

# Draft genome sequences of three *Pantoea* strains promoting the growth of wheat, isolated from desert plant rhizosphere soils

Yungang Liang,<sup>1,2</sup> Fangfang Xu,<sup>3</sup> Kai Tang,<sup>1,2</sup> Fuying Feng,<sup>1,2</sup> Huirong Liu<sup>1,2</sup>

**AUTHOR AFFILIATIONS** See affiliation list on p. 3.

**ABSTRACT** We report three draft genome sequences of wheat growth-promoting *Pantoea* spp. isolated from desert plant rhizosphere soils in western Inner Mongolia, China. Genomes were sequenced using Illumina NovaSeq 6000, yielding an average genome size of 4,285,493 bp and a mean GC content of 53.4%.

**KEYWORDS** *Pantoea*, plant growth-promoting rhizobacteria, genome sequencing

*Pantoea* spp. are recognized as potential plant growth-promoting rhizobacteria (1). To better understand their potential to enhance plant growth, we isolated and sequenced three *Pantoea* strains from desert plants. Rhizosphere soil samples, defined as soil adhering to the roots after shaking off bulk soil, were collected from desert plants at Bayannur, Inner Mongolia, China (40°14'28"N, 107°05'43"E) in October 2015. One gram of rhizosphere soil was serially diluted in 9 mL sterile saline, spread onto 1/2 Reasoner's 2A agar, and incubated at 28°C for 72 hours. Single colonies were picked and purified through successive streaking. Strains FN0302 and FN0307 were isolated from *Caragana intermedia* Kuang et H. C. Fu rhizosphere soils, and strain FN0305 was isolated from *Ammopiptanthus mongolicus* (Maxim. ex Kom.) Cheng f. rhizosphere soil. All three strains promoted wheat seedling growth under salt stress in pot experiments. Each strain was cultured in 1/2 Reasoner's 2A broth at 28°C for 72 h with shaking, centrifuged, and resuspended to an OD<sub>600</sub> of 1.0 in 0.1% (wt/vol) sterile CMC-Na solution for seed soaking (2). Treated seeds were sown in pots with soil salinized to 0.4% NaCl, with five seeds per pot and five replicates per treatment. Seedlings were sampled after 30 days for phenotypic measurements. Under these conditions, all three strains increased seedling emergence rates and shoot dry weights compared to uninoculated controls (Fig. 1).

Single colonies were inoculated into 1/2 Reasoner's 2A broth and incubated at 28°C with shaking for approximately 24 hours until stationary phase, then harvested by centrifugation at 5,000 × *g* for 10 min at 4°C. Genomic DNA was extracted using the SDS method (3) and processed into sequencing libraries using the NEBNext UltraDNA Library Prep Kit for Illumina (NEB, USA) according to the manufacturer's instructions. Whole genomes were sequenced using the Illumina NovaSeq 6000 platform at Novogene Bioinformatics Technology (Beijing, China), generating 150 bp paired-end (PE150) reads. Raw reads were processed using readfq software version 10 with parameters: "-rq1 input\_1.fq, -rq2 input\_2.fq, -oq1 out\_1.fq, -oq2 out\_2.fq, -adp1 adapter\_1.lst, -adp2 adapter\_2.lst, -Q QUAL,PERCENT, -C QUAL,PERCENT, -N PERCENT, -alen INT, -amis INT, -dup, -gz, -check1 read1.check, and -check2 read2.check." Clean data were assembled using SOAPdenovo (4, 5) version 2.04, ABySS (6) version 1.3.6, and SPAdes (7) version 3.10.0. Assembly results were integrated using CISA (8) version 1.3, selecting the assembly with the fewest scaffolds. GapCloser (9) version 1.12 was used to fill assembly gaps, removing fragments below 500 bp.

The genome assemblies were deposited in NCBI GenBank, where they were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (10) for public

**Editor** David A. Baltrus, The University of Arizona, Tucson, Arizona, USA

Address correspondence to Huirong Liu, lhz17@126.com.

The authors declare no conflict of interest.

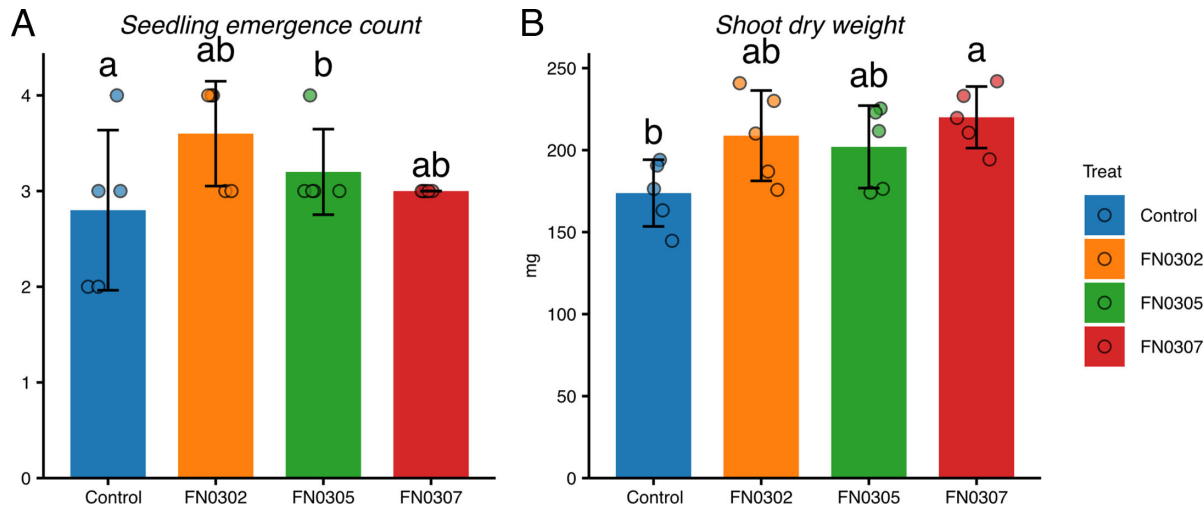
See the funding table on p. 3.

**Received** 6 May 2025

**Accepted** 19 June 2025

**Published** 11 February 2026

Copyright © 2026 Liang et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).



**FIG 1** Effects of three *Pantoea* strains on wheat seedling growth. (A) Seedling emergence count and (B) shoot dry weight of wheat plants inoculated with strains FN0302, FN0305, and FN0307 compared with the uninoculated control. Bars represent mean values, error bars indicate standard deviation, and points represent individual replicates. Different letters above bars indicate significant differences among treatments ( $P < 0.05$ ).

release. For downstream comparative and functional analyses described below, the genomes were re-annotated using Bakta (11) version 1.11.0. Genome sizes ranged from 4,204,445 to 4,345,745 bp with GC content from 53.34% to 53.48%. Contamination and completeness were assessed with CheckM (12) version 1.2.3. Average nucleotide identity (ANI) analysis using FastANI (13) version 1.32 indicated that all three strains were closely related to *Pantoea alhagi* LTYR-11Z (GenBank accession number [CP019706](#)). Sequencing and annotation details are provided in Table 1.

**TABLE 1** Isolation details and genome assembly statistics of three *Pantoea* strains

|                                                                                | FN0302                                          | FN0305                                                     | FN0307                                          |
|--------------------------------------------------------------------------------|-------------------------------------------------|------------------------------------------------------------|-------------------------------------------------|
| Host plant                                                                     | <i>Caragana intermedia</i><br>Kuang et H. C. Fu | <i>Ammopiptanthus mongolicus</i> (Maxim. ex Kom.) Cheng f. | <i>Caragana intermedia</i><br>Kuang et H. C. Fu |
| BioSample                                                                      | <a href="#">SAMN48141140</a>                    | <a href="#">SAMN48141141</a>                               | <a href="#">SAMN48141142</a>                    |
| Whole-genome sequencing (WGS) accession no.                                    | <a href="#">JBNJSM000000000</a>                 | <a href="#">JBNJSN000000000</a>                            | <a href="#">JBNJSO00000000</a>                  |
| SRA accession number                                                           | <a href="#">SRR33873240</a>                     | <a href="#">SRR33873239</a>                                | <a href="#">SRR33873238</a>                     |
| DNA genome size (bp)                                                           | 4,204,445                                       | 4,345,745                                                  | 4,306,289                                       |
| Illumina reads (read pairs)                                                    | 4,068,272                                       | 1,642,470                                                  | 2,000,000                                       |
| Coverage                                                                       | 290.28                                          | 113.38                                                     | 139.33                                          |
| Assembly $N_{50}$ value (bp)                                                   | 2,260,127                                       | 439,991                                                    | 625,342                                         |
| Assembly $N_{90}$ value (bp)                                                   | 128,349                                         | 131,528                                                    | 209,838                                         |
| GC content (%)                                                                 | 53.5                                            | 53.3                                                       | 53.4                                            |
| No. of contigs                                                                 | 14                                              | 26                                                         | 11                                              |
| rRNA gene count                                                                | 7                                               | 11                                                         | 6                                               |
| tmRNA gene count                                                               | 1                                               | 1                                                          | 1                                               |
| tRNA gene count                                                                | 75                                              | 78                                                         | 74                                              |
| Coding DNA sequence (CDS) count                                                | 3,840                                           | 3,947                                                      | 3,976                                           |
| Contamination (%)                                                              | 0.26                                            | 0.26                                                       | 0.46                                            |
| Completeness (%)                                                               | 99.79                                           | 99.79                                                      | 97.82                                           |
| Most closely related genome according to whole genome sequence in NCBI (% ANI) | <i>Pantoea alhagi</i><br>LTYR-11Z<br>(96.24)    | <i>Pantoea alhagi</i><br>LTYR-11Z<br>(96.34)               | <i>Pantoea alhagi</i><br>LTYR-11Z<br>(96.26)    |

## ACKNOWLEDGMENTS

This work was financially supported by the National Natural Science Foundation of China (31960021 and 67 U23A2054).

## AUTHOR AFFILIATIONS

<sup>1</sup>Laboratory for Environmental Microbiology and Biotechnology in Arid and Cold Regions, College of Life Sciences, Inner Mongolia Agricultural University, Hohhot, China

<sup>2</sup>Inner Mongolia Key Laboratory of Biomanufacturing Technology, Hohhot, China

<sup>3</sup>PLA Army Chemical Defense College, Beijing, China

## AUTHOR ORCID*s*

Yungang Liang  <http://orcid.org/0009-0001-5807-6564>

Huirong Liu  <http://orcid.org/0009-0002-5165-7136>

## FUNDING

| Funder                                       | Grant(s) | Author(s)   |
|----------------------------------------------|----------|-------------|
| National Natural Science Foundation of China | 31960021 | Fuying Feng |
| National Natural Science Foundation of China | U23A2054 | Fuying Feng |

## AUTHOR CONTRIBUTIONS

Yungang Liang, Writing – original draft | Fangfang Xu, Methodology | Kai Tang, Writing – review and editing | Fuying Feng, Funding acquisition, Writing – review and editing.

## DATA AVAILABILITY

This project has been deposited in DDBJ/ENA/GenBank under accession number [PRJNA1255394](https://www.ncbi.nlm.nih.gov/PRJNA1255394). BioSample numbers, SRA accession numbers, and the whole-genome shotgun (WGS) accession numbers are detailed in Table 1.

## REFERENCES

- Nascimento FX, Hernandez AG, Glick BR, Rossi MJ. 2020. The extreme plant-growth-promoting properties of *Pantoea phytobeneficialis* MSR2 revealed by functional and genomic analysis. *Environ Microbiol* 22:1341–1355. <https://doi.org/10.1111/1462-2920.14946>
- Xu F, Liang Y, Wang X, Guo Y, Tang K, Feng F. 2022. Synergic mitigation of saline-alkaline stress in wheat plant by silicon and *Enterobacter* sp. FN0603. *Front Microbiol* 13:1100232. <https://doi.org/10.3389/fmicb.2022.1100232>
- Lim HJ, Lee E-H, Yoon Y, Chua B, Son A. 2016. Portable lysis apparatus for rapid single-step DNA extraction of *Bacillus subtilis*. *J Appl Microbiol* 120:379–387. <https://doi.org/10.1111/jam.13011>
- Li R, Zhu H, Ruan J, Qian W, Fang X, Shi Z, Li Y, Li S, Shan G, Kristiansen K, Li S, Yang H, Wang J, Wang J. 2010. *De novo* assembly of human genomes with massively parallel short read sequencing. *Genome Res* 20:265–272. <https://doi.org/10.1101/gr.097261.109>
- Li R, Li Y, Kristiansen K, Wang J. 2008. SOAP: short oligonucleotide alignment program. *Bioinformatics* 24:713–714. <https://doi.org/10.1093/bioinformatics/btn025>
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJM, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. *Genome Res* 19:1117–1123. <https://doi.org/10.1101/gr.089532.108>
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>
- Lin S-H, Liao Y-C. 2013. CISA: contig integrator for sequence assembly of bacterial genomes. *PLoS One* 8:e60843. <https://doi.org/10.1371/journal.pone.0060843>
- Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read *de novo* assembler. *Gigascience* 1:18. <https://doi.org/10.1186/2047-217X-1-18>
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
- Schwengers O, Jelonek L, Dieckmann MA, Beyvers S, Blom J, Goesmann A. 2021. Bakta: rapid & standardized annotation of bacterial genomes via alignment-free sequence identification. *Bioinformatics*. <https://doi.org/10.1101/2021.09.02.458689>
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
- Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>