

Hybrid assembled genomes of three *Salmonella enterica* isolates from municipal wastewater in Tennessee, USA

Jia Wang,¹ Lauren K. Hudson,¹ Daniel W. Bryan,¹ Thomas G. Denes¹

AUTHOR AFFILIATION See affiliation list on p. 2.

ABSTRACT Three *Salmonella enterica* isolates were isolated from a municipal wastewater treatment plant in Knoxville, TN, USA. Genomic DNA was extracted and sequenced on the Illumina and Oxford Nanopore platforms. Annotated hybrid assemblies showed the strains were predicted to be serotypes Bareilly ($n = 2$) and Typhimurium ($n = 1$).

KEYWORDS *Salmonella*, wastewater, surveillance

Salmonella enterica is a globally widespread bacterial pathogen, causing an estimated 100 million infections and over 200,000 deaths annually (1, 2). Reporting novel *S. enterica* strains from municipal wastewater contributes genomic resources for comparative analyses and genomic surveillance. This study presents the complete and draft genome sequences of three *S. enterica* strains (UTK W1-0001, UTK W1-0003, and UTK W1-0011) isolated from municipal wastewater samples collected at a local wastewater treatment plant in Knoxville, Tennessee, USA.

Samples were collected from the primary effluent of municipal wastewater on 9 October 2023. The collected wastewater samples were filtered and enriched for *Salmonella* spp. following the modified Food and Drug Administration's Bacteriological Analytical Manual protocol (3, 4). Single colonies were isolated on selective media, and pure cultures were prepared. The presumptive *Salmonella* isolates were cultured overnight in brain heart infusion broth at 37°C with shaking at 200 rpm. DNA was extracted by the DNeasy Blood and Tissue Kit (Qiagen), assessed with Nanodrop (Thermo Fisher Scientific) and Qubit 4 (Thermo Fisher Scientific), and shipped to SeqCenter (Pittsburgh, PA, USA). Illumina short-read sequencing was performed using the Illumina DNA Prep kit and NovaSeq X Plus sequencer (Illumina), producing 151 bp paired-end reads (5). For Nanopore sequencing, DNA was extracted using the Zymo Quick-DNA HMW MagBead kit (Zymo Research Corp.). The library was prepared using the Oxford Nanopore rapid barcoding kit (SQK-RBK114.24) and sequenced with a MinION sequencer with Flongle (Oxford Nanopore Technologies). Basecalling was performed by MinkNOW (v24.06.5) using the fast basecalling model with adapter trimming, default quality score threshold, and no size-selection (6). Read quality was checked by FastQC v0.11.9 (7), and trimming was performed by Trimmomatic v0.39 (8). The hybrid assembly was performed using Unicycler v0.4.8 in bold mode, including overlap identification and trimming, with rotation to start at dnaA/repA (9). For all software, default parameters were used except where otherwise noted.

BLAST analysis (10) of the 7 kb contig in UTK W1-0011 assembly revealed 96.7% nucleotide identity across 81% of the query sequence length to plasmid NZ_CP128883.1. kSNP4 (v4.1) was used to identify core genome single-nucleotide polymorphism (SNP) among these three *Salmonella* isolates (11), revealing each was a distinct strain. Genome sequencing data and annotation features are summarized in Table 1. The genomes of strains UTK W1-0001, UTK W1-0003, and UTK W1-0011 were predicted to have 151–152 virulence factor genes. Key virulence determinants included fimbrial and

Editor Vincent Michael Bruno, University of Maryland School of Medicine, Baltimore, Maryland, USA

Address correspondence to Thomas G. Denes, tdenes@utk.edu.

The authors declare no conflict of interest.

Received 25 November 2024

Accepted 28 April 2025

Published 27 June 2025

Copyright © 2025 Wang et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Genomic sequence statistics and features of three *S. enterica* isolates from a local wastewater treatment plant

Strain	No. of Illumina reads	No. of ONT reads	N50 of ONT reads	Total length (bp) ^a	No. of chromosome contigs ^a	No. of plasmid contigs ^a	GC content (%) ^a	Predicted serotype ^b	Illumina read coverage (x) ^c	Nanopore read coverage (x) ^c	No. of CDSs ^d	No. of tRNAs ^d	No. of CRISPR arrays ^e	Antibiotic resistance genes ^f
UTK W1-0001	2,419,110	5,000	19,045	4,964,787	1	1	52.17	Typhimurium	66	9	4,739	86	4	<i>aac(6)-laa</i> , <i>aph(6)-Id</i> , <i>blaCMY-2</i> , <i>sul2</i> , <i>tet(A)</i> , <i>floR</i>
UTK W1-0003	3,066,142	8,000	17,340	4,843,513	1	1	52.05	Bareilly	90	14	4,648	85	3	<i>aac(6)-laa</i>
UTK W1-0011	2,943,720	7,000	17,533	5,032,162	1	2	52.14	Bareilly	83	12	4,871	86	2	<i>aac(6)-laa</i>

^aAssembly statistics and quality were generated using QUAST v5.2.0 (16) and CheckM v1.1.3 (17).^bThe species and serotype were analyzed by Enterobase v1.2.0 (18).^cAverage read coverage was determined using Minimap2 v2.14 (19), BMAP v38.63 (20), and SAMtools v1.15.1 (21).^dThe assemblies were annotated by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP, v6.8) (22).^eThe clustered regularly interspaced short palindromic repeat (CRISPR) arrays were predicted using CRISPRFinder (23, 24).^fAntimicrobial resistance and virulence genes were screened using ResFinder v4.6.0 (25) and Virulence Factor Database (VFDB) (26).

nonfimbrial adherence determinants, macrophage inducible gene *mig-14*, and type III secretion system (T3SS) encoded by *Salmonella* pathogenicity island 1 (SPI-1) and 2 (SPI-2). Compared with previously isolated *Salmonella enterica* strains harboring 100–130 virulence genes identified by VFDB (12–15), the three newly isolated *Salmonella enterica* strains exhibit a notable virulence profile potentially indicative of aggressive pathogenic characteristics.

ACKNOWLEDGMENTS

This study was supported by multistate project S1077, “Enhancing Microbial Food Safety by Risk Analysis,” by Epidemiology and Laboratory Capacity for Infectious Diseases (grant number NU50CK000528), and by The University of Tennessee Institute of Agriculture AgResearch.

We thank the sample provider for granting permission and guidance for sample collection.

AUTHOR AFFILIATION

¹Department of Food Science, The University of Tennessee, Knoxville, Tennessee, USA

AUTHOR ORCIDs

Jia Wang  <https://orcid.org/0000-0001-8709-9709>

Lauren K. Hudson  <http://orcid.org/0000-0002-1799-3171>

Thomas G. Denes  <http://orcid.org/0000-0002-2631-0846>

DATA AVAILABILITY

The genomic sequencing data and assemblies for strains UTK W1-0001, UTK W1-0003 and UTK W1-0011 have been deposited in the NCBI database under BioProject number [PRJNA1169800](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1169800). The corresponding BioSample numbers are [SAMN44082895](https://www.ncbi.nlm.nih.gov/biosample/SAMN44082895), [SAMN44082896](https://www.ncbi.nlm.nih.gov/biosample/SAMN44082896), and [SAMN44082897](https://www.ncbi.nlm.nih.gov/biosample/SAMN44082897); SRA accession numbers are [SRR30900535](https://www.ncbi.nlm.nih.gov/sra/SRR30900535), [SRR30900538](https://www.ncbi.nlm.nih.gov/sra/SRR30900538), [SRR30900534](https://www.ncbi.nlm.nih.gov/sra/SRR30900534), [SRR30900537](https://www.ncbi.nlm.nih.gov/sra/SRR30900537), [SRR30900533](https://www.ncbi.nlm.nih.gov/sra/SRR30900533), and [SRR30900536](https://www.ncbi.nlm.nih.gov/sra/SRR30900536). Assembly accession numbers are [CP171353.1](https://www.ncbi.nlm.nih.gov/assembly/CP171353.1), [CP171354.1](https://www.ncbi.nlm.nih.gov/assembly/CP171354.1), [CP171351.1](https://www.ncbi.nlm.nih.gov/assembly/CP171351.1), [CP171352.1](https://www.ncbi.nlm.nih.gov/assembly/CP171352.1), and [JBJDRN000000000.1](https://www.ncbi.nlm.nih.gov/assembly/JBJDRN000000000.1).

REFERENCES

- Wang BX, Butler DSC, Hamblin M, Monack DM. 2023. One species, different diseases: the unique molecular mechanisms that underlie the pathogenesis of typhoidal *Salmonella* infections. *Curr Opin Microbiol* 72:102262. <https://doi.org/10.1016/j.mib.2022.102262>

2. Sears KT, Galen JE, Tennant SM. 2021. Advances in the development of *Salmonella*-based vaccine strategies for protection against Salmonellosis in humans. *J Appl Microbiol* 131:2640–2658. <https://doi.org/10.1111/jam.15055>
3. Strawn LK, Gröhn YT, Warchocki S, Worobo RW, Bihn EA, Wiedmann M. 2013. Risk factors associated with *Salmonella* and *Listeria monocytogenes* contamination of produce fields. *Appl Environ Microbiol* 79:7618–7627. <https://doi.org/10.1128/AEM.02831-13>
4. Strawn LK, Fortes ED, Bihn EA, Nightingale KK, Gröhn YT, Worobo RW, Wiedmann M, Bergholz PW. 2013. Landscape and meteorological factors affecting prevalence of three food-borne pathogens in fruit and vegetable farms. *Appl Environ Microbiol* 79:588–600. <https://doi.org/10.1128/AEM.02491-12>
5. Wang J, Schamp CN, Hudson LK, Chaggar HK, Bryan DW, Garman KN, Radosevich MA, Denes TG. 2025. Whole-genome sequencing and metagenomics reveals diversity and prevalence of soil *Listeria* spp. in the Nantahala National Forest. *Microbiol Spectr* 13:e0171224. <https://doi.org/10.1128/spectrum.01712-24>
6. Technologies ON. 2025. MinkNOW. Available from: <https://nanoporetech.com/document/experiment-companion-minknow>
7. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data, on Babraham Bioinformatics, Babraham Institute, Cambridge, United Kingdom. Available from: <https://www.bioinformatics.babraham.ac.uk/projects/fastqc>
8. Bolger AM, Lohse N, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
9. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLOS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
10. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
11. Hall BG, Nisbet J. 2023. Building phylogenetic trees from genome sequences with kSNP4. *Mol Biol Evol* 40:msad235. <https://doi.org/10.1093/molbev/msad235>
12. Tiengo A, Orsini M, Petrin S, Losasso C, Longo A, Cento G, Ciot L, Ceruti R, Cibi V, Barco L. 2023. Whole-genome sequence of *Salmonella enterica* serovar Bispebjerg from Turkey reveals its pathogenic potential. *Microbiol Resour Announc* 12:e0004323. <https://doi.org/10.1128/mra.00043-23>
13. Popy NN, Hoque MN, Khan MFR, Biswas L, Rahman MH, Saiduzzaman M, Rahman M, Rahman MB. 2024. Draft genome sequencing of a multidrug-resistant *Salmonella enterica* subspecies *enterica* serovar Typhimurium strain isolated from chicken in Bangladesh. *Microbiol Resour Announc* 13:e0061923. <https://doi.org/10.1128/mra.00619-23>
14. Schlund O, Göhler A, Borowiak M, Deneke C, Fischer M, Lamparter MC. 2022. Complete genome sequence of a *Salmonella enterica* subsp. *enterica* serovar Tennessee strain from Tahiti. *Microbiol Resour Announc* 11:e0040722. <https://doi.org/10.1128/mra.00407-22>
15. Helmy Yosra A, Kabir A, Saleh M, Kennedy Laura A, Burns L, Johnson B. 2024. Draft genome sequence analysis of multidrug-resistant *Salmonella enterica* subsp. *enterica* serovar Mbandaka harboring colistin resistance gene mcr-9.1 isolated from foals in Kentucky, USA. *Microbiol Resour Announc* 13:e00737–24. <https://doi.org/10.1128/mra.00737-24>
16. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
17. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
18. Zhou Z, Alikhan N-F, Mohamed K, Fan Y, Achtman M, Brown D, Chattaway M, Dallman T, Delahay R, Kornschöber C. 2020. The Enterobase user's guide, with case studies on *Salmonella* transmissions, *Yersinia pestis* phylogeny, and *Escherichia* core genomic diversity. *Genome Res* 30:138–152.
19. Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34:3094–3100. <https://doi.org/10.1093/bioinformatics/bty191>
20. Bushnell B. 2015. *BBMap short-read aligner, and other bioinformatics tools*. University of California, Berkeley, CA.
21. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, Genome Project Data Processing S. 2009. The sequence alignment/map format and SAMtools. *Bioinformatics* 25:2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>
22. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
23. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. *Nucleic Acids Res* 35:W52–W57. <https://doi.org/10.1093/nar/gkm360>
24. Couvin D, Bernheim A, Toffano-Nioche C, Touchon M, Michalik J, Néron B, Rocha EPC, Vergnaud G, Gautheret D, Pourcel C. 2018. CRISPRCas-Finder, an update of CRISPRFinder, includes a portable version, enhanced performance and integrates search for Cas proteins. *Nucleic Acids Res* 46:W246–W251. <https://doi.org/10.1093/nar/gky425>
25. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <https://doi.org/10.1093/jac/dks261>
26. Chen L, Zheng D, Liu B, Yang J, Jin Q. 2016. VFDB 2016: hierarchical and refined dataset for big data analysis--10 years on. *Nucleic Acids Res* 44:D694–D697. <https://doi.org/10.1093/nar/gkv1239>