

Genome sequence of a human monkeypox virus isolate from Central Europe during the 2022 outbreak

Ármin Gergely Nagy,¹ Ágota Ábrahám,² István Prazsák,¹ Balázs Kakuk,¹ Brigitta Zana,² Ágnes Nagy,³ Dóra Tombác,¹ Gábor Kemenesi,^{2,4} Zsolt Boldogkői¹

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT We report the genome sequence of a human monkeypox virus isolated from an early Central European case. The 197,993 bp genome was assembled from amplicons sequenced using the Oxford Nanopore method. It has a GC content of 32.9% and belongs to subclade IIb B.1.3.

KEYWORDS monkeypox virus, genome sequencing, nanopore sequencing, long-read sequencing, amplicon sequencing

The human monkeypox virus (hMPXV) is a zoonotic virus in the *Orthopoxvirus* genus of the *Poxviridae* family (1, 2), closely related to the smallpox-causing variola virus (3). In August 2024, the World Health Organization declared hMPXV a public health emergency of international concern, and it is now listed among emerging pathogens with pandemic potential (4, 5).

To support a better understanding of hMPXV evolution, we sequenced the virus isolate MPXV_NRL_4279/2022, obtained in 2022, from a patient's skin lesion in Prague, Czechia.

The virus was propagated under BSL-4 conditions at the National Laboratory of Virology, University of Pécs, and passaged once in Vero cells (ATCC, CCL-81) to produce sufficient quantity of infectious material. DNA extraction was carried out using the Direct-zol RNA MiniPrep kit (Zymo Research, USA), following the manufacturer's instructions except for DNase I treatment.

A set of 22 multiplex PCR primer pairs was designed to generate ~10 kb overlapping amplicons covering the viral genome. Primer sequences along with the detailed protocol are available on protocols.io (6). Briefly, repliQa HiFi ToughMix (Quantabio) was used to amplify complex and repetitive genomic regions from fragments with 50–100 bp overlaps. Two distinct PCR rounds were performed, each targeting non-overlapping segments. PCR products were purified using SPRI beads (AmpureXP) following the manufacturer's instructions. Sequencing libraries were prepared with the SQK-LSK110 kit (Oxford Nanopore Technologies, ONT) and barcoded using the EXP-NBD196 kit. Sequencing was performed on a MinION Mk1B device with an R9.4.1 flow cell (FLO-MIN106), yielding 29,404 reads with an N50 of 9,556 (Table 1).

Basecalling using the super-accurate model and adapter trimming was performed with Guppy 6.5.7 (7), and low-quality reads were filtered out using NanoFilt2.8.0. *De novo* assembly using Raven 1.8.3 (8) produced two contigs, which were merged into a single sequence using RagTag 2.1.0 (9).

To determine the exact genomic termini, the results of an additional direct RNA-sequencing of our isolate (10) were utilized to generate a consensus sequence using samtools (11). The consensus matched the termini of the closely related, highly accurate OXO44336.2 genome (12).

Editor Jelle Matthijnssens, Katholieke Universiteit Leuven, Leuven, Belgium

Address correspondence to Zsolt Boldogkői, boldogkoi.zsolt@med.u-szeged.hu.

Ármin Gergely Nagy and Ágota Ábrahám contributed equally to this article. The author order was determined based on decreasing seniority.

Gábor Kemenesi and Zsolt Boldogkői are joint senior authors.

The authors declare no conflict of interest.

See the funding table on p. 3.

Received 23 April 2025

Accepted 5 September 2025

Published 21 October 2025

Copyright © 2025 Nagy et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

TABLE 1 Sequencing summary and structural variations of MPXV_NRL 4279/2022 isolate^{a,b}

Metrics type	Metrics value	Variant type	Position	Length (bp)	Sequence	Affected gene/Location
Number of Reads	29 404	Substitutions	36850–36851	2	AT → TA	OPG055
			64426	1	C → T	IGR
Mean Depth	1309,92		111074	1	C → T	OPG130
			190660	1	G → A	OPG016
Mean BaseQ	23,8	Deletions	607–609	3	TTT	IGR
			133177–133186	10	CAATCTTTCT	IGR
Mean MapQ	60		150586–150602	17	GATATGATGGATATGAT	IGR
			196615–196617	3	AAA	IGR
N50	9556	Insertions	0	118	# ^c	ITR
			133102	1	T	CGR
Min. Amplicon Size	5004 bp		173276	5	ATATA	CGR
			197209	113	#	ITR
Max. Amplicon Size	30 665 bp	VNTR Insertions	4806	128	[TAACCTAATTATGACT]*8	ITR
			179243	324	[CATTATATA]*36	CGR
			192530	128	[AAGTCATAAGTTAGTT]*8	ITR

^aSummary of amplicon sequencing metrics and substitutions, deletions as well as insertions in the consensus sequence compared to the [NC_063383NC_063383NC_063383](#), reference genome.

^bbp, base pair; BaseQ, basecalling quality score; CGR, core genomic region; IGR, intergenic region; ITR, inverted terminal repeat; MapQ, mapping quality; VNTR, variable number of tandem repeat.

^c# indicates TTTTCTAGACACTAAATAAATAGTAAATATAATATTAATGTACTAACTTATGTATTATTAATTTATCTAACTAAAGTTAGTAAATTATATACATAA.

The complete, assembled genome is 197,993 bp in length and has a 32.9% GC content. It shows no frameshift mutations or premature stop codons, indicating no major disruptions in protein-coding regions. Comparison with the reference genome ([NC_063383](#)) revealed four deletions totaling 33 bp, all located in intergenic regions. Five nucleotide substitutions were identified, three of which resulted in amino acid changes within the OPG055 (F11), OPG130 (A5L), and OPG016 (N3R) genes (Table 1). Additionally, seven insertions were detected: two at the terminal ends, three within variable number tandem repeat regions (VNTRs), and two small insertions (Table 1). Whole-genome phylogenetic analysis using Nextclade v3.10.2 (13) indicated that the MPXV_NRL_4279/2022 isolate belongs to Clade IIb B.1.3 of hMPXV. This genome, associated with the 2022 multi-country outbreak, provides valuable insight into the evolutionary dynamics of poxviruses.

ACKNOWLEDGMENTS

The authors thank Gábor E. Tóth for his assistance with the laboratory work. We would acknowledge the Doctoral School of Biology and Sport Biology, University of Pécs, Hungary, for providing research opportunities for Á.N. during their PhD studies.

Z.B. was supported by the National Research, Development and Innovation Office (NRDIO), grant number: K 142674. D.T. was supported by the NRDIO (FK 142676). I.P. was supported by the University Research Scholarship program (EKOP-24-4-SZTE-377) by the Ministry of Culture and Innovation, financed from the NRDIO fund. The development of the novel sequencing method has been implemented with the support provided by the Ministry of Culture and Innovation of Hungary from the National Research, Development and Innovation Fund, financed under the TKP2021 funding scheme with the project no. TKP-NVA-07.

AUTHOR AFFILIATIONS

¹Department of Medical Biology, University of Szeged, Albert Szent-Györgyi Medical School, Szeged, Hungary

²National Laboratory of Virology, Szentágothai Research Centre, University of Pécs, Pécs, Hungary

³Biological Laboratory of the Hungarian Defense Forces, Budapest, Hungary

⁴Faculty of Sciences, University of Pécs, Institute of Biology, Pécs, Baranya, Hungary

AUTHOR ORCID*s*

Dóra Tombácz <https://orcid.org/0000-0001-5520-2978>

Zsolt Boldogkői <http://orcid.org/0000-0003-1184-7293>

FUNDING

Funder	Grant(s)	Author(s)
Nemzeti Kutatási Fejlesztési és Innovációs Hivatal	K 142674	Zsolt Boldogkői
Nemzeti Kutatási Fejlesztési és Innovációs Hivatal	FK 142676	Dóra Tombácz
Ministry of Culture and Innovation	EKOP-24-4-SZTE-377	István Prazsák
Nemzeti Kutatási Fejlesztési és Innovációs Hivatal	TKP-NVA-07	Gábor Kemenesi

AUTHOR CONTRIBUTIONS

Ármin Gergely Nagy, Data curation, Formal analysis, Validation | Ágota Ábrahám, Investigation, Methodology | István Prazsák, Data curation, Formal analysis, Validation, Visualization, Writing – original draft | Balázs Kakuk, Data curation, Formal analysis, Methodology | Brigitta Zana, Investigation, project administration | Ágnes Nagy, Conceptualization, Investigation | Dóra Tombácz, Funding acquisition, Methodology, Writing – review and editing | Gábor Kemenesi, Conceptualization, Funding acquisition, Supervision, Writing – review and editing | Zsolt Boldogkői, Conceptualization, Formal analysis, Funding acquisition, Resources, Supervision, Writing – original draft, Writing – review and editing

DATA AVAILABILITY

The sequence has been deposited in the NCBI Viral Genomes database under the GenBank accession number [PV424067](#). Raw sequencing reads are available under BioProject ID [PRJNA1241293](#) and have been deposited in the Sequence Read Archive (SRA) under accession number [SRP572719](#).

ETHICS APPROVAL

The monkeypox virus strain (ID: MPXV_NRL 4279/2022) was obtained from the certified biobank of the National Institute of Public Health (Czech Republic), originally isolated for diagnostic purposes. No identifiable patient data were used, and ethical approval was not required for the use of anonymized biobank material.

REFERENCES

1. Lansiaux E, Jain N, Laivacuma S, Reinis A. 2022. The virology of human monkeypox virus (hMPXV): a brief overview. *Virus Res* 322:198932. <https://doi.org/10.1016/j.virusres.2022.198932>
2. Elsayed S, Bondy L, Hanage WP. 2022. Monkeypox virus infections in humans. *Clin Microbiol Rev* 35:e0009222. <https://doi.org/10.1128/cmr.00092-22>
3. Izadi M, Mirzaei F, Bagherzadeh MA, Ghiabi S, Khalifeh A. 2024. Discovering conserved epitopes of Monkeypox: novel immunoinformatic and machine learning approaches. *Heliyon* 10:e24972. <https://doi.org/10.1016/j.heliyon.2024.e24972>
4. WHO Media Team. 2024. WHO Director-General declares mpox outbreak a public health emergency of international concern. Available from: <http://www.who.int/news/item/14-08-2024-who-director-general-declares-mpox-outbreak-a-public-health-emergency-of-international-concern>. Retrieved 02 Jul 2025.
5. Ukoaka BM, Okesanya OJ, Daniel FM, Ahmed MM, Udam NG, Wagwula PM, Adigun OA, Udoh RA, Peter IG, Lawal H. 2024. Updated WHO list of emerging pathogens for a potential future pandemic: Implications for public health and global preparedness. *Infez Med* 32:463–477. <https://doi.org/10.53854/liim-3204-5>
6. Nagy GÁ, Ábrahám Á, Prazsák I, Kakuk B, Zana B, Nagy Á, Tombácz D, Kemenesi G, Boldogkői Z. 2025. Amplicon based sequencing of a human monkeypox virus isolate. Available from: <https://www.protocols.io/view/amplicon-based-sequencing-of-a-human-monkeypox-vir-5qpvo0589v4o/v2>
7. Wick RR, Judd LM, Holt KE. 2019. Performance of neural network basecalling tools for Oxford Nanopore sequencing. *Genome Biol* 20:129. <https://doi.org/10.1186/s13059-019-1727-y>

8. Vaser R, Šikić M. 2021. Time- and memory-efficient genome assembly with Raven. *Nat Comput Sci* 1:332–336. <https://doi.org/10.1038/s43588-021-00073-4>
9. Alonge M, Lebeigle L, Kirsche M, Jenike K, Ou S, Aganezov S, Wang X, Lippman ZB, Schatz MC, Soyk S. 2022. Automated assembly scaffolding using RagTag elevates a new tomato system for high-throughput genome editing. *Genome Biol* 23:258. <https://doi.org/10.1186/s13059-022-02823-7>
10. Kakuk B, Dörmő Á, Csabai Z, Kemenesi G, Holoubek J, Růžek D, Prazsák I, Dani VÉ, Dénes B, Torma G, Jakab F, Tóth GE, Földes FV, Zana B, Lanszki Z, Harangozó Á, Fülöp Á, Gulyás G, Mizik M, Kiss AA, Tombácz D, Boldogkői Z. 2023. In-depth temporal transcriptome profiling of monkeypox and host cells using nanopore sequencing. *Sci Data* 10:262. <https://doi.org/10.1038/s41597-023-02149-4>
11. Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, Whitwham A, Keane T, McCarthy SA, Davies RM, Li H. 2021. Twelve years of SAMtools and BCFtools. *Gigascience* 10:giab008. <https://doi.org/10.1093/gigascience/giab008>
12. Monzón S, Varona S, Negredo A, Vidal-Freire S, Patiño-Galindo JA, Ferressini-Gerpe N, Zaballos A, Orviz E, Ayerdi O, Muñoz-Gómez A, et al. 2024. Monkeypox virus genomic accordion strategies. *Nat Commun* 15:3059. <https://doi.org/10.1038/s41467-024-46949-7>
13. Aksamentov I, Roemer C, Hodcroft E, Neher R. 2021. Nextclade: clade assignment, mutation calling and quality control for viral genomes. *J Open Source Softw* 6:3773. <https://doi.org/10.21105/joss.03773>