Reports Methods* **2023**, *3* (9), 100567.
- (328) Tenchov, R.; Sasso, J. M.; Wang, X.; Zhou, Q. A. Aging Hallmarks and Progression and Age-Related Diseases: A Landscape View of Research Advancement. *ACS Chem. Neurosci.* **2024**, *15* (1), 1–30.
- (329) Li, A.; Koch, Z.; Ideker, T. Epigenetic Aging: Biological Age Prediction and Informing a Mechanistic Theory of Aging. *Journal of Internal Medicine* **2022**, *292* (5), 733–744.
- (330) Bell, C. G.; Lowe, R.; Adams, P. D.; Baccarelli, A. A.; Beck, S.; Bell, J. T.; Christensen, B. C.; Gladyshev, V. N.; Heijmans, B. T.; Horvath, S.; Ideker, T.; Issa, J. J.; Kelsey, K. T.; Marioni, R. E.; Reik, W.; Relton, C. L.; Schalkwyk, L. C.; Teschendorff, A. E.; Wagner, W.; Zhang, K.; Rakyan, V. K. DNA Methylation Aging Clocks: Challenges and Recommendations. *Genome Biology* **2019**, *20* (1), 249.
- (331) Duan, R.; Fu, Q.; Sun, Y.; Li, Q. Epigenetic Clock: A Promising Biomarker and Practical Tool in Aging. *Ageing Research Reviews* **2022**, *81*, 101743.
- (332) Horvath, S. DNA Methylation Age of Human Tissues and Cell Types. *Genome Biology* **2013**, *14* (10), R115.
- (333) Hannum, G.; Guinney, J.; Zhao, L.; Zhang, L.; Hughes, G.; Sada, S.; Klotzle, B.; Bibikova, M.; Fan, J. B.; Gao, Y.; Deconde, R.; Chen, M.; Rajapakse, I.; Friend, S.; Ideker, T.; Zhang, K. Genome-wide Methylation Profiles Reveal Quantitative Views of Human Aging Rates. *Mol. Cell* **2013**, *49* (2), 359–367.
- (334) Lu, A. T.; Quach, A.; Wilson, J. G.; Reiner, A. P.; Aviv, A.; Raj, K.; Hou, L.; Baccarelli, A. A.; Li, Y.; Stewart, J. D.; Whitsel, E. A.; Assimes, T. L.; Ferrucci, L.; Horvath, S. DNA Methylation GrimAge Strongly Predicts Lifespan and Healthspan. *Aging (Albany NY)* **2019**, *11* (2), 303–327.
- (335) Levine, M. E.; Lu, A. T.; Quach, A.; Chen, B. H.; Assimes, T. L.; Bandinelli, S.; Hou, L.; Baccarelli, A. A.; Stewart, J. D.; Li, Y.; Whitsel, E. A.; Wilson, J. G.; Reiner, A. P.; Aviv, A.; Lohman, K.; Liu, Y.; Ferrucci, L.; Horvath, S. An Epigenetic Biomarker of Aging for Lifespan and Healthspan. *Aging (Albany NY)* **2018**, *10* (4), 573–591.
- (336) McCrory, C.; Fiorito, G.; Hernandez, B.; Polidoro, S.; O'Halloran, A. M.; Hever, A.; Ni Cheallaigh, C.; Lu, A. T.; Horvath, S.; Vineis, P.; Kenny, R. A. GrimAge Outperforms Other Epigenetic Clocks in the Prediction of Age-Related Clinical Phenotypes and All-Cause Mortality. *Journals of Gerontology. Series A, Biological Sciences and Medical Sciences* **2021**, *76* (5), 741–749.
- (337) Protsenko, E.; Yang, R.; Nier, B.; Reus, V.; Hammamieh, R.; Rampersaud, R.; Wu, G. W. Y.; Hough, C. M.; Epel, E.; Prather, A. A.; Jett, M.; Gautam, A.; Mellon, S. H.; Wolkowitz, O. M. GrimAge, an

Epigenetic Predictor of Mortality, is Accelerated in Major Depressive Disorder. *Translational Psychiatry* **2021**, *11* (1), 193.

(338) Levy, J. J.; Diallo, A. B.; Saldias Montivero, M. K.; Gabbita, S.; Salas, L. A.; Christensen, B. C. Insights to Aging Prediction with AI Based Epigenetic Clocks. *Epigenomics* **2025**, *17* (1), 49–57.

(339) Tenchov, R.; Sasso, J. M.; Wang, X.; Zhou, Q. A. Antiaging Strategies and Remedies: A Landscape of Research Progress and Promise. *ACS Chem. Neurosci.* **2024**, *15* (3), 408–446.

(340) Rajado, A. T.; Silva, N.; Esteves, F.; Brito, D.; Binnie, A.; Araujo, I. M.; Nobrega, C.; Braganca, J.; Castelo-Branco, P.; Consortium, A. S. How Can We Modulate Aging Through Nutrition and Physical Exercise? An Epigenetic Approach. *Aging (Albany NY)* **2023**, *15* (8), 3191–3217.

(341) Ryan, C. P. Epigenetic Clocks: Theory and Applications in Human Biology. *American Journal of Human Biology* **2021**, *33* (3), No. e23488.

(342) Ecker, S.; Beck, S. The Epigenetic Clock: A Molecular Crystal Ball for Human Aging? *Aging (Albany NY)* **2019**, *11* (2), 833–835.

(343) Hao, Y.; Han, K.; Wang, T.; Yu, J.; Ding, H.; Dao, F. Exploring the Potential of Epigenetic Clocks in Aging Research. *Methods* **2024**, *231*, 37–44.

(344) Qazi, T. J.; Quan, Z.; Mir, A.; Qing, H. Epigenetics in Alzheimer's Disease: Perspective of DNA Methylation. *Molecular Neurobiology* **2018**, *55* (2), 1026–1044.

(345) Song, H.; Chen, J.; Huang, J.; Sun, P.; Liu, Y.; Xu, L.; Wei, C.; Mu, X.; Lu, X.; Wang, W.; Zhang, N.; Shang, M.; Mo, M.; Zhang, W.; Zhao, H.; Han, F. Epigenetic Modification in Parkinson's Disease. *Frontiers in Cell and Developmental Biology* **2023**, *11*, 1123621.

(346) Masala, A.; Sanna, S.; Esposito, S.; Rassu, M.; Galioto, M.; Zinellu, A.; Carru, C.; Carri, M. T.; Iaccarino, C.; Crosio, C. Epigenetic Changes Associated with the Expression of Amyotrophic Lateral Sclerosis (ALS) Causing Genes. *Neuroscience* **2018**, *390*, 1–11.

(347) Bennett, S. A.; Tanaz, R.; Cobos, S. N.; Torrente, M. P. Epigenetics in Amyotrophic Lateral Sclerosis: A Role for Histone Post-Translational Modifications in Neurodegenerative Disease. *Translational Research* **2019**, *204*, 19–30.

(348) Noches, V.; Campos-Melo, D.; Droppelmann, C. A.; Strong, M. J. Epigenetics in the Formation of Pathological Aggregates in Amyotrophic Lateral Sclerosis. *Frontiers in Molecular Neuroscience* **2024**, *17*, 1417961.

(349) Park, C.; Rosenblat, J. D.; Brietzke, E.; Pan, Z.; Lee, Y.; Cao, B.; Zuckerman, H.; Kalantarova, A.; McIntyre, R. S. Stress, Epigenetics and Depression: A Systematic Review. *Neuroscience & Biobehavioral Reviews* **2019**, *102*, 139–152.

(350) Chen, Q.; Li, D.; Jin, W.; Shi, Y.; Li, Z.; Ma, P.; Sun, J.; Chen, S.; Li, P.; Lin, P. Research Progress on the Correlation Between Epigenetics and Schizophrenia. *Frontiers in Neuroscience* **2021**, *15*, 688727.

(351) Herrera-Espejo, S.; Santos-Zorroza, B.; Alvarez-Gonzalez, P.; Lopez-Lopez, E.; Garcia-Orad, A. A Systematic Review of MicroRNA Expression as Biomarker of Late-Onset Alzheimer's Disease. *Molecular Neurobiology* **2019**, *56* (12), 8376–8391.

(352) Mei, Z.; Liu, J.; Schroeder, J. P.; Weinshenker, D.; Duong, D. M.; Seyfried, N. T.; Li, Y.; Jin, P.; Wingo, A. P.; Wingo, T. S. Lowering Hippocampal miR-29a Expression Slows Cognitive Decline and Reduces Beta-Amyloid Deposition in 5xFAD Mice. *Molecular Neurobiology* **2024**, *61* (6), 3343–3356.

(353) Raia, T.; Cavallaro, R. A.; Borges, L. D. F.; Cinti, S.; Bizzarri, M.; Ferrer, I.; Lucarelli, M.; Fuso, A. One-carbon Metabolism Modulates miR-29a-DNA Methylation Crosstalk in Alzheimer's Disease. *Alzheimer's & Dementia* **2025**, *21* (9), No. e70703.

(354) Cosin-Tomas, M.; Antonell, A.; Llado, A.; Alcolea, D.; Fortea, J.; Ezquerro, M.; Lleo, A.; Marti, M. J.; Pallas, M.; Sanchez-Valle, R.; Molinuevo, J. L.; Sanfeliu, C.; Kaliman, P. Plasma miR-34a-5p and miR-545-3p as Early Biomarkers of Alzheimer's Disease: Potential and Limitations. *Molecular Neurobiology* **2017**, *54* (7), 5550–5562.

(355) Du, X.; Guan, L.; Chen, C.; Wang, X.; Geng, X. Long-Term Exposure to PM(2.5) Exacerbates Dopaminergic Neuronal Loss

Through CpG Hypermethylation Induced Down-Regulation of PINK1 and DJ-1 Genes. *Sci. Rep.* **2025**, *15* (1), 10778.

(356) Pavlou, M. A. S.; Pinho, R.; Paiva, I.; Outeiro, T. F. The Yin and Yang of Alpha-synuclein-associated Epigenetics in Parkinson's Disease. *Brain* **2016**, *140* (4), 878–886.

(357) Toker, L.; Tran, G. T.; Sundaresan, J.; Tysnes, O. B.; Alves, G.; Haugarvoll, K.; Nido, G. S.; Dolle, C.; Tzoulis, C. Genome-wide Histone Acetylation Analysis Reveals Altered Transcriptional Regulation in the Parkinson's Disease Brain. *Molecular Neurodegeneration* **2021**, *16* (1), 31.

(358) Miranda-Morales, E.; Meier, K.; Sandoval-Carrillo, A.; Salas-Pacheco, J.; Vazquez-Cardenas, P.; Arias-Carrion, O. Implications of DNA Methylation in Parkinson's Disease. *Frontiers in Molecular Neuroscience* **2017**, *10*, 225.

(359) Lord, C.; Bruga, T. S.; Charman, T.; Cusack, J.; Dumas, G.; Frazier, T.; Jones, E. J. H.; Jones, R. M.; Pickles, A.; State, M. W.; Taylor, J. L.; Veenstra-VanderWeele, J. Autism Spectrum Disorder. *Nature Reviews Disease Primers* **2020**, *6* (1), 5.

(360) Zhu, L.; Wang, X.; Li, X. L.; Towers, A.; Cao, X.; Wang, P.; Bowman, R.; Yang, H.; Goldstein, J.; Li, Y. J.; Jiang, Y. H. Epigenetic Dysregulation of SHANK3 in Brain Tissues from Individuals with Autism Spectrum Disorders. *Hum. Mol. Genet.* **2014**, *23* (6), 1563–1578.

(361) Tremblay, M. W.; Jiang, Y. H. DNA Methylation and Susceptibility to Autism Spectrum Disorder. *Annual Review of Medicine* **2019**, *70*, 151–166.

(362) Mouat, J. S.; LaSalle, J. M. The Promise of DNA Methylation in Understanding Multigenerational Factors in Autism Spectrum Disorders. *Frontiers in Genetics* **2022**, *13*, 831221.

(363) Sun, W.; Poschmann, J.; Cruz-Herrera Del Rosario, R.; Parikshak, N. N.; Hajan, H. S.; Kumar, V.; Ramasamy, R.; Belgard, T. G.; Elangovan, B.; Wong, C. C. Y.; Mill, J.; Geschwind, D. H.; Prabhakar, S. Histone Acetylome-wide Association Study of Autism Spectrum Disorder. *Cell* **2016**, *167* (5), 1385–1397.e11.

(364) Tseng, C. J.; McDougall, C. J.; Hooker, J. M.; Zurcher, N. R. Epigenetics of Autism Spectrum Disorder: Histone Deacetylases. *Biol. Psychiatry* **2022**, *91* (11), 922–933.

(365) Choi, C. S.; Gonzales, E. L.; Kim, K. C.; Yang, S. M.; Kim, J. W.; Mabunga, D. F.; Cheong, J. H.; Han, S. H.; Bahn, G. H.; Shin, C. Y. The Transgenerational Inheritance of Autism-Like Phenotypes in Mice Exposed to Valproic Acid During Pregnancy. *Sci. Rep.* **2016**, *6*, 36250.

(366) Cao, D. F.; Zhou, X. Y.; Guo, Q.; Xiang, M. Y.; Bao, M. H.; He, B. S.; Mao, X. Y. Unveiling the Role of Histone Deacetylases in Neurological Diseases: Focus on Epilepsy. *Biomarker Research* **2024**, *12* (1), 142.

(367) Xie, G.; Chen, H.; He, C.; Hu, S.; Xiao, X.; Luo, Q. The Dysregulation of miRNAs in Epilepsy and Their Regulatory Role in Inflammation and Apoptosis. *Functional & Integrative Genomics* **2023**, *23* (3), 287.

(368) Good, K. V.; Vincent, J. B.; Ausio, J. MeCP2: The Genetic Driver of Rett Syndrome Epigenetics. *Frontiers in Genetics* **2021**, *12*, 620859.

(369) Jin, P.; Warren, S. T. Understanding the Molecular Basis of Fragile X Syndrome. *Hum. Mol. Genet.* **2000**, *9* (6), 901–908.

(370) Liu, X. S.; Wu, H.; Krzisch, M.; Wu, X.; Graef, J.; Muffat, J.; Hnisz, D.; Li, C. H.; Yuan, B.; Xu, C.; Li, Y.; Vershkov, D.; Cacace, A.; Young, R. A.; Jaenisch, R. Rescue of Fragile X Syndrome Neurons by DNA Methylation Editing of the FMR1 Gene. *Cell* **2018**, *172* (5), 979–992.e6.

(371) Skinner, M. K. Epigenetic Biomarkers for Disease Susceptibility and Preventative Medicine. *Cell Metabolism* **2024**, *36* (2), 263–277.

(372) Zheleznyakova, G. Y.; Cao, H.; Schioth, H. B. BDNF DNA Methylation Changes as a Biomarker of Psychiatric Disorders: Literature Review and Open Access Database Analysis. *Behavioral and Brain Functions* **2016**, *12* (1), 17.

(373) Alece Arantes Moreno, I.; Rodrigues de Oliveira, D.; Ribeiro Borcoi, A.; Fungaro Rissatti, L.; Vitorino Freitas, F.; Arantes, L.; Oliveira Mendes, S.; Dos Santos Vieira, T.; Risse Quaioto, B.; Cerbino Doblas, P.; Sgrancio Olinda, A.; Ribeiro Cunha, E.; Gasparini Dos

Santos, J.; Assis Pinheiro, J.; Pereira Sorroche, B.; Madeira Alvares da Silva, A. Methylation of BDNF Gene in Association with Episodic Memory in Women. *Frontiers in Neuroscience* **2023**, *17*, 1092406.

(374) Gao, S.; Cheng, J.; Li, G.; Sun, T.; Xu, Y.; Wang, Y.; Du, X.; Xu, G.; Duan, S. Catechol-O-methyltransferase Gene Promoter Methylation as a Peripheral Biomarker in Male Schizophrenia. *European Psychiatry* **2017**, *44*, 39–46.

(375) Liu, S.; Fan, M.; Zheng, Q.; Hao, S.; Yang, L.; Xia, Q.; Qi, C.; Ge, J. MicroRNAs in Alzheimer's Disease: Potential Diagnostic Markers and Therapeutic Targets. *Biomedicine & Pharmacotherapy* **2022**, *148*, 112681.

(376) Zhang, M.; Bian, Z. Alzheimer's Disease and microRNA-132: A Widespread Pathological Factor and Potential Therapeutic Target. *Frontiers in Neuroscience* **2021**, *15*, 687973.

(377) Sbriscia, M.; Spadoni, T.; Ambrogini, P.; Guescini, M.; Agostini, R.; Graciotti, L.; Piacenza, F.; Giuli, C.; Cecati, M.; Bonfigli, A. R.; Vaiasicca, S.; Pagliarini, M.; Rusanova, I.; Fazioli, F.; Sabbatinelli, J.; Albertini, M. C.; Olivieri, F.; Giuliani, A. miR-132–3p is Down-regulated in Plasma and CD171(+) Extracellular Vesicles Isolated from Patients with Mild Alzheimer's Disease. *Mechanisms of Ageing and Development* **2025**, *225*, 112063.

(378) Angelopoulou, E.; Paudel, Y. N.; Piperi, C. miR-124 and Parkinson's disease: A Biomarker with Therapeutic Potential. *Pharmacol. Res.* **2019**, *150*, 104515.

(379) Raghubeer, S. The Influence of Epigenetics and Inflammation on Cardiometabolic Risks. *Seminars in Cell & Developmental Biology* **2024**, *154* (Pt C), 175–184.

(380) Gevaert, A. B.; Wood, N.; Boen, J. R. A.; Davos, C. H.; Hansen, D.; Hanssen, H.; Krenning, G.; Moholdt, T.; Osto, E.; Paneni, F.; Pedretti, R. F. E.; Plosch, T.; Simonenko, M.; Bowen, T. S. Epigenetics in the Primary and Secondary Prevention of Cardiovascular Disease: Influence of Exercise and Nutrition. *European Journal of Preventive Cardiology* **2022**, *29* (17), 2183–2199.

(381) Zhang, W.; Song, M.; Qu, J.; Liu, G. H. Epigenetic Modifications in Cardiovascular Aging and Diseases. *Circ. Res.* **2018**, *123* (7), 773–786.

(382) Jiang, W.; Agrawal, D. K.; Boosani, C. S. Cell-specific Histone Modifications in Atherosclerosis. *Molecular Medicine Reports* **2018**, *18* (2), 1215–1224.

(383) Zhang, L.; Xia, C.; Yang, Y.; Sun, F.; Zhang, Y.; Wang, H.; Liu, R.; Yuan, M. DNA Methylation and Histone Post-translational Modifications in Atherosclerosis and a Novel Perspective for Epigenetic Therapy. *Cell Communication and Signaling* **2023**, *21* (1), 344.

(384) Mao, S. Q.; Sun, J. H.; Gu, T. L.; Zhu, F. B.; Yin, F. Y.; Zhang, L. N. Hypomethylation of Interleukin-6 (IL-6) Gene Increases the Risk of Essential Hypertension: A Matched Case-control Study. *Journal of Human Hypertension* **2017**, *31* (8), 530–536.

(385) Yu, J.; Qiu, Y.; Yang, J.; Bian, S.; Chen, G.; Deng, M.; Kang, H.; Huang, L. DNMT1-PPARgamma Pathway in Macrophages Regulates Chronic Inflammation and Atherosclerosis Development in Mice. *Sci. Rep.* **2016**, *6*, 30053.

(386) Levy, E.; Spahis, S.; Bigras, J. L.; Delvin, E.; Borys, J. M. The Epigenetic Machinery in Vascular Dysfunction and Hypertension. *Current Hypertension Reports* **2017**, *19* (6), S2.

(387) Xu, X. F.; Ma, X. L.; Shen, Z.; Wu, X. L.; Cheng, F.; Du, L. Z. Epigenetic Regulation of the Endothelial Nitric Oxide Synthase Gene in Persistent Pulmonary Hypertension of the Newborn Rat. *Journal of Hypertension* **2010**, *28* (11), 2227–2235.

(388) Liao, Y.; Chu, C.; Yan, Y.; Wang, D.; Ma, Q.; Gao, K.; Sun, Y.; Hu, J.; Zheng, W.; Mu, J. High Dietary Salt Intake Is Associated With Histone Methylation in Salt-Sensitive Individuals. *Frontiers in Nutrition* **2022**, *9*, 857562.

(389) Fujita, T. Recent Advances in Hypertension: Epigenetic Mechanism Involved in Development of Salt-Sensitive Hypertension. *Hypertension* **2023**, *80* (4), 711–718.

(390) Liao, Y.; Chu, C.; Ma, Q.; Yan, Y.; Wang, D.; Sun, Y.; Wang, Y.; Hu, J.; Chen, C.; Mu, J. Transient High Salt Intake Causes Epigenetic Changes and Leads to Persistent Inflammatory Activation to Produce "Salt Memory. *Journal of Nutritional Biochemistry* **2023**, *115*, 109281.

(391) Salvarani, N.; Crasto, S.; Miragoli, M.; Bertero, A.; Paulis, M.; Kunderfranco, P.; Serio, S.; Forni, A.; Lucarelli, C.; Dal Ferro, M.; Larcher, V.; Sinagra, G.; Vezzoni, P.; Murry, C. E.; Faggian, G.; Condorelli, G.; Di Pasquale, E. The K219T-Lamin Mutation Induces Conduction Defects Through Epigenetic Inhibition of SCN5A in Human Cardiac Laminopathy. *Nat. Commun.* **2019**, *10* (1), 2267.

(392) Duygu, B.; Poels, E. M.; da Costa Martins, P. A. Genetics and Epigenetics of Arrhythmia and Heart Failure. *Frontiers in Genetics* **2013**, *4*, 219.

(393) Krolevets, M.; Cate, V. T.; Prochaska, J. H.; Schulz, A.; Rapp, S.; Tenzer, S.; Andrade-Navarro, M. A.; Horvath, S.; Niehrs, C.; Wild, P. S. DNA Methylation and Cardiovascular Disease in Humans: A Systematic Review and Database of Known CpG Methylation Sites. *Clinical Epigenetics* **2023**, *15* (1), S6.

(394) Funamoto, M.; Imanishi, M.; Tsuchiya, K.; Ikeda, Y. Roles of Histone Acetylation Sites in Cardiac Hypertrophy and Heart Failure. *Frontiers in Cardiovascular Medicine* **2023**, *10*, 1133611.

(395) Huang, X. H.; Li, J. L.; Li, X. Y.; Wang, S. X.; Jiao, Z. H.; Li, S. Q.; Liu, J.; Ding, J. miR-208a in Cardiac Hypertrophy and Remodeling. *Frontiers in Cardiovascular Medicine* **2021**, *8*, 773314.

(396) Zeng, M.; Zhen, J.; Zheng, X.; Qiu, H.; Xu, X.; Wu, J.; Lin, Z.; Hu, J. The Role of DNA Methylation in Ischemic Stroke: A Systematic Review. *Frontiers in Neurology* **2020**, *11*, 566124.

(397) Su, Y.; Zhang, L.; Zhou, Y.; Ding, L.; Li, L.; Wang, Z. The Progress of Research on Histone Methylation in Ischemic Stroke Pathogenesis. *Journal of Physiology and Biochemistry* **2022**, *78* (1), 1–8.

(398) Soler-Botija, C.; Galvez-Monton, C.; Bayes-Genis, A. Epigenetic Biomarkers in Cardiovascular Diseases. *Frontiers in Genetics* **2019**, *10*, 950.

(399) Baccarelli, A. A.; Ordovas, J. Epigenetics of Early Cardiometabolic Disease: Mechanisms and Precision Medicine. *Circ. Res.* **2023**, *132* (12), 1648–1662.

(400) Corbin, L. J.; White, S. J.; Taylor, A. E.; Williams, C. M.; Taylor, K.; van den Bosch, M. T.; Teasdale, J. E.; Jones, M.; Bond, M.; Harper, M. T.; Falk, L.; Groom, A.; Hazell, G. G. J.; Paternoster, L.; Munafo, M. R.; Nordestgaard, B. G.; Tybjaerg-Hansen, A.; Bojesen, S. E.; Relton, C.; Min, J. L.; Davey Smith, G.; Mumford, A. D.; Poole, A. W.; Timpson, N. J. Epigenetic Regulation of F2RL3 Associates With Myocardial Infarction and Platelet Function. *Circ. Res.* **2022**, *130* (3), 384–400.

(401) Langsted, A.; Bojesen, S. E.; Stroses, E. S. G.; Nordestgaard, B. G. AHRR Hypomethylation as an Epigenetic Marker of Smoking History Predicts Risk of Myocardial Infarction in Former Smokers. *Atherosclerosis* **2020**, *312*, 8–15.

(402) Hohl, M.; Wagner, M.; Reil, J. C.; Muller, S. A.; Tauchnitz, M.; Zimmer, A. M.; Lehmann, L. H.; Thiel, G.; Bohm, M.; Backs, J.; Maack, C. HDAC4 Controls Histone Methylation in Response to Elevated Cardiac Load. *J. Clin. Invest.* **2013**, *123* (3), 1359–1370.

(403) Pei, J.; Harakalova, M.; Treibel, T. A.; Lumbers, R. T.; Boukens, B. J.; Efimov, I. R.; van Dinter, J. T.; Gonzalez, A.; Lopez, B.; El Azzouzi, H.; van den Dungen, N.; van Dijk, C. G. M.; Krebber, M. M.; den Ruijter, H. M.; Pasterkamp, G.; Duncker, D. J.; Nieuwenhuis, E. E. S.; de Weger, R.; Huibers, M. M.; Vink, A.; Moore, J. H.; Moon, J. C.; Verhaar, M. C.; Kararigas, G.; Mokry, M.; Asselbergs, F. W.; Cheng, C. H3K27ac Acetylation Signatures Reveal the Epigenomic Reorganization in Remodeled Non-failing Human Hearts. *Clinical Epigenetics* **2020**, *12* (1), 106.

(404) Xue, S.; Liu, D.; Zhu, W.; Su, Z.; Zhang, L.; Zhou, C.; Li, P. Circulating MiR-17–5p, MiR-126–5p and MiR-145–3p Are Novel Biomarkers for Diagnosis of Acute Myocardial Infarction. *Frontiers in Physiology* **2019**, *10*, 123.

(405) Zhang, L.; Chen, X.; Su, T.; Li, H.; Huang, Q.; Wu, D.; Yang, C.; Han, Z. Circulating miR-499 are Novel and Sensitive Biomarker of Acute Myocardial Infarction. *Journal of Thoracic Disease* **2015**, *7* (3), 303–308.

(406) Zhai, C.; Li, R.; Hou, K.; Chen, J.; Alzogool, M.; Hu, Y.; Zhang, J.; Zhang, Y.; Wang, L.; Zhang, R.; Cong, H. Value of Blood-Based microRNAs in the Diagnosis of Acute Myocardial Infarction: A

Systematic Review and Meta-Analysis. *Frontiers in Physiology* **2020**, *11*, 691.

(407) Pillon, N. J.; Loos, R. J. F.; Marshall, S. M.; Zierath, J. R. Metabolic Consequences of Obesity and Type 2 Diabetes: Balancing Genes and Environment for Personalized Care. *Cell* **2021**, *184* (6), 1530–1544.

(408) Long, Y.; Mao, C.; Liu, S.; Tao, Y.; Xiao, D. Epigenetic Modifications in Obesity-associated Diseases. *MedComm* (2020) **2024**, *5* (2), No. e496.

(409) Maude, H.; Sanchez-Cabanillas, C.; Cebola, I. Epigenetics of Hepatic Insulin Resistance. *Frontiers in Endocrinology* **2021**, *12*, 681356.

(410) Chew, N. W. S.; Ng, C. H.; Tan, D. J. H.; Kong, G.; Lin, C.; Chin, Y. H.; Lim, W. H.; Huang, D. Q.; Quek, J.; Fu, C. E.; Xiao, J.; Syn, N.; Foo, R.; Khoo, C. M.; Wang, J. W.; Dimitriadis, G. K.; Young, D. Y.; Siddiqui, M. S.; Lam, C. S. P.; Wang, Y.; Figtree, G. A.; Chan, M. Y.; Cummings, D. E.; Noureddin, M.; Wong, V. W.; Ma, R. C. W.; Mantzoros, C. S.; Sanyal, A.; Muthiah, M. D. The Global Burden of Metabolic Disease: Data from 2000 to 2019. *Cell Metabolism* **2023**, *35* (3), 414–428.e3.

(411) Sales, V. M.; Ferguson-Smith, A. C.; Patti, M. E. Epigenetic Mechanisms of Transmission of Metabolic Disease across Generations. *Cell Metabolism* **2017**, *25* (3), 559–571.

(412) Iacomino, G.; Siani, A. Role of microRNAs in Obesity and Obesity-related Diseases. *Genes & Nutrition* **2017**, *12*, 23.

(413) Davegardh, C.; Garcia-Calzon, S.; Bacos, K.; Ling, C. DNA Methylation in the Pathogenesis of Type 2 Diabetes in Humans. *Molecular Metabolism* **2018**, *14*, 12–25.

(414) Xu, F.; Guo, W. The Progress of Epigenetics in the Development and Progression of Non-alcoholic Fatty Liver Disease. *Liver Research* **2020**, *4* (3), 118–123.

(415) Gallego-Duran, R.; Romero-Gomez, M. Epigenetic Mechanisms in Non-alcoholic Fatty Liver Disease: An Emerging Field. *World Journal of Hepatology* **2015**, *7* (24), 2497–2502.

(416) Carson, C.; Lawson, H. A. Epigenetics of Metabolic Syndrome. *Physiological Genomics* **2018**, *50* (11), 947–955.

(417) Silva-Ochoa, A. D.; Velastegui, E.; Falconi, I. B.; Garcia-Solorzano, V. I.; Rendon-Riofrio, A.; Sanguna-Soliz, G. A.; Vanden Berghe, W.; Orellana-Manzano, A. Metabolic Syndrome: Nutri-epigenetic Cause or Consequence? *Heliyon* **2023**, *9* (11), No. e21106.

(418) Dayeh, T.; Volkov, P.; Salo, S.; Hall, E.; Nilsson, E.; Olsson, A. H.; Kirkpatrick, C. L.; Wollheim, C. B.; Eliasson, L.; Ronn, T.; Bacos, K.; Ling, C. Genome-Wide DNA Methylation Analysis of Human Pancreatic Islets from Type 2 Diabetic and Non-Diabetic Donors Identifies Candidate Genes that Influence Insulin Secretion. *PLoS Genetics* **2014**, *10* (3), No. e1004160.

(419) An, F.; Liu, C.; Wang, X.; Li, T.; Fu, H.; Bao, B.; Cong, H.; Zhao, J. Effect of ABCA1 Promoter Methylation on Premature Coronary Artery Disease and Its Relationship with Inflammation. *BMC Cardiovascular Disorders* **2021**, *21* (1), 78.

(420) Soriano-Tarraga, C.; Jimenez-Conde, J.; Giralte-Steinhauer, E.; Mola-Caminal, M.; Vivanco-Hidalgo, R. M.; Ois, A.; Rodriguez-Campello, A.; Cuadrado-Godia, E.; Sayols-Baixeras, S.; Elosua, R.; Roquer, J.; Consortium, G. Epigenome-wide Association Study Identifies TXNIP Gene Associated with Type 2 Diabetes Mellitus and Sustained Hyperglycemia. *Hum. Mol. Genet.* **2016**, *25* (3), 609–619.

(421) Wu, Y.; Chen, W.; Zhao, Y.; Gu, M.; Gao, Y.; Ke, Y.; Wang, L.; Wang, M.; Zhang, W.; Chen, Y.; Huo, W.; Fu, X.; Li, X.; Zhang, D.; Qin, P.; Hu, F.; Liu, Y.; Sun, X.; Zhang, M.; Hu, D. Visit to Visit Transition in TXNIP Gene Methylation and the Risk of Type 2 Diabetes Mellitus: A Nested Case-control Study. *Journal of Human Genetics* **2024**, *69* (7), 311–319.

(422) Krause, C.; Sievert, H.; Geissler, C.; Grohs, M.; El Gammal, A. T.; Wolter, S.; Ohlei, O.; Kilpert, F.; Kramer, U. M.; Kasten, M.; Klein, C.; Brabant, G. E.; Mann, O.; Lehnert, H.; Kirchner, H. Critical Evaluation of the DNA-Methylation Markers ABCG1 and SREBF1 for Type 2 Diabetes Stratification. *Epigenomics* **2019**, *11* (8), 885–897.

(423) Zeng, H.; Li, W.; Xia, M.; Ge, J.; Ma, H.; Chen, L.; Pan, B.; Lin, H.; Wang, S.; Gao, X. Longitudinal Association of Peripheral Blood

DNA Methylation with Liver Fat Content: Distinguishing Between Predictors and Biomarkers. *Lipids in Health and Disease* **2024**, *23* (1), 309.

(424) Miao, F.; Chen, Z.; Genuth, S.; Paterson, A.; Zhang, L.; Wu, X.; Li, S. M.; Cleary, P.; Riggs, A.; Harlan, D. M.; Lorenzi, G.; Kolterman, O.; Sun, W.; Lachin, J. M.; Natarajan, R.; Group, D. E. R. Evaluating the Role of Epigenetic Histone Modifications in the Metabolic Memory of Type 1 Diabetes. *Diabetes* **2014**, *63* (5), 1748–1762.

(425) Higuchi, C.; Nakatsuka, A.; Eguchi, J.; Teshigawara, S.; Kanzaki, M.; Katayama, A.; Yamaguchi, S.; Takahashi, N.; Murakami, K.; Ogawa, D.; Sasaki, S.; Makino, H.; Wada, J. Identification of Circulating miR-101, miR-375 and miR-802 as Biomarkers for Type 2 Diabetes. *Metabolism* **2015**, *64* (4), 489–497.

(426) Nemecek, M.; Stefan, D. S.; Comarita, I. K.; Constantin, A.; Tanko, G.; Guja, C.; Georgescu, A. Microvesicle-associated and Circulating microRNAs in Diabetic Dyslipidemia: miR-218, miR-132, miR-143, and miR-21, miR-122, miR-155 have Biomarker Potential. *Cardiovascular Diabetology* **2023**, *22* (1), 260.

(427) Willmer, T.; Johnson, R.; Louw, J.; Pfeiffer, C. Blood-Based DNA Methylation Biomarkers for Type 2 Diabetes: Potential for Clinical Applications. *Frontiers in Endocrinology* **2018**, *9*, 744.

(428) Sun, B.; Hu, L.; Luo, Z. Y.; Chen, X. P.; Zhou, H. H.; Zhang, W. DNA Methylation Perspectives in the Pathogenesis of Autoimmune Diseases. *Clinical Immunology* **2016**, *164*, 21–27.

(429) Pan, W.; Zhu, S.; Yuan, M.; Cui, H.; Wang, L.; Luo, X.; Li, J.; Zhou, H. H.; Tang, Y.; Shen, N. MicroRNA-21 and MicroRNA-148a contribute to DNA hypomethylation in lupus CD4+ T cells by directly and indirectly targeting DNA methyltransferase. *J. Immunol.* **2010**, *184*, 6773–6781.

(430) Frank-Bertoncelj, M.; Trenkmann, M.; Klein, K.; Karouzakis, E.; Rehrauer, H.; Bratus, A.; Kolling, C.; Armaka, M.; Filer, A.; Michel, B. A.; Gay, R. E.; Buckley, C. D.; Kollias, G.; Gay, S.; Ospelt, C. Epigenetically-driven Anatomical Diversity of Synovial Fibroblasts Guides Joint-specific Fibroblast Functions. *Nat. Commun.* **2017**, *8*, 14852.

(431) Du, C.; Liu, C.; Kang, J.; Zhao, G.; Ye, Z.; Huang, S.; Li, Z.; Wu, Z.; Pei, G. MicroRNA miR-326 Regulates TH-17 Differentiation and is Associated with the Pathogenesis of Multiple Sclerosis. *Nature Immunology* **2009**, *10* (12), 1252–1259.

(432) McCoy, C. E. miR-155 Dysregulation and Therapeutic Intervention in Multiple Sclerosis. *Advances in Experimental Medicine and Biology* **2017**, *1024*, 111–131.

(433) Zhang, J.; Chen, L. M.; Zou, Y.; Zhang, S.; Xiong, F.; Wang, C. Y. Implication of Epigenetic Factors in the Pathogenesis of Type 1 Diabetes. *Chinese Medical Journal* **2021**, *134* (9), 1031–1042.

(434) Xiao, X.; Mao, X.; Chen, D.; Yu, B.; He, J.; Yan, H.; Wang, J. miRNAs Can Affect Intestinal Epithelial Barrier in Inflammatory Bowel Disease. *Frontiers in Immunology* **2022**, *13*, 868229.

(435) Yarani, R.; Shojaeian, A.; Palasca, O.; Doncheva, N. T.; Jensen, L. J.; Gorodkin, J.; Pociot, F. Differentially Expressed miRNAs in Ulcerative Colitis and Crohn's Disease. *Frontiers in Immunology* **2022**, *13*, 865777.

(436) de Oliveira, E. C. S.; Quaglio, A. E. V.; Grillo, T. G.; Di Stasi, L. C.; Sasaki, L. Y. MicroRNAs in Inflammatory Bowel Disease: What Do We Know and What Can We Expect? *World Journal of Gastroenterology* **2024**, *30* (16), 2184–2190.

(437) Kronfol, M. M.; Dozmorov, M. G.; Huang, R.; Slattum, P. W.; McClay, J. L. The Role of Epigenomics in Personalized Medicine. *Expert Review of Precision Medicine and Drug Development* **2017**, *2* (1), 33–45.

(438) De Riso, G.; Cocozza, S. Artificial Intelligence for Epigenetics: Towards Personalized Medicine. *Curr. Med. Chem.* **2021**, *28* (32), 6654–6674.

(439) Cao, Q.; Dan, Z.; Hou, N.; Yan, L.; Yuan, X.; Lu, H.; Yu, S.; Zhang, J.; Xiao, H.; Liu, Q.; Zhang, X.; Zhang, M.; Pang, M. Discovery and Validation of Colorectal Cancer Tissue-Specific Methylation Markers: A Dual-Center Retrospective Cohort Study. *Clinical Epigenetics* **2024**, *16* (1), 122.

- (440) Locke, W. J.; Guanzon, D.; Ma, C.; Liew, Y. J.; Duesing, K. R.; Fung, K. Y. C.; Ross, J. P. DNA Methylation Cancer Biomarkers: Translation to the Clinic. *Frontiers in Genetics* **2019**, *10*, 1150.
- (441) Lim, J. H.; Lee, D. E.; Park, S. Y.; Kim, D. J.; Ahn, H. K.; Han, Y. J.; Kim, M. Y.; Ryu, H. M. Disease Specific Characteristics of Fetal Epigenetic Markers for Non-invasive Prenatal Testing of Trisomy 21. *BMC Medical Genomics* **2014**, *7*, 1.
- (442) Kwon, E. J.; Kim, Y. J. What is Fetal Programming?: A Lifetime Health is Under the Control of In utero Health. *Obstetrics & Gynecology Science* **2017**, *60* (6), 506–519.
- (443) Tokarz, P.; Pawlowska, E.; Bialkowska-Warzecha, J.; Blasiak, J. The Significance of DNA Methylation Profile in Metastasis-related Genes for the Progression of Colorectal Cancer. *Cellular and Molecular Biology* **2017**, *63* (2), 79–87.
- (444) Li, W.; Guo, L.; Tang, W.; Ma, Y.; Wang, X.; Shao, Y.; Zhao, H.; Ying, J. Identification of DNA Methylation Biomarkers for Risk of Liver Metastasis in Early-stage Colorectal Cancer. *Clinical Epigenetics* **2021**, *13* (1), 126.
- (445) Zhou, J.; Li, M.; Wang, X.; He, Y.; Xia, Y.; Sweeney, J. A.; Kopp, R. F.; Liu, C.; Chen, C. Drug Response-Related DNA Methylation Changes in Schizophrenia, Bipolar Disorder, and Major Depressive Disorder. *Frontiers in Neuroscience* **2021**, *15*, 674273.
- (446) Nepali, K.; Liou, J. P. Recent Developments in Epigenetic Cancer Therapeutics: Clinical Advancement and Emerging Trends. *Journal of Biomedical Science* **2021**, *28* (1), 27.
- (447) Farani, M. R.; Sarlak, M.; Gholami, A.; Azaraian, M.; Binabaj, M. M.; Kakavandi, S.; Tambuwala, M. M.; Taheriazam, A.; Hashemi, M.; Ghasemi, S. Epigenetic Drugs as New Emerging Therapeutics: What is the Scale's Orientation of Application and Challenges? *Pathology - Research and Practice* **2023**, *248*, 154688.
- (448) Seto, E.; Yoshida, M. Erasers of histone acetylation: the histone deacetylase enzymes. *Cold Spring Harbor Perspectives in Biology* **2014**, *6* (4), a018713.
- (449) Narita, T.; Weinert, B. T.; Choudhary, C. Functions and mechanisms of non-histone protein acetylation. *Nat. Rev. Mol. Cell Biol.* **2019**, *20* (3), 156–174.
- (450) Shanmugam, G.; Rakshit, S.; Sarkar, K. HDAC Inhibitors: Targets for Tumor Therapy, Immune Modulation and Lung Diseases. *Translational Oncology* **2022**, *16*, 101312.
- (451) Bondarev, A. D.; Attwood, M. M.; Jonsson, J.; Chubarev, V. N.; Tarasov, V. V.; Schioth, H. B. Recent Developments of HDAC Inhibitors: Emerging Indications and Novel Molecules. *Br. J. Clin. Pharmacol.* **2021**, *87* (12), 4577–4597.
- (452) Luo, J.; Su, F.; Chen, D.; Shiloh, A.; Gu, W. Deacetylation of p53 modulates its effect on cell growth and apoptosis. *Nature* **2000**, *408* (6810), 377–381. From NLM Medline
- (453) Sabari, B. R.; Zhang, D.; Allis, C. D.; Zhao, Y. Metabolic regulation of gene expression through histone acylations. *Nat. Rev. Mol. Cell Biol.* **2017**, *18* (2), 90–101. From NLM Medline
- (454) Greer, C. B.; Tanaka, Y.; Kim, Y. J.; Xie, P.; Zhang, M. Q.; Park, I. H.; Kim, T. H. Histone Deacetylases Positively Regulate Transcription through the Elongation Machinery. *Cell Reports* **2015**, *13* (7), 1444–1455.
- (455) Bali, P.; Pranpat, M.; Bradner, J.; Balasis, M.; Fiskus, W.; Guo, F.; Rocha, K.; Kumaraswamy, S.; Boyapalle, S.; Atadja, P.; Seto, E.; Bhalla, K. Inhibition of Histone Deacetylase 6 Acetylates and Disrupts the Chaperone Function of Heat Shock Protein 90: A Novel Basis for Antileukemia Activity of Histone Deacetylase Inhibitors. *J. Biol. Chem.* **2005**, *280* (29), 26729–26734.
- (456) Gravina, G. L.; Festuccia, C.; Marampon, F.; Popov, V. M.; Pestell, R. G.; Zani, B. M.; Tombolini, V. Biological Rationale for the Use of DNA Methyltransferase Inhibitors as New Strategy for Modulation of Tumor Response to Chemotherapy and Radiation. *Molecular Cancer* **2010**, *9*, 305.
- (457) Giri, A. K.; Aittokallio, T. DNMT Inhibitors Increase Methylation in the Cancer Genome. *Frontiers in Pharmacology* **2019**, *10*, 385.
- (458) Mehdipour, P.; Chen, R.; De Carvalho, D. D. The Next Generation of DNMT Inhibitors. *Nature Cancer* **2021**, *2* (10), 1000–1001.
- (459) Stathis, A.; Bertoni, F. BET Proteins as Targets for Anticancer Treatment. *Cancer Discovery* **2018**, *8* (1), 24–36.
- (460) Morgado-Pascual, J. L.; Rayego-Mateos, S.; Tejedor, L.; Suarez-Alvarez, B.; Ruiz-Ortega, M. Bromodomain and Extraterminal Proteins as Novel Epigenetic Targets for Renal Diseases. *Frontiers in Pharmacology* **2019**, *10*, 1315.
- (461) Alqahtani, A.; Choucair, K.; Ashraf, M.; Hammouda, D. M.; Alloghbi, A.; Khan, T.; Senzer, N.; Nemunaitis, J. Bromodomain and Extra-Terminal Motif Inhibitors: A Review of Preclinical and Clinical Advances in Cancer Therapy. *Future Science OA* **2019**, *5* (3), FSO372.
- (462) Wang, Z. Q.; Zhang, Z. C.; Wu, Y. Y.; Pi, Y. N.; Lou, S. H.; Liu, T. B.; Lou, G.; Yang, C. Bromodomain and Extraterminal (BET) Proteins: Biological Functions, Diseases, and Targeted Therapy. *Signal Transduction and Targeted Therapy* **2023**, *8* (1), 420.
- (463) Thinnies, C. C.; England, K. S.; Kawamura, A.; Chowdhury, R.; Schofield, C. J.; Hopkinson, R. J. Targeting Histone Lysine Demethylases - Progress, Challenges, and the Future. *Biochim. Biophys. Acta* **2014**, *1839* (12), 1416–1432.
- (464) McAllister, T. E.; England, K. S.; Hopkinson, R. J.; Brennan, P. E.; Kawamura, A.; Schofield, C. J. Recent Progress in Histone Demethylase Inhibitors. *J. Med. Chem.* **2016**, *59* (4), 1308–1329.
- (465) Jambhekar, A.; Anastas, J. N.; Shi, Y. Histone Lysine Demethylase Inhibitors. *Cold Spring Harbor Perspectives in Medicine* **2017**, *7* (1), a026484.
- (466) Lin, H.; Li, Q.; Li, Q.; Zhu, J.; Gu, K.; Jiang, X.; Hu, Q.; Feng, F.; Qu, W.; Chen, Y.; Sun, H. Small Molecule KDM4s Inhibitors as Anti-cancer Agents. *Journal of Enzyme Inhibition and Medicinal Chemistry* **2018**, *33* (1), 777–793.
- (467) Gray, Z. H.; Honer, M. A.; Ghatalia, P.; Shi, Y.; Whetstone, J. R. 20 years of histone lysine demethylases: From discovery to the clinic and beyond. *Cell* **2025**, *188* (7), 1747–1783. From NLM Medline
- (468) Rugo, H. S.; Jacobs, I.; Sharma, S.; Scappaticci, F.; Paul, T. A.; Jensen-Pergakes, K.; Malouf, G. G. The Promise for Histone Methyltransferase Inhibitors for Epigenetic Therapy in Clinical Oncology: A Narrative Review. *Advances in Therapy* **2020**, *37* (7), 3059–3082.
- (469) Liao, Q.; Yang, J.; Ge, S.; Chai, P.; Fan, J.; Jia, R. Novel Insights into Histone Lysine Methyltransferases in Cancer Therapy: From Epigenetic Regulation to Selective Drugs. *Journal of Pharmaceutical Analysis* **2023**, *13* (2), 127–141.
- (470) Marzochi, L. L.; Cuzziol, C. I.; Nascimento Filho, C.; Dos Santos, J. A.; Castanhole-Nunes, M. M. U.; Pavarino, E. C.; Guerra, E. N. S.; Goloni-Bertollo, E. M. Use of Histone Methyltransferase Inhibitors in Cancer Treatment: A Systematic Review. *Eur. J. Pharmacol.* **2023**, *944*, 175590.
- (471) Hu, H.; Qian, K.; Ho, M. C.; Zheng, Y. G. Small Molecule Inhibitors of Protein Arginine Methyltransferases. *Expert Opinion on Investigational Drugs* **2016**, *25* (3), 335–358.
- (472) Zhu, Y.; Xia, T.; Chen, D. Q.; Xiong, X.; Shi, L.; Zuo, Y.; Xiao, H.; Liu, L. Promising Role of Protein Arginine Methyltransferases in Overcoming Anti-cancer Drug Resistance. *Drug Resistance Updates* **2024**, *72*, 101016.
- (473) Li, Y.; Ding, L.; Ren, S.; Zhang, W.; Rao, G. W. Protein Lysine Methyltransferases Inhibitors. *Curr. Med. Chem.* **2023**, *30* (27), 3060–3089.
- (474) Velez, J.; Kaniskan, H. U.; Jin, J. Recent Advances in Developing Degradable & Inhibitors of Lysine Methyltransferases. *Curr. Opin. Chem. Biol.* **2023**, *76*, 102356.
- (475) Aziz, N.; Hong, Y. H.; Kim, H. G.; Kim, J. H.; Cho, J. Y. Tumor-Suppressive Functions of Protein Lysine Methyltransferases. *Experimental & Molecular Medicine* **2023**, *55* (12), 2475–2497.
- (476) Gulati, N.; Beguelin, W.; Giulino-Roth, L. Enhancer of Zeste Homolog 2 (EZH2) Inhibitors. *Leukemia & Lymphoma* **2018**, *59* (7), 1574–1585.
- (477) Duan, R.; Du, W.; Guo, W. EZH2: A Novel Target for Cancer Treatment. *Journal of Hematology & Oncology* **2020**, *13* (1), 104.

- (478) Morschhauser, F.; Salles, G.; Batlevi, C. L.; Tilly, H.; Chaidos, A.; Phillips, T.; Burke, J.; Melnick, A. Taking the EZ Way: Targeting Enhancer of Zeste Homolog 2 in B-cell Lymphomas. *Blood Reviews* **2022**, *56*, 100988.
- (479) Upadhyay, V. A.; Brunner, A. M.; Fathi, A. T. Isocitrate Dehydrogenase (IDH) Inhibition as Treatment of Myeloid Malignancies: Progress and Future Directions. *Pharmacology & Therapeutics* **2017**, *177*, 123–128.
- (480) Liu, X.; Gong, Y. Isocitrate Dehydrogenase Inhibitors in Acute Myeloid Leukemia. *Biomark Research* **2019**, *7*, 22.
- (481) Liu, Y.; Xu, W.; Li, M.; Yang, Y.; Sun, D.; Chen, L.; Li, H.; Chen, L. The Regulatory Mechanisms and Inhibitors of Isocitrate Dehydrogenase 1 in Cancer. *Acta Pharmaceutica Sinica B* **2023**, *13* (4), 1438–1466.
- (482) Tian, W.; Zhang, W.; Wang, Y.; Jin, R.; Wang, Y.; Guo, H.; Tang, Y.; Yao, X. Recent Advances of IDH1 Mutant Inhibitor in Cancer Therapy. *Frontiers in Pharmacology* **2022**, *13*, 982424.
- (483) Roy, L.; Chatterjee, O.; Bose, D.; Roy, A.; Chatterjee, S. Noncoding RNA as an Influential Epigenetic Modulator with Promising Roles in Cancer Therapeutics. *Drug Discovery Today* **2023**, *28* (9), 103690.
- (484) Yang, X.; Liu, M.; Li, M.; Zhang, S.; Hiju, H.; Sun, J.; Mao, Z.; Zheng, M.; Feng, B. Epigenetic Modulations of Noncoding RNA: A Novel Dimension of Cancer Biology. *Molecular Cancer* **2020**, *19* (1), 64. From NLM Medline
- (485) A Study to Assess the Safety, Pharmacokinetics, and Antitumor Activity of Oral TACH101 in Participants With Advanced or Metastatic Cancer. 2024. <https://clinicaltrials.gov/study/NCT05076552?term=NCT05076552&rank=1> (accessed 2025 November 6).
- (486) Wu, Q.; Young, B.; Wang, Y.; Davidoff, A. M.; Rankovic, Z.; Yang, J. Recent Advances with KDM4 Inhibitors and Potential Applications. *J. Med. Chem.* **2022**, *65* (14), 9564–9579.
- (487) Schapira, M. Chemical Inhibition of Protein Methyltransferases. *Cell Chem. Biol.* **2016**, *23* (9), 1067–1076. From NLM Medline
- (488) Kaniskan, H. U.; Konze, K. D.; Jin, J. Selective inhibitors of protein methyltransferases. *J. Med. Chem.* **2015**, *58* (4), 1596–1629. From NLM Medline
- (489) Luo, M. Inhibitors of protein methyltransferases as chemical tools. *Epigenomics* **2015**, *7* (8), 1327–1338. From NLM Medline
- (490) Pirozzi, C. J.; Yan, H. The Implications of IDH Mutations for Cancer Development and Therapy. *Nature Reviews Clinical Oncology* **2021**, *18* (10), 645–661.
- (491) Waitkus, M. S.; Diplas, B. H.; Yan, H. Biological Role and Therapeutic Potential of IDH Mutations in Cancer. *Cancer Cell* **2018**, *34* (2), 186–195.
- (492) Nakhate, V.; Lasica, A. B.; Wen, P. Y. The Role of Mutant IDH Inhibitors in the Treatment of Glioma. *Current Neurology and Neuroscience Reports* **2024**, *24* (12), 631–643.
- (493) de Lera, A. R.; Ganesan, A. Epigenetic Polypharmacology: From Combination Therapy to Multitargeted Drugs. *Clinical Epigenetics* **2016**, *8*, 105.
- (494) Miranda Furtado, C. L.; Dos Santos Luciano, M. C.; Silva Santos, R. D.; Furtado, G. P.; Moraes, M. O.; Pessoa, C. Epidrugs: Targeting Epigenetic Marks in Cancer Treatment. *Epigenetics* **2019**, *14* (12), 1164–1176. From NLM Medline
- (495) Mondal, P.; Natesh, J.; Penta, D.; Meeran, S. M. Progress and Promises of Epigenetic Drugs and Epigenetic Diets in Cancer Prevention and Therapy: A Clinical Update. *Seminars in Cancer Biology* **2022**, *83*, 503–522. From NLM Medline
- (496) Morel, D.; Jeffery, D.; Aspeslagh, S.; Almouzni, G.; Postel-Vinay, S. Combining Epigenetic Drugs with Other Therapies for Solid Tumours - Past Lessons and Future Promise. *Nature Reviews Clinical Oncology* **2020**, *17* (2), 91–107. From NLM Medline
- (497) Stahl, M.; Kohrman, N.; Gore, S. D.; Kim, T. K.; Zeidan, A. M.; Prebet, T. Epigenetics in Cancer: A Hematological Perspective. *PLoS Genetics* **2016**, *12* (10), No. e1006193.
- (498) Duan, Y. C.; Zhang, S. J.; Shi, X. J.; Jin, L. F.; Yu, T.; Song, Y.; Guan, Y. Y. Research progress of dual inhibitors targeting crosstalk between histone epigenetic modulators for cancer therapy. *Eur. J. Med. Chem.* **2021**, *222*, 113588.
- (499) San Jose-Eneriz, E.; Agirre, X.; Rabal, O.; Vilas-Zornoza, A.; Sanchez-Arias, J. A.; Miranda, E.; Ugarte, A.; Roa, S.; Paiva, B.; Estella-Hermoso de Mendoza, A.; Alvarez, R. M.; Casares, N.; Segura, V.; Martin-Subero, J. I.; Ogi, F. X.; Soule, P.; Santiveri, C. M.; Campos-Olivas, R.; Castellano, G.; de Barrena, M. G. F.; Rodriguez-Madoz, J. R.; Garcia-Barchino, M. J.; Lasarte, J. J.; Avila, M. A.; Martinez-Climent, J. A.; Oyarzabal, J.; Prosper, F. Discovery of first-in-class reversible dual small molecule inhibitors against G9a and DNMTs in hematological malignancies. *Nat. Commun.* **2017**, *8*, 15424.
- (500) Jose-Eneriz, E. S.; Rabal, O.; Agirre, X.; Oyarzabal, J.; Prosper, F. Dual epigenetic modifiers for cancer therapy. *Molecular & Cellular Oncology* **2017**, *4* (4), No. e1342748.
- (501) Cai, X.; Zhai, H. X.; Wang, J.; Forrester, J.; Qu, H.; Yin, L.; Lai, C. J.; Bao, R.; Qian, C.; Discovery of 7-(4-(3-ethynylphenylamino)-7-methoxyquinazolin-6-yloxy)-N-hydroxyheptanamide. CUDC-101) as a potent multi-acting HDAC, EGFR, and HER2 inhibitor for the treatment of cancer. *J. Med. Chem.* **2010**, *53* (5), 2000–2009.
- (502) Morshed, M. N.; Parvez, S. A.; Akanda, R. I.; Saha, M. K.; Fardous, J.; Alam, M.; Zhao, Y.; Franco, O. E.; Hosen, M. J. Identification of novel small molecule inhibitors targeting multiple methyltransferase like proteins against hepatocellular carcinoma. *Sci. Rep.* **2025**, *15* (1), 33087.
- (503) Barcena-Varela, M.; Caruso, S.; Llerena, S.; Alvarez-Sola, G.; Uriarte, I.; Latasa, M. U.; Urtasun, R.; Rebouissou, S.; Alvarez, L.; Jimenez, M.; Santamaria, E.; Rodriguez-Ortigosa, C.; Mazza, G.; Rombouts, K.; San Jose-Eneriz, E.; Rabal, O.; Agirre, X.; Iraburu, M.; Santos-Laso, A.; Banales, J. M.; Zucman-Rossi, J.; Prosper, F.; Oyarzabal, J.; Berasain, C.; Avila, M. A.; Fernandez-Barrena, M. G. Dual Targeting of Histone Methyltransferase G9a and DNA-Methyltransferase 1 for the Treatment of Experimental Hepatocellular Carcinoma. *Hepatology* **2019**, *69* (2), 587–603.
- (504) Nylund, P.; Garrido-Zabala, B.; Tziola, S. I.; Mohajershojai, T.; Berglund, H.; Muylaert, C.; Van Hemelrijck, L. A.; Atienza Parraga, A.; Vasquez, L.; Jacob, J.; Bergquist, E.; Martin-Subero, J. I.; Oberg, F.; Karlsson, T.; Nestor, M.; De Bruyne, E.; Kalushkova, A.; Wiklund, H. J. Dual targeting of G9a and DNMTs induces antitumor effects in multiple myeloma. *Blood Advances* **2025**, *9* (19), 4825–4841.
- (505) Huang, W.; Zhu, Q.; Shi, Z.; Tu, Y.; Li, Q.; Zheng, W.; Yuan, Z.; Li, L.; Zu, X.; Hao, Y.; Chu, B.; Jiang, Y. Dual inhibitors of DNMT and HDAC induce viral mimicry to induce antitumor immunity in breast cancer. *Cell Death Discovery* **2024**, *10* (1), 143.
- (506) Rosenberg, L.; Vabret, N. Viral mimicry in cancer therapy. *Trends in Cancer* **2025**, *11* (12), 1185–1202.
- (507) Wei, L.; Mei, D.; Hu, S.; Du, S. Dual-target EZH2 inhibitor: latest advances in medicinal chemistry. *Future Medicinal Chemistry* **2024**, *16* (15), 1561–1582.
- (508) Cheng, Y.; He, C.; Wang, M.; Ma, X.; Mo, F.; Yang, S.; Han, J.; Wei, X. Targeting epigenetic regulators for cancer therapy: Mechanisms and advances in clinical trials. *Signal Transduction and Targeted Therapy* **2019**, *4*, 62.
- (509) Shirbhate, E.; Singh, V.; Jahoriya, V.; Mishra, A.; Veerasamy, R.; Tiwari, A. K.; Rajak, H. Dual inhibitors of HDAC and other epigenetic regulators: A novel strategy for cancer treatment. *Eur. J. Med. Chem.* **2024**, *263*, 115938.
- (510) Bell, H. W.; Feng, R.; Shah, M.; Yao, Y.; Douglas, J.; Doerfler, P. A.; Mayuranathan, T.; O'Dea, M. F.; Li, Y.; Wang, Y. D.; Zhang, J.; Mackay, J. P.; Cheng, Y.; Quinlan, K. G. R.; Weiss, M. J.; Crossley, M. Removal of promoter CpG methylation by epigenome editing reverses HBG silencing. *Nat. Commun.* **2025**, *16* (1), 6919.
- (511) Goudy, L.; Ha, A.; Borah, A. A.; Umhoefer, J. M.; Chow, L.; Tran, C.; Winters, A.; Talbot, A.; Hernandez, R.; Li, Z.; Subramanya, S.; Arab, A.; Kale, N.; Lee, J. H. J.; Muldoon, J. J.; Liu, C.; Schmidt, R.; Santangelo, P.; Carnevale, J.; Eyquem, J.; Shy, B. R.; Marson, A.; Gilbert, L. A. Integrated epigenetic and genetic programming of primary human T cells. *Nat. Biotechnol.* **2025**, DOI: 10.1038/s41587-025-02856-w.

- (512) Tune Therapeutics Presents First Data Supporting TUNE-401: a First-in-Class Epigenetic Silencer for Hepatitis B. <https://tunetx.com/tune-therapeutics-presents-first-data-supporting-tune-401-a-first-in-class-epigenetic-silencer-for-hepatitis-b/> (accessed 2026 January 2).
- (513) Stopper, D.; Honin, I.; Feller, F.; Hansen, F. K. Development of ethyl-hydrazide-based selective histone deacetylase 6 (HDAC6) PROTACs. *ACS Med. Chem. Lett.* **2025**, *16* (3), 487–495.
- (514) Livingston, J. A.; Blay, J. Y.; Trent, J.; Valverde, C.; Agulnik, M.; Gounder, M.; Le Cesne, A.; McKean, M.; Wagner, M. J.; Stacchiotti, S.; Agresta, S.; Quintas-Cardama, A.; Reilly, S. A.; Healy, K.; Hickman, D.; Zhao, T.; Ballesteros-Perez, A.; Khalil, A.; Collins, M. P.; Piel, J.; Horrigan, K.; Lefkovich, A.; Innis, S.; Lazar, A. J.; Cote, G. M.; Wagner, A. J. A phase I study of FHD-609, a heterobifunctional degrader of bromodomain-containing protein 9, in patients with advanced synovial sarcoma or SMARCB1-deficient tumors. *Clin. Cancer Res.* **2025**, *31* (4), 628–638.
- (515) Jackson, K. L.; Yin, N.; Agafonov, R. V.; Chaturvedi, P.; Cole, K.; Eron, S. J.; Good, A. C.; Hart, J. A.; He, M.; Henderson, C. S.; Huang, H.; Kirby, R. J.; Lee, L.; Moustakim, M.; Nasveschuk, C. G.; Phillips, A. J.; Pollock, R. M.; Poling, L. L.; Veits, G. K.; Yap, J.; Zeid, R.; Ali, A.; Crystal, A. S.; Hurh, E.; Liu, H.; Lobbardi, R.; Reyno, L.; Fisher, S. L. Discovery of CFT8634, a potent, selective, and orally bioavailable heterobifunctional degrader of BRD9. *J. Med. Chem.* **2025**, *68* (23), 24848–24868.
- (516) Hsu, J. H.; Rasmusson, T.; Robinson, J.; Pachl, F.; Read, J.; Kawatkar, S.; DH, O. D.; Bagal, S.; Code, E.; Rawlins, P.; Argyrou, A.; Tomlinson, R.; Gao, N.; Zhu, X.; Chiarparin, E.; Jacques, K.; Shen, M.; Woods, H.; Bednarski, E.; Wilson, D. M.; Drew, L.; Castaldi, M. P.; Fawell, S.; Bloecher, A. EED-targeted PROTACs degrade EED, EZH2, and SUZ12 in the PRC2 complex. *Cell Chemical Biology* **2020**, *27* (1), 41–46.e17.
- (517) Farnaby, W.; Koegl, M.; Roy, M. J.; Whitworth, C.; Diers, E.; Trainor, N.; Zollman, D.; Steurer, S.; Karolyi-Oezguer, J.; Riedmueller, C.; Gmaschitz, T.; Wachter, J.; Dank, C.; Galant, M.; Sharps, B.; Rumpel, K.; Traxler, E.; Gerstberger, T.; Schnitzer, R.; Petermann, O.; Greb, P.; Weinstabl, H.; Bader, G.; Zoephel, A.; Weiss-Puxbaum, A.; Ehrenhofer-Wolfer, K.; Wohrle, S.; Boehmelt, G.; Rinnenthal, J.; Arnhof, H.; Wiechens, N.; Wu, M. Y.; Owen-Hughes, T.; Ettmayer, P.; Pearson, M.; McConnell, D. B.; Ciulli, A. BAF complex vulnerabilities in cancer demonstrated via structure-based PROTAC design. *Nat. Chem. Biol.* **2019**, *15* (7), 672–680.
- (518) Xiao, Y.; Wang, J.; Zhao, L. Y.; Chen, X.; Zheng, G.; Zhang, X.; Liao, D. Discovery of histone deacetylase 3 (HDAC3)-specific PROTACs. *Chem. Commun.* **2020**, *56* (68), 9866–9869.
- (519) Macabuag, N.; Esmieu, W.; Breccia, P.; Jarvis, R.; Blackaby, W.; Lazari, O.; Urbonas, L.; Eznarriaga, M.; Williams, R.; Strijbosch, A.; Van de Bospoort, R.; Matthews, K.; Clissold, C.; Ladduwahetty, T.; Vater, H.; Heaphy, P.; Stafford, D. G.; Wang, H. J.; Mangette, J. E.; McAllister, G.; Beaumont, V.; Vogt, T. F.; Wilkinson, H. A.; Doherty, E. M.; Dominguez, C. Developing HDAC4-Selective Protein Degradators To Investigate the Role of HDAC4 in Huntington's Disease Pathology. *J. Med. Chem.* **2022**, *65* (18), 12445–12459.
- (520) Yang, K.; Song, Y.; Xie, H.; Wu, H.; Wu, Y. T.; Leisten, E. D.; Tang, W. Development of the first small molecule histone deacetylase 6 (HDAC6) degraders. *Bioorg. Med. Chem. Lett.* **2018**, *28* (14), 2493–2497.
- (521) A Study to Assess the Safety and Tolerability of CFT8634 in Locally Advanced or Metastatic SMARCB1-Perturbed Cancers, Including Synovial Sarcoma and SMARCB1-Null Tumors. 2022. <https://clinicaltrials.gov/ct2/show/NCT05355753> (accessed 2026 January 2).
- (522) FHD-609 in Subjects With Advanced Synovial Sarcoma or Advanced SMARCB1-Loss Tumors. 2021. <https://clinicaltrials.gov/study/NCT04965753?term=FHD-609&rank=1> (accessed 2026 January 2).
- (523) C4 Therapeutics Reports Third Quarter 2023 Financial Results and Recent Business Highlights. GlobalNewswire, 2023. <https://www.globenewswire.com/news-release/2023/11/01/2770978/0/en/C4-Therapeutics-Reports-Third-Quarter-2023-Financial-Results-and-Recent-Business-Highlights.html> (accessed 2026 January 2).
- (524) Foghorn Therapeutics Provides an Update on FHD-609. Foghorn Therapeutics, 2023. <https://ir.foghornrx.com/news-releases/news-release-details/foghorn-therapeutics-provides-update-fhd-609> (accessed 2026 January 2).
- (525) Ma, Z.; Bolinger, A. A.; Zhou, J. RIPTACs: A groundbreaking approach to drug discovery. *Drug Discovery Today* **2023**, *28* (11), 103774.
- (526) Raina, K.; Forbes, C. D.; Stronk, R.; Rappi, J. P., Jr.; Eastman, K. J.; Zaware, N.; Yu, X.; Li, H.; Bhardwaj, A.; Gerritz, S. W.; Forgione, M.; Hundt, A.; King, M. P.; Posner, Z. M.; Correia, A. D.; McGovern, A.; Puleo, D. E.; Chenard, R.; Mousseau, J. J.; Vergara, J. I.; Garvin, E.; Macaluso, J.; Martin, M.; Bassoli, K.; Jones, K.; Garcia, M.; Howard, K.; Yaggi, M.; Smith, L. M.; Chen, J. M.; Mayfield, A. B.; De Leon, C. A.; Hines, J.; Kayser-Bricker, K. J.; Crews, C. M. Regulated induced proximity targeting chimeras-RIPTACs-A heterobifunctional small molecule strategy for cancer selective therapies. *Cell Chemical Biology* **2024**, *31* (8), 1490–1502.e42.
- (527) A Study of RNK05047 in Subjects With Advanced Solid Tumors/Diffuse Large B-cell Lymphoma (CHAMP-1). 2022. <https://clinicaltrials.gov/study/NCT05487170?term=RNK05047&rank=1> (accessed 2026 January 2).
- (528) Peng, X.; Hu, Z.; Zeng, L.; Zhang, M.; Xu, C.; Lu, B.; Tao, C.; Chen, W.; Hou, W.; Cheng, K.; Bi, H.; Pan, W.; Chen, J. Overview of epigenetic degraders based on PROTAC, molecular glue, and hydrophobic tagging technologies. *Acta Pharmaceutica Sinica B* **2024**, *14* (2), 533–578.
- (529) Collotta, D.; Bertocchi, I.; Chiapello, E.; Collino, M. Antisense oligonucleotides: A novel Frontier in pharmacological strategy. *Frontiers in Pharmacology* **2023**, *14*, 1304342.
- (530) Krey-Grauert, I.; Ferro, I.; Wagner, M. Antisense oligonucleotide therapies for monogenic disorders. *Medizinische Genetik* **2025**, *37* (3), 179–187.
- (531) Lauffer, M. C.; van Roon-Mom, W.; Aartsma-Rus, A.; Collaborative, N. Possibilities and limitations of antisense oligonucleotide therapies for the treatment of monogenic disorders. *Communications Medicine* **2024**, *4* (1), 6.
- (532) Rom, A.; Melamed, L.; Gil, N.; Goldrich, M. J.; Kadir, R.; Golan, M.; Biton, I.; Perry, R. B.; Ulitsky, I. Regulation of CHD2 expression by the Chaserr long noncoding RNA gene is essential for viability. *Nat. Commun.* **2019**, *10* (1), 5092.
- (533) Xiao, L.; Tien, J. C.; Vo, J.; Tan, M.; Parolia, A.; Zhang, Y.; Wang, L.; Qiao, Y.; Shukla, S.; Wang, X.; Zheng, H.; Su, F.; Jing, X.; Luo, E.; Delekt, A.; Juckette, K. M.; Xu, A.; Cao, X.; Alva, A. S.; Kim, Y.; MacLeod, A. R.; Chinnaiyan, A. M. Epigenetic reprogramming with antisense oligonucleotides enhances the effectiveness of androgen receptor inhibition in castration-resistant prostate cancer. *Cancer Res.* **2018**, *78* (20), 5731–5740. From NLM Medline
- (534) Wang, Z. The guideline of the design and validation of MiRNA mimics. *Methods Mol. Biol.* **2011**, *676*, 211–223.
- (535) Diener, C.; Keller, A.; Meese, E. Emerging concepts of miRNA therapeutics: From cells to clinic. *Trends in Genetics* **2022**, *38* (6), 613–626.
- (536) Bouchie, A. First microRNA mimic enters clinic. *Nat. Biotechnol.* **2013**, *31* (7), 577.
- (537) What will it take to get miRNA therapies to market? *Nat. Biotechnol.* **2024**, *42* (11), 1623–1624. DOI: .
- (538) A Multicenter Phase I Study of MRX34, MicroRNA miR-RX34 Liposomal Injection. 2013. <https://clinicaltrials.gov/study/NCT01829971?term=NCT01829971&rank=1> (accessed 2026 January 2).
- (539) Pharmacodynamics Study of MRX34, MicroRNA Liposomal Injection in Melanoma Patients With Biopsy Accessible Lesions (MRX34–102). 2016. <https://clinicaltrials.gov/study/NCT02862145?term=NCT02862145&rank=1> (accessed 2026 January 2).
- (540) MesomiR 1: A Phase I Study of TargomiRs as 2nd or 3rd Line Treatment for Patients With Recurrent MPM and NSCLC. 2014.

<https://clinicaltrials.gov/study/NCT02369198?term=NCT02369198&rank=1> (accessed 2026 January 2).

(541) First-in-Human Study of INT-1B3 in Patients With Advanced Solid Tumors. 2020. <https://clinicaltrials.gov/study/NCT04675996?term=NCT04675996&rank=1> (accessed 2026 January 2).

(542) Efficacy, Safety, and Tolerability of Remlarsen (MRG-201) Following Intradermal Injection in Subjects With a History of Keloids. 2018. <https://clinicaltrials.gov/study/NCT03601052?term=NCT03601052&rank=1> (accessed 2026 January 2).

(543) van Zandwijk, N.; Pavlakis, N.; Kao, S. C.; Linton, A.; Boyer, M. J.; Clarke, S.; Huynh, Y.; Chrzanowska, A.; Fulham, M. J.; Bailey, D. L.; Cooper, W. A.; Kritharides, L.; Ridley, L.; Pattison, S. T.; MacDiarmid, J.; Brahmbhatt, H.; Reid, G. Safety and activity of microRNA-loaded minicells in patients with recurrent malignant pleural mesothelioma: a first-in-man, phase 1, open-label, dose-escalation study. *Lancet Oncology* **2017**, *18* (10), 1386–1396.

(544) Viteri, S.; Rosell, R. An innovative mesothelioma treatment based on miR-16 mimic loaded EGFR targeted minicells (TargomiRs). *Translational Lung Cancer Research* **2018**, *7* (Suppl 1), S1–S4.

(545) Chioccioli, M.; Roy, S.; Newell, R.; Pestano, L.; Dickinson, B.; Rigby, K.; Herazo-Maya, J.; Jenkins, G.; Ian, S.; Saini, G.; Johnson, S. R.; Braybrooke, R.; Yu, G.; Sauler, M.; Ahangari, F.; Ding, S.; DeLulius, J.; Aurelien, N.; Montgomery, R. L.; Kaminski, N. A lung targeted miR-29 mimic as a therapy for pulmonary fibrosis. *EBioMedicine* **2022**, *85*, 104304.

(546) ClinicalTrials.gov. <https://clinicaltrials.gov/> (accessed 2025 October 20).

(547) Phase 3 Study of Pelabresib (CPI-0610) in Myelofibrosis (MF) (MANIFEST-2) (MANIFEST-2). 2024. <https://clinicaltrials.gov/study/NCT04603495?term=NCT04603495&rank=1> (accessed 2025 November 6).

(548) Sumitomo Pharma America Announces that Nuvisertib (TP-3654) Has Received FDA Fast Track Designation for the Treatment of Myelofibrosis. <https://news.us.sumitomo-pharma.com/2025-06-12-Sumitomo-Pharma-America-Announces-that-Nuvisertib-TP-3654-Has-Received-FDA-Fast-Track-Designation-for-the-Treatment-of-Myelofibrosis> (accessed 2025 August 13).

(549) Mascarenhas, J.; Kremianskaya, M.; Patriarca, A.; Palandri, F.; Devos, T.; Passamonti, F.; Rampal, R. K.; Mead, A. J.; Hobbs, G.; Scandura, J. M.; Talpaz, M.; Granacher, N.; Somerville, T. C. P.; Hoffman, R.; Wondergem, M. J.; Salama, M. E.; Colak, G.; Cui, J.; Kiladjian, J. J.; Vannucchi, A. M.; Verstovsek, S.; Curto-Garcia, N.; Harrison, C.; Gupta, V. MANIFEST: Pelabresib in Combination With Ruxolitinib for Janus Kinase Inhibitor Treatment-Naive Myelofibrosis. *Journal of Clinical Oncology* **2023**, *41* (32), 4993–5004.

(550) Doroshov, D. B.; Eder, J. P.; LoRusso, P. M. BET Inhibitors: A Novel Epigenetic Approach. *Annals of Oncology* **2017**, *28* (8), 1776–1787.

(551) Pipeline and About Menin Inhibitors. *Kura Oncology*, <https://kuraoncology.com/pipeline/#about-menin-inhibitors> (accessed 2025 August 10).

(552) Breakthrough Therapy Approvals. <https://www.fda.gov/drugs/nda-and-bla-approvals/breakthrough-therapy-approvals> (accessed 2024 March).

(553) First in Human Study of Ziftomenib in Relapsed or Refractory Acute Myeloid Leukemia. 2025. <https://clinicaltrials.gov/study/NCT04067336?term=NCT04067336&rank=1> (accessed 2025 November 6).

(554) Wang, E. S.; Issa, G. C.; Erba, H. P.; Altman, J. K.; Montesinos, P.; DeBotton, S.; Walter, R. B.; Pettit, K.; Savona, M. R.; Shah, M. V.; Kremianskaya, M.; Baer, M. R.; Foran, J. M.; Schiller, G.; Ades, L.; Heiblig, M.; Berthon, C.; Peterlin, P.; Rodriguez-Arboli, E.; Salamero, O.; Patnaik, M. M.; Papayannidis, C.; Grembecka, J.; Cierpicki, T.; Clegg, B.; Ray, J.; Linhares, B. M.; Nie, K.; Mitra, A.; Ahsan, J. M.; Tabachri, M.; Soifer, H. S.; Corum, D.; Leoni, M.; Dale, S.; Fathi, A. T. Ziftomenib in Relapsed or Refractory Acute Myeloid Leukemia (KOMET-001): A Multicentre, Open-label, Multi-cohort, Phase 1 Trial. *Lancet Oncology* **2024**, *25* (10), 1310–1324.

(555) Klossowski, S.; Miao, H.; Kempinska, K.; Wu, T.; Purohit, T.; Kim, E.; Linhares, B. M.; Chen, D.; Jih, G.; Perkey, E.; Huang, H.; He, M.; Wen, B.; Wang, Y.; Yu, K.; Lee, S. C.; Danet-Desnoyers, G.; Trotman, W.; Kandarpa, M.; Cotton, A.; Abdel-Wahab, O.; Lei, H.; Dou, Y.; Guzman, M.; Peterson, L.; Gruber, T.; Choi, S.; Sun, D.; Ren, P.; Li, L. S.; Liu, Y.; Burrows, F.; Maillard, I.; Cierpicki, T.; Grembecka, J. Menin inhibitor MI-3454 Induces Remission in MLL1-rearranged and NPM1-mutated Models of Leukemia. *J. Clin. Invest.* **2019**, *130* (2), 981–997.

(556) A Two-Part Phase 2a Study of RVX000222 in Patients With End-Stage Renal Disease Treated With Hemodialysis. 2023. <https://clinicaltrials.gov/study/NCT03160430?term=NCT03160430&rank=1> (accessed 2025 November 6).

(557) Park, B. Apabetalone Gets Breakthrough Therapy Designation for MACE Prevention. 2020. <https://www.empr.com/home/news/drugs-in-the-pipeline/apabetalone-gets-breakthrough-tx-designation-for-mace-prevention/> (accessed 2025 Aug 10, 2025).

(558) Tsujikawa, L. M.; Fu, L.; Das, S.; Halliday, C.; Rakai, B. D.; Stotz, S. C.; Sarsons, C. D.; Gilham, D.; Daze, E.; Wasiaik, S.; Studer, D.; Rinker, K. D.; Sweeney, M.; Johansson, J. O.; Wong, N. C. W.; Kulikowski, E. Apabetalone (RVX-208) Reduces Vascular Inflammation In vitro and in CVD Patients by a BET-dependent Epigenetic Mechanism. *Clinical Epigenetics* **2019**, *11* (1), 102.

(559) A Phase 2b Study in Subjects With Alcoholic Hepatitis to Evaluate Safety and Efficacy of DUR-928 Treatment (AHFIRM). 2024. <https://clinicaltrials.gov/study/NCT04563026?term=NCT04563026&rank=1> (accessed 2025 November 6).

(560) Corporation, D. Press Release: DURECT Corporation Receives FDA Breakthrough Therapy Designation for Larsucosterol in Alcohol-Associated Hepatitis, 2024.

(561) Wang, Y.; Ren, J.; Ren, S. Larsucosterol: Endogenous Epigenetic Regulator for Treating Chronic and Acute Liver Diseases. *American Journal of Physiology-Endocrinology and Metabolism* **2024**, *326* (5), E577–E587.

(562) Hassanein, T.; McClain, C. J.; Vatsalya, V.; Stein, L. L.; Flamm, S. L.; Martin, P.; Cave, M. C.; Mitchell, M., Jr.; Barton, B.; Nagy, L.; Szabo, G.; McCullough, A.; Dasarthy, S.; Shah, J.; Blevins, C.; Scott, D.; Krebs, W.; Brown, J. E.; Lin, W. Safety, Pharmacokinetics, and Efficacy Signals of Larsucosterol (DUR-928) in Alcohol-Associated Hepatitis. *American Journal of Gastroenterology* **2024**, *119* (1), 107–115.

(563) Study to Test the Efficacy and Safety of Vafidemstat in Adult Borderline Personality Disorder Population. 2023. <https://clinicaltrials.gov/study/NCT04932291?term=NCT04932291&rank=1> (accessed 2025 November 6).

(564) Ferrer, M.; Richarte, V.; Gisbert, L.; Xaus, J.; Gutierrez, S.; Arevalo, M. I.; Ropacki, M.; Bullock, R.; Buesa, C.; Ramos-Quiroga, J. A. REIMAGINE: A Central Nervous System Basket Trial Showing Safety and Efficacy of Vafidemstat on Aggression in Different Psychiatric Disorders. *Psychiatry and Clinical Neurosciences* **2025**, *79* (5), 257–265.

(565) FDA Grants Accelerated Approval to Dordaviprone for Diffuse Midline Glioma. 2025. <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-grants-accelerated-approval-dordaviprone-diffuse-midline-glioma> (accessed 2025 Aug 13, 2025).

(566) Arrillaga-Romany, I.; Gardner, S. L.; Oda, Y.; Aguilera, D.; Allen, J. E.; Batchelor, T.; Butowski, N.; Chen, C.; Cloughesy, T.; Cluster, A.; de Groot, J.; Dixit, K. S.; Graber, J. J.; Haggagi, A. M.; Harrison, R. A.; Kheradpour, A.; Kilburn, L. B.; Kurz, S. C.; Lu, G.; MacDonald, T. J.; Mehta, M.; Melemed, A. S.; Nghiemphu, P. L.; Ramage, S. C.; Shonka, N.; Sumrall, A.; Tarapore, R. S.; Taylor, L.; Umemura, Y.; Wen, P. Y. ONC201 (Dordaviprone) in Recurrent H3 K27M-Mutant Diffuse Midline Glioma. *Journal of Clinical Oncology* **2024**, *42* (13), 1542–1552.

(567) Yang, Z.; Sun, L.; Chen, H.; Sun, C.; Xia, L. New Progress in the Treatment of Diffuse Midline Glioma with H3K27M Alteration. *Heliyon* **2024**, *10* (2), No. e24877.

(568) Arrillaga-Romany, I.; Lassman, A.; McGovern, S. L.; Mueller, S.; Nabors, B.; van den Bent, M.; Vogelbaum, M. A.; Allen, J. E.; Melemed,

- A. S.; Tarapore, R. S.; Wen, P. Y.; Cloughesy, T. ACTION: A Randomized Phase 3 Study of ONC201 (Dordaviprone) in Patients with Newly Diagnosed H3 K27M-Mutant Diffuse Glioma. *Neuro-Oncology* **2024**, *26* (Supplement 2), S173–S181.
- (569) ONC201 in H3 K27M-mutant Diffuse Glioma Following Radiotherapy (the ACTION Study) (ACTION). 2025. <https://clinicaltrials.gov/study/NCT05580562?term=NCT05580562&rank=1> (accessed 2025 November 6).
- (570) FDA Approves Revumenib for Relapsed or Refractory Acute Leukemia with a KMT2A Translocation. 2024.
- (571) A Study of Revumenib in R/R Leukemias Including Those With an MLL/KMT2A Gene Rearrangement or NPM1 Mutation (AUGMENT-101). 2025. <https://clinicaltrials.gov/study/NCT04065399?term=NCT04065399&rank=1> (accessed 2025 November 6).
- (572) Aldoss, I.; Issa, G. C.; Thirman, M.; DiPersio, J.; Arellano, M.; Blachly, J. S.; Mannis, G. N.; Perl, A.; Dickens, D. S.; McMahon, C. M.; Traer, E.; Zwaan, C. M.; Grove, C.; Stone, R.; Shami, P. J.; Mantzaris, I.; Greenwood, M.; Shukla, N.; Cuglievan, B.; Gu, Y.; Bagley, R. G.; Madigan, K.; Sunkaraneni, S.; Nguyen, H. V.; McNeer, N.; Stein, E. M. Revumenib Monotherapy in Patients with Relapsed/Refractory KMT2Ar Acute Leukemia: Topline Efficacy and Safety Results from the Pivotal Augment-101 Phase 2 Study. *Blood* **2023**, *142*, LBA-5.
- (573) Syndax. Syndax Announces FDA Approval of Revuforj® (revumenib) in Adult and Pediatric Patients with Relapsed or Refractory NPM1-Mutated Acute Myeloid Leukemia. 2025.
- (574) Revumenib in Combination With Azacitidine + Venetoclax in Patients NPM1-mutated or KMT2A-rearranged AML. 2025. <https://clinicaltrials.gov/study/NCT06652438?term=NCT06652438&rank=1> (accessed 2025 November 6).
- (575) Evaluation of Revumenib in Participants With Colorectal Cancer and Other Solid Tumors. 2025. <https://clinicaltrials.gov/study/NCT05731947?term=NCT05731947&rank=1> (accessed 2025 November 6).
- (576) Gao, J.; Shi, W.; Wang, J.; Guan, C.; Dong, Q.; Sheng, J.; Zou, X.; Xu, Z.; Ge, Y.; Yang, C.; Li, J.; Bao, H.; Zhong, X.; Cui, Y. Research Progress and Applications of Epigenetic Biomarkers in Cancer. *Frontiers in Pharmacology* **2024**, *15*, 1308309.
- (577) Peedicayil, J. Identification of Biomarkers in Neuropsychiatric Disorders Based on Systems Biology and Epigenetics. *Frontiers in Genetics* **2019**, *10*, 985.
- (578) Gupta, M. K.; Kushwah, A. S.; Singh, R.; Srivastava, K.; Banerjee, M. Genetic and Epigenetic Alterations in MGMT Gene and Correlation with Concomitant Chemoradiotherapy (CRT) in Cervical Cancer. *Journal of Cancer Research and Clinical Oncology* **2023**, *149* (16), 15159–15170.
- (579) Liu, Y.; Ming, H.; Xu, L.; Li, L.; Liu, Q.; Zhao, J.; Zhong, C.; Li, H. DNA Methylation Analysis of the SDC2, SEPT9 and VIM Genes in Fecal DNA for Colorectal Cancer Diagnosis. *BMC Cancer* **2024**, *24* (1), 1205.
- (580) Panagopoulou, M.; Panou, T.; Gkoutakos, A.; Tarapatzi, G.; Karagiani, M.; Tsamardinos, I.; Chatzaki, E. BRCA1 & BRCA2 Methylation as a Prognostic and Predictive Biomarker in Cancer: Implementation in Liquid Biopsy in the Era of Precision Medicine. *Clinical Epigenetics* **2024**, *16* (1), 178.
- (581) Paoli, C.; Misztak, P.; Mazzini, G.; Musazzi, L. DNA Methylation in Depression and Depressive-Like Phenotype: Biomarker or Target of Pharmacological Intervention? *Current Neuropharmacology* **2022**, *20* (12), 2267–2291.
- (582) Shirley, M. Epi proColon((R)) for Colorectal Cancer Screening: A Profile of Its Use in the USA. *Molecular Diagnosis & Therapy* **2020**, *24* (4), 497–503.
- (583) Poon, M. T. C.; Keni, S.; Vimalan, V.; Ip, C.; Smith, C.; Erridge, S.; Weir, C. J.; Brennan, P. M. Extent of MGMT Promoter Methylation Modifies the Effect of Temozolomide on Overall Survival in Patients with Glioblastoma: A Regional Cohort Study. *Neurooncology Advances* **2021**, *3* (1), vdb171.
- (584) Xu, X.; Gammon, M. D.; Zhang, Y.; Bestor, T. H.; Zeisel, S. H.; Wetmur, J. G.; Wallenstein, S.; Bradshaw, P. T.; Garbowski, G.; Teitelbaum, S. L.; Neugut, A. I.; Santella, R. M.; Chen, J. BRCA1 Promoter Methylation is Associated with Increased Mortality Among Women with Breast Cancer. *Breast Cancer Research and Treatment* **2009**, *115* (2), 397–404.
- (585) Ruscito, I.; Dimitrova, D.; Vasconcelos, I.; Gellhaus, K.; Schwachula, T.; Bellati, F.; Zeillinger, R.; Benedetti-Panici, P.; Vergote, I.; Mahner, S.; Cacsire-Tong, D.; Concin, N.; Darb-Esfahani, S.; Lambrechts, S.; Sehouli, J.; Olek, S.; Braicu, E. I. BRCA1 Gene Promoter Methylation Status in High-grade Serous Ovarian Cancer Patients—A Study of the Tumour Bank Ovarian Cancer (TOC) and Ovarian Cancer Diagnosis Consortium (OVCAD). *Eur. J. Cancer* **2014**, *50* (12), 2090–2098.
- (586) Fraga, M. F.; Ballestar, E.; Villar-Garea, A.; Boix-Chornet, M.; Espada, J.; Schotta, G.; Bonaldi, T.; Haydon, C.; Ropero, S.; Petrie, K.; Iyer, N. G.; Perez-Rosado, A.; Calvo, E.; Lopez, J. A.; Cano, A.; Calasanz, M. J.; Colomer, D.; Piris, M. A.; Ahn, N.; Imhof, A.; Caldas, C.; Jenuwein, T.; Esteller, M. Loss of Acetylation at Lys16 and Trimethylation at Lys20 of Histone H4 is a Common Hallmark of Human Cancer. *Nat. Genet.* **2005**, *37* (4), 391–400.
- (587) Plagg, B.; Ehrlich, D.; Kniewallner, K. M.; Marksteiner, J.; Humpel, C. Increased acetylation of histone H4 at lysine 12 (H4K12) in monocytes of transgenic Alzheimer's mice and in human patients. *Current Alzheimer Research* **2015**, *12* (8), 752–760. From NLM Medline
- (588) Ngollo, M.; Lebert, A.; Daures, M.; Judes, G.; Rifai, K.; Dubois, L.; Kemeny, J. L.; Penault-Llorca, F.; Bignon, Y. J.; Guy, L.; Bernard-Gallon, D. Global Analysis of H3K27me3 as an Epigenetic Marker in Prostate Cancer Progression. *BMC Cancer* **2017**, *17* (1), 261.
- (589) Liu, J.; Li, Y.; Liao, Y.; Mai, S.; Zhang, Z.; Liu, Z.; Jiang, L.; Zeng, Y.; Zhou, F.; Xie, D. High Expression of H3K27me3 is an Independent Predictor of Worse Outcome in Patients with Urothelial Carcinoma of Bladder Treated with Radical Cystectomy. *BioMed. Research International* **2013**, *2013*, 390482.
- (590) Tilwala, R.; Nguyen, S. H.; Maurais, A. J.; Nemmara, V. V.; Nagar, M.; Salinger, A. J.; Nagpal, S.; Weerapana, E.; Thompson, P. R. The Rheumatoid Arthritis-Associated Citrullinome. *Cell Chemical Biology* **2018**, *25* (6), 691–704.e6.
- (591) Peng, W.; Wu, S.; Wang, W. Correlation of Serum Citrullinated Histone H3 Levels with Disease Activity in Patients with Rheumatoid Arthritis. *Clinical and Experimental Rheumatology* **2023**, *41* (9), 1792–1800. From NLM Medline
- (592) Hou, J. Y.; Li, N.; Wang, J.; Gao, L. J.; Chang, J. S.; Cao, J. M. Histone crotonylation of peripheral blood mononuclear cells is a potential biomarker for diagnosis of colorectal cancer. *Epigenetics & Chromatin* **2023**, *16* (1), 35. From NLM Medline
- (593) The Predictive Value of Serum Histone Succinylation in Malignant Solid Tumors. <https://ichgcp.net/clinical-trials-registry/NCT07132606> (accessed 2026 January 2).
- (594) Zhu, Y.; Fu, Y.; Liu, F.; Yan, S.; Yu, R. Appraising histone H4 lysine 5 lactylation as a novel biomarker in breast cancer. *Sci. Rep.* **2025**, *15* (1), 8205. From NLM Medline.
- (595) Bautista-Sanchez, D.; Arriaga-Canon, C.; Pedroza-Torres, A.; De La Rosa-Velazquez, I. A.; Gonzalez-Barrios, R.; Contreras-Espinosa, L.; Montiel-Manriquez, R.; Castro-Hernandez, C.; Fragos-Ontiveros, V.; Alvarez-Gomez, R. M.; Herrera, L. A. The Promising Role of miR-21 as a Cancer Biomarker and Its Importance in RNA-Based Therapeutics. *Molecular Therapy Nucleic Acids* **2020**, *20*, 409–420.
- (596) Shao, C.; Yang, F.; Qin, Z.; Jing, X.; Shu, Y.; Shen, H. The Value of miR-155 as a Biomarker for the Diagnosis and Prognosis of Lung Cancer: A Systematic Review with Meta-analysis. *BMC Cancer* **2019**, *19* (1), 1103. From NLM Medline
- (597) Todén, S.; Goel, A. Non-coding RNAs as Liquid Biopsy Biomarkers in Cancer. *Br. J. Cancer* **2022**, *126* (3), 351–360.
- (598) Wang, J.; Xu, L.; Tian, L.; Sun, Q. Circulating microRNA-208 Family as Early Diagnostic Biomarkers for Acute Myocardial Infarction: A Meta-analysis. *Medicine (Baltimore)* **2021**, *100* (51), No. e27779.
- (599) Arshi, A.; Raeisi, F.; Mahmoudi, E.; Mohajerani, F.; Kabiri, H.; Fazel, R.; Zabihian-Langeroudi, M.; Jusic, A. A Comparative Study of HOTAIR Expression in Breast Cancer Patient Tissues and Cell Lines. *Cell* **2020**, *22* (2), 178–184.

- (600) Chen, S.; Zhang, C.; Feng, M. Prognostic Value of LncRNA HOTAIR in Colorectal Cancer: A Meta-analysis. *Open Medicine (Warsaw, Poland)* **2020**, *15*, 76–83.
- (601) Nazari, M.; Babakhanzadeh, E.; Mollazadeh, A.; Ahmadvade, M.; Mohammadi Soleimani, E.; Hajimaqsoudi, E. HOTAIR in Cancer: Diagnostic, Prognostic, and Therapeutic Perspectives. *Cancer Cell International* **2024**, *24* (1), 415.
- (602) Hao, F.; Zhang, Y.; Hou, J.; Zhao, B. Chromatin remodeling and cancer: the critical influence of the SWI/SNF complex. *Epigenetics & Chromatin* **2025**, *18* (1), 22.
- (603) Hsu, H. K.; Weng, Y. I.; Hsu, P. Y.; Huang, T. H.; Huang, Y. W. Detection of DNA Methylation by MeDIP and MBDCap Assays: An Overview of Techniques. *Methods Mol. Biol.* **2020**, *2102*, 225–234.
- (604) Meissner, A.; Gnirke, A.; Bell, G. W.; Ramsahoye, B.; Lander, E. S.; Jaenisch, R. Reduced Representation Bisulfite Sequencing for Comparative High-resolution DNA Methylation Analysis. *Nucleic Acids Res.* **2005**, *33* (18), 5868–5877.
- (605) Kelsey, G.; Stegle, O.; Reik, W. Single-cell Epigenomics: Recording the Past and Predicting the Future. *Science* **2017**, *358* (6359), 69–75.
- (606) Preissl, S.; Gaulton, K. J.; Ren, B. Characterizing Cis-regulatory Elements Using Single-cell Epigenomics. *Nat. Rev. Genet.* **2023**, *24* (1), 21–43.
- (607) Schwartzman, O.; Tanay, A. Single-cell Epigenomics: Techniques and Emerging Applications. *Nat. Rev. Genet.* **2015**, *16* (12), 716–726.
- (608) Yu, F.; Sankaran, V. G.; Yuan, G. C. CUT&RUNTools 2.0: A Pipeline for Single-cell and Bulk-level CUT&RUN and CUT&Tag Data Analysis. *Bioinformatics* **2021**, *38* (1), 252–254.
- (609) Lajoie, B. R.; Dekker, J.; Kaplan, N. The Hitchhiker's Guide to Hi-C Analysis: Practical Guidelines. *Methods* **2015**, *72*, 65–75.
- (610) Chen, C.; Wang, J.; Pan, D.; Wang, X.; Xu, Y.; Yan, J.; Wang, L.; Yang, X.; Yang, M.; Liu, G. P. Applications of Multi-Omics Analysis in Human Diseases. *MedComm* (2020) **2023**, *4* (4), No. e315.
- (611) Ruprecht, N. A.; Kennedy, J. D.; Bansal, B.; Singhal, S.; Sens, D.; Maggio, A.; Doe, V.; Hawkins, D.; Campbell, R.; O'Connell, K.; Gill, J. S.; Schaefer, K.; Singhal, S. K. Transcriptomics and Epigenetic Data Integration Learning Module on Google Cloud. *Briefings in Bioinformatics* **2024**, *25* (Supplement_1), DOI: 10.1093/bib/bbae352.
- (612) Josipovic, G.; Tadic, V.; Klasic, M.; Zanki, V.; Beccheli, I.; Chung, F.; Ghantous, A.; Keser, T.; Madunic, J.; Boskovic, M.; Lauc, G.; Herceg, Z.; Vojta, A.; Zoldos, V. Antagonistic and Synergistic Epigenetic Modulation Using Orthologous CRISPR/dCas9-based Modular System. *Nucleic Acids Res.* **2019**, *47* (18), 9637–9657.
- (613) Policarpi, C.; Munafo, M.; Tsagkris, S.; Carlini, V.; Hackett, J. A. Systematic Epigenome Editing Captures the Context-dependent Instructive Function of Chromatin Modifications. *Nat. Genet.* **2024**, *56* (6), 1168–1180.
- (614) Kubik, G.; Summerer, D. TALEdred Epigenetics: A DNA-Binding Scaffold for Programmable Epigenome Editing and Analysis. *Chembiochem* **2016**, *17* (11), 975–980.
- (615) Waryah, C. B.; Moses, C.; Arooj, M.; Blancafort, P. Zinc Fingers, TALEs, and CRISPR Systems: A Comparison of Tools for Epigenome Editing. *Methods Mol. Biol.* **2018**, *1767*, 19–63.
- (616) Zhang, R.; Yao, T.; Fan, M.; Jiang, X.; Wang, K.; Cui, M.; Bing, K.; Xia, X. Precision Scalpels for the Epigenome: Next-gen Editing Tools in Targeted Therapies. *Frontiers in Medicine* **2025**, *12*, 1613722.
- (617) Xu, J.; Ma, H.; Liu, Y. Stochastic Optical Reconstruction Microscopy (STORM). *Current Protocols in Cytometry* **2017**, *81* (1), 12.46.1–12.46.27.
- (618) Lippincott-Schwartz, J.; Manley, S.; Burnette, D.; Gillette, J.; Patterson, G. PALM-Based Super-Resolution Imaging and its Applications. *Biophys. J.* **2010**, *98*, 619A.
- (619) Barth, R.; Bystricky, K.; Shaban, H. A. Coupling Chromatin Structure and Dynamics by Live Super-Resolution Imaging. *Science Advances* **2020**, *6* (27), DOI: 10.1126/sciadv.aaz2196.
- (620) Mehra, D.; Adhikari, S.; Banerjee, C.; Puchner, E. M. Characterizing Locus Specific Chromatin Structure and Dynamics with Correlative Conventional and Super-resolution Imaging in Living Cells. *Nucleic Acids Res.* **2022**, *50* (13), No. e78.
- (621) Lucas, M. C.; Novoa, E. M. Long-read Sequencing in the Era of Epigenomics and Epitranscriptomics. *Nat. Methods* **2023**, *20* (1), 25–29.
- (622) Liu, T.; Conesa, A. Profiling the Epigenome Using Long-read Sequencing. *Nat. Genet.* **2025**, *57* (1), 27–41.
- (623) Santalo, J.; Berdasco, M. Ethical Implications of Epigenetics in the Era of Personalized Medicine. *Clinical Epigenetics* **2022**, *14* (1), 44.
- (624) Dupras, C.; Knoppers, T.; Palmour, N.; Beauchamp, E.; Liosi, S.; Siebert, R.; Berner, A. M.; Beck, S.; Charest, I.; Joly, Y. Researcher Perspectives on Ethics Considerations in Epigenetics: An International Survey. *Clinical Epigenetics* **2022**, *14* (1), 110.
- (625) Bunnik, E. M.; Timmers, M.; Bolt, I. L. Ethical Issues in Research and Development of Epigenome-wide Technologies. *Epigenetics Insights* **2020**, *13*, 2516865720913253.
- (626) *Epigenetics: Ethical, Legal and Social Aspects*; Springer VS: Wiesbaden, 2017. DOI: 10.1007/978-3-658-14460-9.
- (627) Dalpe, G.; Huerne, K.; Dupras, C.; Cheung, K.; Palmour, N.; Winkler, E.; Alex, K.; Mehlman, M.; Holloway, J. W.; Bunnik, E.; Konig, H.; Mansuy, I. M.; Rots, M. G.; Erwin, C.; Erler, A.; Libertini, E.; Joly, Y. Defusing the Legal and Ethical Minefield of Epigenetic Applications in the Military, Defense, and Security Context. *Journal of Law and the Biosciences* **2023**, *10* (2), lsad034.
- (628) Chiapperino, L. Epigenetics: Ethics, Politics, Biosociality. *British Medical Bulletin* **2018**, *128* (1), 49–60.
- (629) Cusenza, V. Y.; Tameni, A.; Neri, A.; Frazzi, R. The lncRNA epigenetics: The significance of m6A and m5C lncRNA modifications in cancer. *Frontiers in Oncology* **2023**, *13*, 1063636.
- (630) Li, J.; Wang, X.; Wang, H. RNA modifications in long non-coding RNAs and their implications in cancer biology. *Bioorg. Med. Chem.* **2024**, *113*, 117922.
- (631) Yang, L.; Tang, L.; Min, Q.; Tian, H.; Li, L.; Zhao, Y.; Wu, X.; Li, M.; Du, F.; Chen, Y.; Li, W.; Li, X.; Chen, M.; Gu, L.; Sun, Y.; Xiao, Z.; Shen, J. Emerging role of RNA modification and long noncoding RNA interaction in cancer. *Cancer Gene Ther.* **2024**, *31* (6), 816–830.
- (632) Alaei, A.; Kakumani, P. K. MicroRNA chemical modifications in post-transcriptional gene silencing and human diseases. *Molecular Therapy Nucleic Acids* **2025**, *36* (4), 102745.
- (633) Sighel, D.; Destefanis, E.; Quattrone, A. Therapeutic strategies to target the epitranscriptomic machinery. *Current Opinion in Genetics & Development* **2024**, *87*, 102230.
- (634) Kayzuka, C.; Rondon-Pereira, V. C.; Nogueira Tavares, C.; Pacheco Pachado, M.; Monica, F. Z.; Tanus-Santos, J. E.; Lacchini, R. Epigenetics is involved in the pleiotropic effects of statins. *Expert Opinion on Drug Metabolism & Toxicology* **2025**, *21* (6), 689–701.