

# The complete mitochondrial genome of *Glyptotendipes tokunagai* (Arthropoda; Insecta; Diptera; Chironomidae)

Xinyue Huang,<sup>1</sup> Dihao Huang,<sup>1</sup> Hengying Chen,<sup>1</sup> Chunmei Zhu,<sup>1</sup> Wenhao Wang,<sup>1</sup> Shan Ouyang,<sup>1</sup> Xiaoping Wu,<sup>1</sup> Chunhua Zhou<sup>1</sup>

**AUTHOR AFFILIATION** See affiliation list on p. 3.

**ABSTRACT** We present the complete mitochondrial genome sequence of *Glyptotendipes tokunagai* in this study. This circular genome encompasses 15,241 base pairs and exhibits a high AT content of 77%. It contains a total of 13 protein-coding genes, 22 transfer RNA (tRNA) genes, and 2 ribosomal RNA (rRNA) genes.

**KEYWORDS** *Glyptotendipes tokunagai*, mitochondrial genome, China

*Glyptotendipes tokunagai* is an aquatic insect of the family Chironomidae (Diptera), with larvae inhabiting aquatic ecosystem sediments (1). As a representative Chironomidae species, it is closely linked to freshwater quality, serving as a key bioindicator for freshwater ecosystem assessment and widely used in aquatic environmental protection research (2). This study reports its complete mitochondrial genome, providing a valuable basis for advancing systematics, population genetics, and database construction.

The sample was collected from Poyang Lake (GPS coordinates: 29.253°N, 116.190°E) in 2023 using a Peterson grab sampler (1/16 m<sup>2</sup>). Sediment samples were washed through a 40-mesh sieve, and macroinvertebrates were manually selected from a white porcelain dish. The specimens were preserved in absolute ethanol at low temperature (4°C). Genomic DNA was extracted from a single, whole specimen using the Rapid Animal Genomic DNA Isolation Kit. The library was constructed with the Hieff NGSMaxUp II DNA Library Prep Kit for Illumina. Sequencing was carried out on the Illumina NovaSeq 6000 platform, generating paired-end reads of 150 bp in length. A total of 49,016,436 raw reads were generated, corresponding to 7,352 Mbp of sequencing data with an average coverage depth of 1,371×. Quality control was conducted using fastp v0.36 (3) to remove adapter sequences, low-quality reads (where >40% of bases had a quality score < Q15), and paired-end reads shorter than 35 nt. To assess potential contamination, 10,000 randomly selected reads were aligned to the NCBI NT database using blastn, with *Chironomus tepperi* (JN861749.1) as the top-matching species (4). *De novo* assembly was carried out with SPAdes v3.15 (5), with GapFiller v1.11 (6) for gap closure and Pilon v1.24 (7) for base-error correction. Gene annotation was performed using MITOS v1.1.7 with the Invertebrate Mitochondrial Code (8). We used the homology with the reference genome to determine the most likely start and stop codons, and the determination process was clear. Manual curation refined gene boundaries through NCBI-BLAST comparisons with reference mitochondrial genome of *Glyptotendipes caulicola* (NC\_016167.1). The assembled genome showed 87% identity to the genome of *G. caulicola*.

The complete mitochondrial genome was 15,241 bp (GenBank accession number PV781185). Protein-coding genes have 77% AT and 23% CG content, showing significant AT bias. It included 13 PCGs, 2 ribosomal RNA (rRNA) genes (12S and 16S), and 22 transfer RNA (tRNA) genes (Table 1). PCGs used diverse start codons: ATG (COX2, ATP6, COX3, ND4, ND4L, and CYTB), ATT (ND2, ATP8, ND3, ND6, and ND1), TTG (COX1), and GTG (ND5).

**Editor** Julie C. Dunning Hotopp, University of Maryland School of Medicine, Baltimore, Maryland, USA

Address correspondence to Chunhua Zhou, zhouchunhua@ncu.edu.cn, or Xiaoping Wu, xpwu@ncu.edu.cn.

The authors declare no conflict of interest.

See the funding table on p. 3.

**Received** 2 October 2025

**Accepted** 19 December 2025

**Published** 3 February 2026

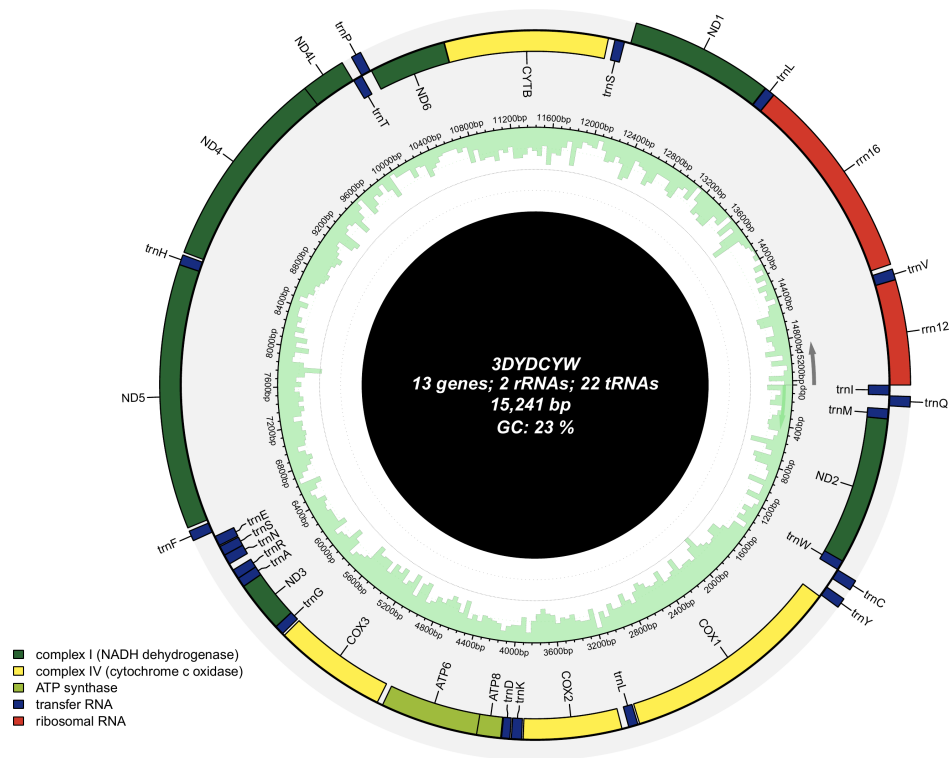
Copyright © 2026 Huang et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

**TABLE 1** Mitochondrial genome content, organization, and codon information of *Glyptotendipes tokunagai*

| Gene   | Type | Minimum nucleotide position | Maximum nucleotide position | Length | Start codon | Stop codon | H/L strand <sup>a</sup> |
|--------|------|-----------------------------|-----------------------------|--------|-------------|------------|-------------------------|
| trnI   | tRNA | 1                           | 68                          | 68     | –           | –          | –                       |
| trnQ   | tRNA | 74                          | 142                         | 69     | –           | –          | –                       |
| trnM   | tRNA | 164                         | 232                         | 69     | –           | –          | –                       |
| ND2    | CDS  | 233                         | 1261                        | 1,029  | ATT         | TAA        | –                       |
| trnW   | tRNA | 1260                        | 1327                        | 68     | –           | –          | –                       |
| trnC   | tRNA | 1329                        | 1398                        | 70     | –           | –          | +                       |
| trnY   | tRNA | 1468                        | 1534                        | 67     | –           | –          | +                       |
| COX1   | CDS  | 1548                        | 3083                        | 1,536  | TTG         | TAA        | –                       |
| trnL   | tRNA | 3096                        | 3161                        | 66     | –           | –          | –                       |
| COX2   | CDS  | 3209                        | 3892                        | 684    | ATG         | TAA        | –                       |
| trnK   | tRNA | 3903                        | 3974                        | 72     | –           | –          | –                       |
| trnD   | tRNA | 3984                        | 4041                        | 58     | –           | –          | –                       |
| ATP8   | CDS  | 4047                        | 4220                        | 174    | ATT         | TAA        | –                       |
| ATP6   | CDS  | 4214                        | 4891                        | 678    | ATG         | TAA        | –                       |
| COX3   | CDS  | 4931                        | 5719                        | 789    | ATG         | TAA        | –                       |
| trnG   | tRNA | 5736                        | 5801                        | 66     | –           | –          | –                       |
| ND3    | CDS  | 5802                        | 6155                        | 354    | ATT         | TAA        | –                       |
| trnA   | tRNA | 6156                        | 6222                        | 67     | –           | –          | –                       |
| trnR   | tRNA | 6223                        | 6289                        | 67     | –           | –          | –                       |
| trnN   | tRNA | 6337                        | 6406                        | 70     | –           | –          | –                       |
| trnS   | tRNA | 6407                        | 6473                        | 67     | –           | –          | –                       |
| trnE   | tRNA | 6482                        | 6550                        | 69     | –           | –          | –                       |
| trnF   | tRNA | 6585                        | 6652                        | 68     | –           | –          | +                       |
| ND5    | CDS  | 6675                        | 8411                        | 1,737  | GTG         | TAA        | +                       |
| trnH   | tRNA | 8412                        | 8478                        | 67     | –           | –          | +                       |
| ND4    | CDS  | 8497                        | 9834                        | 1,338  | ATG         | TAA        | +                       |
| ND4L   | CDS  | 9828                        | 10121                       | 294    | ATG         | TAA        | +                       |
| trnT   | tRNA | 10126                       | 10190                       | 65     | –           | –          | –                       |
| trnP   | tRNA | 10191                       | 10257                       | 67     | –           | –          | +                       |
| ND6    | CDS  | 10265                       | 10798                       | 534    | ATT         | TAA        | –                       |
| CYTB   | CDS  | 10801                       | 11937                       | 1,137  | ATG         | TAA        | –                       |
| trnS   | tRNA | 11983                       | 12050                       | 68     | –           | –          | –                       |
| ND1    | CDS  | 12093                       | 13031                       | 939    | ATT         | TAA        | +                       |
| trnL   | tRNA | 13035                       | 13101                       | 67     | –           | –          | +                       |
| l-rRNA | rRNA | 13103                       | 14447                       | 1,345  | –           | –          | +                       |
| trnV   | tRNA | 14480                       | 14551                       | 72     | –           | –          | +                       |
| s-rRNA | rRNA | 14552                       | 15241                       | 690    | –           | –          | +                       |

<sup>a</sup>+, heavy (H) strand; –, light (L) strand.

ND5, ND4, ND4L, and ND1 were on the heavy strand; COX2, ATP6, COX3, CYTB, ND2, ATP8, ND3, ND6, and COX1 on the light strand. The circular mitomap was visualized via Chloroplast (9), showing transcription directions, gene locations, and GC content (Fig. 1).



**FIG 1** Mitochondrial genome map of *Glyptotendipes tokunagai*. The direction of transcription is indicated by arrows. The color coding corresponds to different genome groups, and the specific information is shown in the legend at the bottom left corner. The outermost ring displays the genes, and the innermost ring shows the GC content in light green. Genes transcribed clockwise are located on the inside of the circle, while genes transcribed counterclockwise are on the outside.

ACKNOWLEDGMENTS

This work was funded by the Natural Science Foundation of Jiangxi Province (20232BAB205060) and the "Science and Technology and Water Conservancy" joint plan project of Jiangxi Province (2023KSG01006).

We thank Zengsheng Yue at Nanchang University, China, for helping with the collection work. We also thank Wenxin Xia and Ruobing Zhao at Nanchang University, China, for their help with the lab work.

AUTHOR AFFILIATION

<sup>1</sup>School of Life Sciences, Nanchang University, Nanchang, China

AUTHOR ORCIDs

Xinyue Huang <https://orcid.org/0009-0004-8046-4899>

Chunhua Zhou <http://orcid.org/0000-0002-3053-1270>

FUNDING

| Funder                                         | Grant(s)       | Author(s)    |
|------------------------------------------------|----------------|--------------|
| Natural Science Foundation of Jiangxi Province | 20232BAB205060 | Chunhua Zhou |

DATA AVAILABILITY

The mitochondrial genome of *Glyptotendipes tokunagai* has been completely sequenced and deposited in GenBank with accession number [PV781185](https://www.ncbi.nlm.nih.gov/nuccore/PV781185). The reads deposited are for

the whole organism. For this genome, the corresponding BioProject, SRA, and BioSample accessions are [PRJNA1291952](https://doi.org/10.1089/cmb.2012.0021), [SRR34564647](https://doi.org/10.1186/gb-2012-13-6-r56), and [SAMN49982289](https://doi.org/10.1371/journal.pone.0112963), respectively.

## REFERENCES

1. Xu ZG, Li CR, Wang JX, Yu P, Xiao YL, Fu Y. 2024. Investigation on diversity of *Chironomidae* in Dabie Mountains. *Journal of Environmental Entomology* 46:606–615. <https://doi.org/10.3969/j.issn.1674-0858.2024.03.7>
2. Hirabayashi K, Yoshizawa K, Yoshida N, Kazama F. 2004. Progress of eutrophication and change of chironomid fauna in Lake Yamanakako, Japan. *Limnology* 5:47–53. <https://doi.org/10.1007/s10201-003-0113-2>
3. Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>
4. Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 25:3389–3402. <https://doi.org/10.1093/nar/25.17.3389>
5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>
6. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with GapFiller. *Genome Biol* 13:R56. <https://doi.org/10.1186/gb-2012-13-6-r56>
7. Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>
8. Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsch G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. *Mol Phylogenet Evol* 69:313–319. <https://doi.org/10.1016/j.ympev.2012.08.023>
9. Zheng S, Pocza P, Hyvönen J, Tang J, Amiryousefi A. 2020. Chloroplot: an online program for the versatile plotting of organelle genomes. *Front Genet* 11:576124. <https://doi.org/10.3389/fgene.2020.576124>