

Complete *de novo* assembly of *Wolbachia* endosymbiont of contemporary *Drosophila simulans* using long-read genome sequencing

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ABSTRACT We present a contemporary high-quality, complete *de novo* assembly of *Wolbachia pipientis* (wRi Merrill 23, [OZ411647](#)), an alphaproteobacterial endosymbiont of *Drosophila simulans* (*D. simulans*). This assembly was generated using long-read sequencing of wRi-infected *D. simulans* embryos collected from Merrill College at the University of California, Santa Cruz, in October 2023.

KEYWORDS genome, symbiosis, *Wolbachia*, *Drosophila*, evolution, evolutionary biology, biological control, ecology

Wolbachia pipientis infects diverse arthropods and nematodes, manipulating host phenotypes through cytoplasmic incompatibility (CI), male killing, and fertility rescue (1, 2). The Riverside strain (wRi) was first identified in California *Drosophila simulans* (*D. simulans*) in the 1980s (3) and rapidly spread statewide due to exceptionally strong CI (4). Despite its significance (5), the only reference genome reflects wRi present in 1984 (3, 6). Here, we present a complete *de novo* wRi genome assembly from contemporary *D. simulans* collected at the UC Santa Cruz Alan Chadwick Garden in Merrill College in October 2023.

To generate a contemporary wRi genome assembly, we collected wild *D. simulans* flies, established isofemale lines, and sequenced wRi-infected embryos. We established isofemale lines by deploying banana-baited bottles for ~5 days and collecting gravid females onto white food medium. After offspring eclosed, species identity was confirmed by phenotyping males and by PCR using silf-F/R primers to distinguish *D. simulans* from *D. melanogaster* (7) and wsp_1F/592R primers to confirm wRi identity (8). We extracted DNA from wRi-infected embryos using the Wizard HMW DNA Extraction Kit (Promega) without shearing and prepared libraries with the Native Barcoding Kit V14 (SQK-NBD114-24) without size selection. We sequenced these libraries on the Nanopore MinION Mk1B with an R10 flow cell (FLO-MIN-114) and MinKNOW v23.07.8 with adaptive sampling (fast model) to deplete *D. simulans* reads ([GCF_016746395.2](#)) yielding 4.26M reads, (N50 749 bp, 20 h) that were basecalled with Dorado (v0.7.3, hac model, --min-qscore 10). After filtering for host-free reads >3 kb (93K reads remaining, N50 10.5 kb), we assembled the wRi genome using Flye (9) following the methods described by Jacobs and Nakamoto *et al.* (10), yielding a 1.26 Mb circular assembly with 30× coverage. Flye determined circularization (4,000 bp minimum overlap), the genome was not rotated.

To polish the assembly, we generated Illumina short-read whole-genome sequencing data from whole wRi-infected *D. simulans* flies (Merrill 23 stocks). Illumina libraries were prepared using a Tn5-based tagmentation protocol (11). The Tn5 enzyme (Tn5R27S,

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TABLE 1 Annotation summary statistics

wRi Merrill 23 annotation summary	
Annotation pipeline	Prokka v1.1.1
Annotation method	kingdom:bacteria
Length (bp)	1,447,516
GC Content	35.21%
Genes (total)	1,392
CDSs (total)	1,392
Genes (RNA)	37
rRNAs	1, 1, 1 (5S, 16S, 23S)
tRNAs	34
ncRNAs	0
Pseudogenes (total)	0

E54K, and L372P) was obtained from QB3 MacroLab (University of California, Berkeley). Tagmentation simultaneously fragmented genomic DNA and ligated adapter sequences using custom oligos (Tn5-A: FC-121-1030, Tn5-B: FC-121-1031, and Tn5-rev; IDT). Libraries were indexed and amplified using the Nextera XT Index Kit v2 (i5/i7 adapters, Illumina) with KAPA HiFi (Roche) and sequenced on a NovaSeqX Plus (2 × 150 bp). Reads were trimmed to remove sequencing adapters and low-quality regions using fastp v1.0.1 (11) before polishing the assembly with Pilon v1.24 (12) (849K reads). We assessed the quality

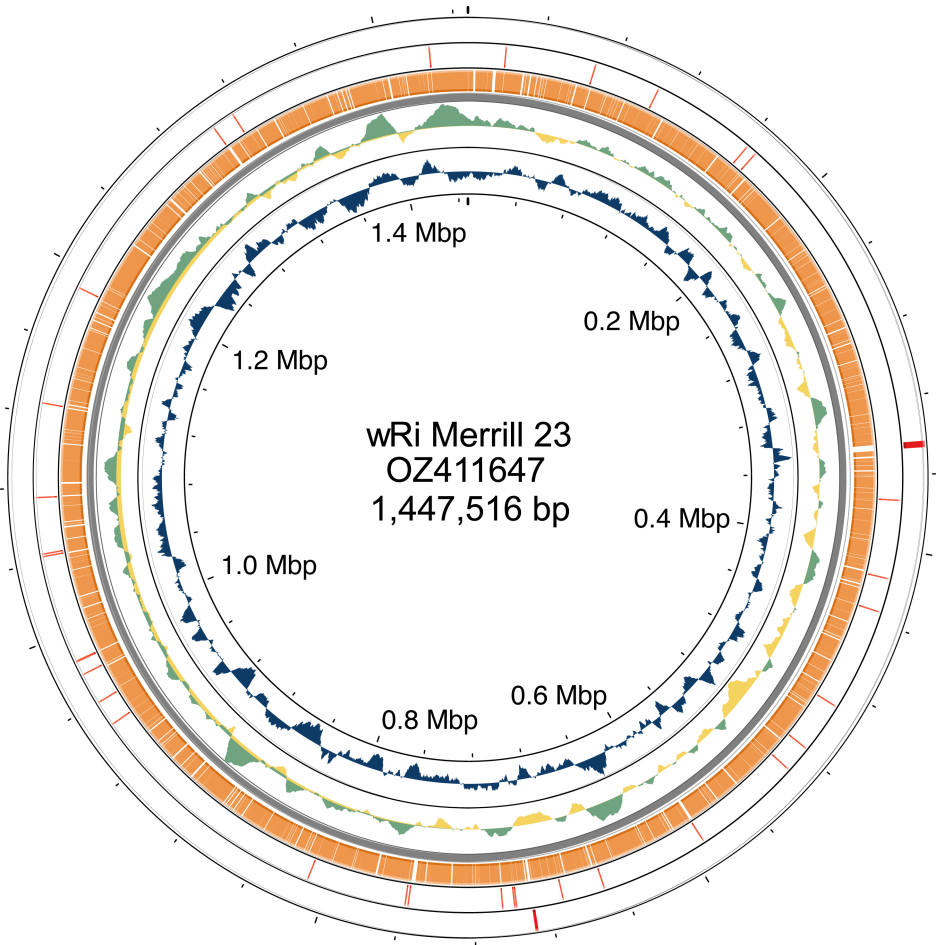


FIG 1 *Wolbachia* wRi genome map. Concentric circles show (outer to inner) rRNA genes (red), tRNA genes (red), coding sequences (orange), GC skew (green/yellow for high/low), and GC content (blue), with GC metrics plotted as deviations from the genome-wide average.

of the polished assembly with BUSCO v5.7.0 (13) (rickettsiales_odb10), achieving 99.2% completeness; annotated the assembly with Prokka v1.1.1 (14) (kingdom:bacteria) (Table 1); and calculated and visualized GC content and GC skew with Proksee v1.1.2 (15) (Fig. 1). Default parameters were used unless otherwise specified.

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AUTHOR CONTRIBUTIONS

Jodie Jacobs, Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Resources, Validation, Visualization, Writing – original draft, Writing – review and editing | Alexandra Lum, Data curation | Elyse Mina, Formal analysis | Camryn N. Morey, Investigation, Writing – review and editing | Darren D. Lee, Validation | Emry Gutierrez, Validation, Writing – review and editing | Jonah Dionisio, Validation | Cade Mirchandani, Data curation, Methodology, Resources | Luke Sylvester, Data curation | Anne Nakamoto, Data curation, Methodology | Hailey Loucks, Data curation | Ciara Wanket, Data curation | Ariana Cisneros, Writing – review and editing | Alessandro Calicchio, Writing – review and editing | Alexis N. Enstrom, Writing – review and editing | Camille Headrick, Writing – review and editing | Faith Okamoto, Writing – review and editing | Harrison D. Heath,

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DATA AVAILABILITY

Raw sequencing reads, [ERX15644803](https://doi.org/10.1038/s41587-019-0072-8) (Illumina) and [ERX15644805](https://doi.org/10.1038/s41587-019-0072-8) (ONT) and the assembled genome ([OZ411647](https://doi.org/10.1038/s41587-019-0072-8)) are available under BioProject accession number [PRJEB107302](https://doi.org/10.1038/s41587-019-0072-8). Assembly pipeline: https://github.com/jodieja-cobs/Jacobs_et_al_2026_de_novo_wRi_merrill_23_assembly.

REFERENCES

1. Russell S.L, Castillo J.R. 2020. Trends in symbiont-induced host cellular differentiation. *Results Probl Cell Differ* 69:137–176. https://doi.org/10.1007/978-3-030-51849-3_5
2. Russell S.L, Castillo J.R, Sullivan W.T. 2023. *Wolbachia* endosymbionts manipulate the self-renewal and differentiation of germline stem cells to reinforce fertility of their fruit fly host. *PLoS Biol* 21:e3002335. <https://doi.org/10.1371/journal.pbio.3002335>
3. Hoffmann A.A, Turelli M, Harshman L.G. 1990. Factors affecting the distribution of cytoplasmic incompatibility in *Drosophila simulans*. *Genetics* 126:933–948. <https://doi.org/10.1093/genetics/126.4.933>
4. Turelli M, Hoffmann A.A. 1995. Cytoplasmic incompatibility in *Drosophila simulans*: dynamics and parameter estimates from natural populations. *Genetics* 140:1319–1338. <https://doi.org/10.1093/genetics/140.4.1319>
5. Carrington L.B, Lipkowitz J.R, Hoffmann A.A, Turelli M. 2011. A Re-examination of *Wolbachia*-induced cytoplasmic incompatibility in California *Drosophila simulans*. *PLoS One* 6:e22565. <https://doi.org/10.1371/journal.pone.0022565>
6. Klasson L, Westberg J, Sapountzis P, Näslund K, Lutnaes Y, Darby A.C, Veneti Z, Chen L, Braig H.R, Garrett R, Bourtzis K, Andersson S.G.E. 2009. The mosaic genome structure of the *Wolbachia* wRi strain infecting *Drosophila simulans*. *Proc Natl Acad Sci USA* 106:5725–5730. <https://doi.org/10.1073/pnas.0810753106>
7. Faria V.G, Sucena É. 2017. From nature to the lab: establishing *Drosophila* resources for evolutionary genetics. *Front Ecol Evol* 5. <https://doi.org/10.3389/fevo.2017.00061>
8. Casper-Lindley C, Kimura S, Saxton D.S, Essaw Y, Simpson I, Tan V, Sullivan W. 2011. Rapid fluorescence-based screening for *Wolbachia* endosymbionts in *Drosophila* germ line and somatic tissues. *Appl Environ Microbiol* 77:4788–4794. <https://doi.org/10.1128/AEM.00215-11>
9. Kolmogorov M, Yuan J, Lin Y, Pevzner P.A. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>
10. Jacobs J, Nakamoto A, Mastoras M, Loucks H, Mirchandani C, Karim L, Penunuri G, Wanket C, Russell S.L. 2024. Complete *de novo* assembly of *Wolbachia* endosymbiont of *Drosophila willistoni* using long-read genome sequencing. *Sci Rep* 14:17770. <https://doi.org/10.1038/s41598-024-68716-w>
11. Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>
12. Walker B.J, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo C.A, Zeng Q, Wortman J, Young S.K, Earl A.M. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>
13. Manni M, Berkeley M.R, Seppely M, Simão F.A, Zdobnov E.M. 2021. BUSCO update: novel and streamlined workflows along with broader and deeper phylogenetic coverage for scoring of eukaryotic, prokaryotic, and viral genomes. *Mol Biol Evol* 38:4647–4654. <https://doi.org/10.1093/molbev/msab199>
14. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>
15. Grant J.R, Enns E, Marinier E, Mandal A, Herman E.K, Chen C, Graham M, Van Domselaar G, Stothard P. 2023. Proksee: in-depth characterization and visualization of bacterial genomes. *Nucleic Acids Res* 51:W484–W492. <https://doi.org/10.1093/nar/gkad326>