

Draft genome sequences of *Lactococcus petauri* isolated from Nile tilapia (*Oreochromis niloticus*) and rainbow trout (*Oncorhynchus mykiss*) in Mexican farms

Eduardo Matínez-Matus,^{1,2,3} César Ortega-Santana,⁴ Ruben Avendaño-Herrera^{1,2,5}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT *Lactococcus petauri* is an emerging fish pathogen linked to septicemia. We report draft genome sequences of two *L. petauri* isolates from independent lactococcosis outbreaks in Mexican fish farms. This study confirms *L. petauri* presence in Mexico and provides the first genomic data on this pathogen.

KEYWORDS *Lactococcus petauri*, lactococcosis, rainbow trout, Mexican farm

Lactococcus petauri is a gram-positive bacterium first identified in a sugar glider (*Petaurus breviceps*) in the United States (1). Recently, it has been linked to lactococcosis outbreaks in Nile tilapia (*Oreochromis niloticus*) (2) and rainbow trout (*Oncorhynchus mykiss*) across several countries (3). In Mexico, lactococcosis in rainbow trout was previously attributed to *Lactococcus garvieae* (4), with no reports of *L. petauri* until now. Here, we report the draft genome sequences of two *L. petauri* isolates from Mexican outbreaks: Que21-Lp08 from diseased tilapia spleen (Querétaro, 2021) and EP23-Lp16 from rainbow trout kidney (Estado de México, 2023), both confirmed to cause lactococcosis.

Samples for bacterial isolation were aseptically collected from the spleen and kidney of each fish. The samples were streaked onto tryptic soy agar (TSA) and incubated at 25°C for 24–72 h. Resulting pure isolates were stored at –80°C in CryoBank vials (Mast Diagnostica, Reinhold, Germany). Phenotypically, colonies were small, gram-positive cocci, catalase-negative, and γ-hemolytic.

For genome sequencing, isolates were grown on TSA at 25°C for 24 h. Genomic DNA was extracted using the NucleoSpin Soil kit (MACHEREY-NAGEL, Düren, Germany) and sequenced at SeqCoast Genomics (New Hampshire, USA). Libraries were prepared with the Illumina DNA Prep tagmentation kit and sequenced on an Illumina NextSeq 2000 platform using a 300-cycle flow cell to generate 2 × 150 bp paired-end reads. Raw read quality was assessed using FastQC v0.12.1 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>).

Sequencing generated 2,615,544 raw reads for Que21-Lp08 and 1,811,876 for EP23-Lp16. Adapters and read ends were trimmed with Trim Galore v0.6.10 (<https://github.com/FelixKrueger/TrimGalore>), and coverage was normalized by down-sampling reads with the BBnorm function implemented in BBTools v39.73 (5). Genome assemblies were generated using Unicycler v0.4.8 (6) and evaluated with QUAST v5.2.0 (7). Genome completeness and contamination were assessed using CheckM v1.2.4 (8), and functional annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.10 (20 January 2026) (9). Antimicrobial resistance genes were identified using the Resistance Gene Identifier in the Comprehensive Antibiotic Resistance Database (<https://card.mcmaster.ca/analyze/rgi>). All software was executed with default parameters.

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Address correspondence to Ruben Avendaño-Herrera, ravendano@unab.cl.

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TABLE 1 Assembly statistics and accession numbers of *Lactococcus petauri* strains isolated in Mexico and deposited in the NCBI database

Strain	SRA	Accession no.	No. of reads ^{a,b}	Total length ^c	Completeness ^d	Genome coverage	Contamination (%) ^d	Contigs ^c	%GC ^c	N50 (bp) ^c	Genes ^e	CDS ^e
Que21-Lp08	SRR3686552	JBTTGY000000000	2,615,544/2,564,030	2,028,205	98.6	187×	1.4	10	38.08	1,677,321	1,995	1,921
EP23-Lp16	SRR3686552	JBTTTHV000000000	1,811,876/1,782,122	2,028,373	98.6	130×	1.4	9	38.08	1,677,957	1,999	1,923

^aOriginal read number.^bNormalized read depth using BBNorm.^cDetermined using QUAST v5.2.0.^dDetermined by CheckM v1.2.4.^eDetermined using PGAP v6.10.

The draft genome assemblies (Table 1) comprised 10 (Que21-Lp08) and 9 (EP23-Lp16) contigs, with total lengths of 2,028,205 and 2,028,373 bp, respectively. Both genomes had a G + C content of 38.08%. The N50 values were 1,677,321 bp for Que21-Lp08 and 1,677,957 bp for EP23-Lp16, with average sequencing coverages of 187× and 130×, respectively. Genome annotation was performed using the automated NCBI PGAP with default parameters. A total of 1,995 genes, including 1,921 CDSs, 46 tRNAs, and 6 rRNAs, were predicted in Que21-Lp08, whereas 1,999 genes, 1,923 CDSs, 48 tRNAs, and 6 rRNAs were predicted in EP23-Lp16.

Species identification was confirmed using the Type (Strain) Genome Server (10) and JSpeciesWS—Taxonomic Thresholds (11). Both isolates clustered with the *L. petauri* 159469^T (accession no. [GCF_002154895.1](#)), with a digital DNA–DNA hybridization value of 86%. Average nucleotide identity (12) relative to *L. petauri* 159,469^T was 98.31% for Que21-Lp08 and 98.34% for EP23-Lp16, confirming their taxonomic assignment. The *IsaD* antimicrobial resistance gene was detected in both genomes.

These genomes represent the first *L. petauri* sequences from Mexican aquaculture and provide a resource for studies on genomic diversity and molecular diagnosis of lactococcosis in fish.

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AUTHOR AFFILIATIONS

¹Laboratorio de Patología de Organismos Acuáticos y Biotecnología Acuicola, Facultad de Ciencias de la Vida, Universidad Andrés Bello, Viña del Mar, Chile

²Interdisciplinary Center for Aquaculture Research—Applied Research (INCAR2), Universidad Andrés Bello, Viña del Mar, Chile

³Actino Lab USM, Centro de Biotecnología DAL, Universidad Técnica Federico Santa María, Valparaíso, Chile

⁴Centro de Investigación y Estudios Avanzados en Salud Animal, Facultad de Medicina Veterinaria y Zootecnia, Universidad Autónoma del Estado de México, Toluca, México

⁵Centro de Investigación Marina Quintay (CIMARQ), Universidad Andrés Bello, Quintay, Chile

AUTHOR ORCIDS

Ruben Avendaño-Herrera  <http://orcid.org/0000-0001-5368-4475>

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AUTHOR CONTRIBUTIONS

Eduardo Matínez-Matus, Formal analysis, Investigation, Methodology, Software, Writing – original draft | César Ortega-Santana, Conceptualization, Investigation, Supervision, Visualization, Writing – original draft | Ruben Avendaño-Herrera, Conceptualization, Funding acquisition, Investigation, Project administration, Resources, Supervision, Validation, Writing – original draft, Writing – review and editing

DATA AVAILABILITY

This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession numbers [JBTTGY0000000000](#) (Que21-Lp08) and [JBTHV0000000000](#) (EP23-Lp16). BioProject ID [PRJNA1403790](#) is publicly available and contains BioSample [SAMN54690575](#) and [SAMN54691117](#). The corresponding SRA records are [SRR36865524](#) (Que21-Lp08) and [SRR36865523](#) (EP23-Lp16).

ETHICS APPROVAL

All sampling and experimental procedures were conducted in accordance with institutional guidelines and were approved by the Ethics Committee of the Universidad Autónoma del Estado de México, Mexico (CICUAL-DISP/08-2025).

REFERENCES

- Goodman LB, Lawton MR, Franklin-Guild RJ, Anderson RR, Schaan L, Thachil AJ, Wiedmann M, Miller CB, Alcaine SD, Kovac J. 2017. *Lactococcus petauri* sp. nov., isolated from an abscess of a sugar glider. *Int J Syst Evol Microbiol* 67:4397–4404. <https://doi.org/10.1099/ijsem.0.02303>
- Egger RC, Rosa JCC, Resende LFL, de Pádua SB, de Oliveira Barbosa F, Zerbini MT, Tavares GC, Figueiredo HCP. 2023. Emerging fish pathogens *Lactococcus petauri* and *L. garvieae* in Nile tilapia (*Oreochromis niloticus*) farmed in Brazil. *Aquaculture* 565:739093. <https://doi.org/10.1016/j.aquaculture.2022.739093>
- Vela AI, del Mar Blanco M, Colussi S, Kotzamanidis C, Prearo M, Altinok I, Acutis PL, Volpatti D, Alba P, Feltrin F, Ianzano A, Domínguez L, Fernández-Garayzábal JF. 2024. The association of *Lactococcus petauri* with lactococcosis is older than expected. *Aquaculture* 578:740057. <https://doi.org/10.1016/j.aquaculture.2023.740057>
- Ortega C, Irgang R, Valladares-Carranza B, Collarte C, Avendaño-Herrera R. 2020. First identification and characterization of *Lactococcus garvieae* isolated from rainbow trout (*Oncorhynchus mykiss*) cultured in Mexico. *Animals (Basel)* 10:1609. <https://doi.org/10.3390/ani10091609>
- Bushnell B. 2014. BBMap: a fast, accurate, splice-aware aligner. <https://github.com/bbushnell/BBTools>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci USA* 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>