

Complete genome sequences of 12 *Azospirillum* strains isolated from diverse agroecosystems

Gabriele Bellotti,¹ Marta Bisaschi,¹ Edoardo Puglisi¹

AUTHOR AFFILIATION See affiliation list on p. 3.

ABSTRACT We report complete genome sequences of 12 *Azospirillum* strains generated using a hybrid Oxford Nanopore–Illumina sequencing approach. High-quality assemblies reveal the full multi-replicon structure of each genome, providing a valuable resource for comparative genomics, taxonomy, and studies on environmental adaptation in plant-associated bacteria.

KEYWORDS diazotrophs, biofertilizers, rhizobacteria, phytohormones, agroecosystems, genomics, plant-microbe interactions

Azospirillum are plant growth-promoting rhizobacteria known for their close associations with grasses and other crops. Several strains are commercially available due to their ability to fix atmospheric nitrogen and modulate phytohormones (1). In a previous work, some species were found to be misclassified, while many recognized species remain represented by a single genome (2). To address this gap, we acquired 10 *Azospirillum* strains from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ). In addition, two *A. argentinense* strains isolated in 2023 from tomato rhizospheres in Mali, cryopreserved in 20% glycerol at –20°C, were reactivated prior to sequencing to complete previous draft genomes (3).

DSMZ strains were obtained as freeze-dried cultures in sealed glass ampoules and revived following the supplier's instructions. The Mali strains were activated from cryotubes on agar plates incubated aerobically at 30°C for 48 h. Colonies were transferred to DSMZ-221 broth and incubated for 24 h at 180 rpm. Cells were harvested, and genomic DNA was extracted using the EZNA Bacterial DNA Kit (Omega Bio-tek, USA), following the manufacturer's protocol. DNA integrity number (DIN) was assessed using an Agilent 4500 TapeStation (High Sensitivity Kit; Agilent Technologies, USA), and purity was checked by NanoDrop (Thermo-Fisher Scientific, USA). Samples with DIN > 7, and A260/280 and A260/230 ratios >1.8 and 2.0, respectively, were sequenced.

ONT libraries were prepared using the Rapid Barcoding Kit 24 v14 (SQK-RBK114.24, ONT, UK) and loaded onto an R10.4.1 flow cell FLO-MIN114 on a MinION Mk1B device with MinKNOW v23.11.5. Raw signal data stored as POD5 were basecalled with Dorado v0.5.3 in super-accurate mode, generating FASTQ reads. Additional filtering was performed using NanoFilt v2.8.0, removing reads with Phred score <10 and reads <1,000 bp (4). From the same DNA extracts, Illumina short reads were generated by Novogene (Cambridge, UK) on a NovaSeq 6000 platform using PE250 technology. Library preparation followed the provider's workflow with a custom kit, including DNA fragmentation, end repair, A-tailing, adapter ligation, size selection, and PCR amplification. Libraries were purified using the AMPure XP system (Beverly). The resulting libraries were assessed on an Agilent Fragment Analyzer system and quantified to 1.5 nM using Qubit fluorometry (Thermo-Fisher Scientific) and qPCR. According to the Novogene QC report, raw reads had average Phred scores between 30 and 40.

Editor Leighton Pritchard, University of Strathclyde, Glasgow, United Kingdom

Address correspondence to Gabriele Bellotti, gabriele.bellotti@unicatt.it.

The authors declare no conflict of interest.

Received 2 December 2025

Accepted 30 March 2026

Published 20 April 2026

Copyright © 2026 Bellotti et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Read statistics, assembly quality metrics, and taxonomic assignment of the 12 *Azospirillum* genomes^a

Strain ID (DSMZ ID)	Ma4 (DSMZ 13400)	Sp 75 (DSMZ 2295)	Sp A2 (DSMZ 2297)	Sp A3a (DSMZ 1838)	Sp Br17 (DSMZ 1842)	Sp Col2b (DSMZ 2293)	Sp Col3 (DSMZ 2294)	Sp Col5 (DSMZ 1860)	Sp RG20a (DSMZ 1840)	Sp USA 5b (DSMZ 2292)	UC4320	UC4321
Isolate data	Origin	Germany, Freising	Brazil	Nigeria, Ibadan	Senegal, Dakar	Brazil	Colombia, Papayan	Colombia, Papayan	Colombia, Papayan	USA, Pullman	Mali, Bamako	Mali, Bamako
	Host/environment	<i>Miscanthus sinensis</i>	Maize roots	Maize roots	Grass roots	Maize roots	Maize roots	Bracharia roots	Wheat roots	Agricultural soil	Tomato rhizosphere	Tomato rhizosphere
Genome data	Sample acc. no.	SAMEA120325263	SAMEA120325264	SAMEA120325265	SAMEA120325266	SAMEA120325267	SAMEA120325268	SAMEA120325269	SAMEA120325270	SAMEA120325271	SAMEA120325272	SAMEA120325273
	Assembly acc. no.	GCA_977006905.2	GCA_977006955.2	GCA_977006985.2	GCA_977007015.2	GCA_977006935.2	GCA_977006915.2	GCA_977006952.2	GCA_977006975.2	GCA_977006945.2	GCA_977007025.2	GCA_977007005.2
	Genome coverage	76x	91x	113x	101x	55x	60x	50x	64x	57x	82x	116x
Raw reads	illumina no. reads (x250 bp)	2,197,436	2,286,004	2,116,656	5,418,784	5,199,096	2,080,502	3,369,880	2,682,922	2,354,872	17,292,764	18,423,740
data	illumina reads acc. no.	ERR15701807	ERR15701810	ERR15701811	ERR15701813	ERR15701814	ERR15701815	ERR15701816	ERR15701838	ERR15701839	ERR15701840	ERR15701844
	ONT tot reads length (bp)	27,313,058	544,419,509	475,034,816	216,270,867	183,833,901	307,558,085	368,846,639	34,798,674	57,623,640	113,354,585	39,818,961
	ONT no. reads	75,996.0	92,087.0	86,999.0	61,162.0	85,086.0	59,546.0	69,475.0	165,096.0	55,160.0	80,784.0	90,641.0
	ONT mean read length (bp)	3,962.4	4,010.5	4,141.9	5,223.0	4,474.6	2,825.5	3,306.1	1,635.4	4,755.1	4,690.4	4,311.8
	ONT reads acc. no.	ERR15700330	ERR15700317	ERR15700344	ERR15700345	ERR15700347	ERR15700386	ERR15700398	ERR15700400	ERR15700404	ERR15700432	ERR15700433
BUSCO	Completeness (%)	100	100	100	100	100	100	99.8	100	100	100	100
	Fragmented BUSCOs (%)	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
	Missing BUSCOs (%)	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
QUAST	No. of contigs	7	6	7	8	8	8	7	8	8	8	7
	Total genome size (bp)	7,501,717	6,863,639	7,591,753	7,106,214	7,379,528	7,719,855	6,977,525	8,855,533	6,770,114	7,470,993	7,172,418
	GC content (%)	68.8	69.1	68.4	68.3	67.8	67.7	67.5	67.8	67.8	68.6	69.0
N50 (bp)		1,357,979	1,822,607	2,064,099	1,107,683	1,154,421	1,085,136	1,348,664	1,192,050	976,033	1,960,025	1,788,884
Taxonomy	DSMZ reported as	<i>Azospirillum dobereineriae</i>	<i>Azospirillum brasilense</i>	<i>Azospirillum brasilense</i>	<i>Azospirillum</i>	<i>Azospirillum</i> sp.	<i>Azospirillum</i>	<i>Azospirillum</i>	<i>Azospirillum</i>	<i>Azospirillum</i>	<i>Azospirillum</i>	–
	Closest type strain	<i>Azospirillum dobereineriae</i>	<i>Azospirillum dobereineriae</i>	<i>Azospirillum dobereineriae</i>	<i>Azospirillum humicireducens</i>	<i>Azospirillum palustre</i>	<i>Azospirillum palustre</i>	<i>Azospirillum melinis</i>	<i>Azospirillum</i>	<i>Azospirillum palustre</i>	<i>Azospirillum</i>	<i>Azospirillum</i>
		DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000	DSMZ 11000
		GSE711	GCA_000632475.2	Az39 ⁺	humicireducens	B2 ⁺	palustre B2 ⁺	lipofurum Sp 59b ⁺	B2 ⁺	ramasanyii	argentinense Az39 ⁺	argentinense Az39 ⁺
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2	GCA_000632475.2
		1	2	1	2	1	1	1	1	1	1	1
		GCA_003989665.1	GCA_00063247									

Unless otherwise noted, default parameters were used for all software. Both long-read and short-read sequencing data were used to generate assemblies. Long-read assemblies were generated using Flye v2.9.6 (5), Raven v1.8.3 (6), and Miniasm v03r179 (7). Tricycler v0.5.5 was used to compare assemblies, resolve structural differences, remove terminal overlaps, close gaps, and circularize replicons (8). The presence of multiple replicons was inferred from consistent recovery of multiple clusters, each yielding a distinct circular contig. Consensus assemblies were polished with Illumina short reads using Polypolish v0.6.0 (9), enabling the recovery of complete multi-replicon genomes typical of *Azospirillum*.

Genome completeness was assessed using BUSCO v6.0.0 (10). Assembly statistics are reported in Table 1. Genomes were annotated via BV-BRC v3.56.67 (11) and assigned taxID 191. The hybrid approach produced high-quality genomes suitable for downstream phylogenomic and taxonomic analyses.

ACKNOWLEDGMENTS

Part of this research was supported by the project “APPRENDO – Agroecological, social, policy and financial approaches for a fair and equal access to technologies for efficient water use,” financed by Università Cattolica del Sacro Cuore within the D3.2 2023 funding scheme.

AUTHOR AFFILIATION

¹Department for Sustainable Food Process (DiSTAS), Università Cattolica del Sacro Cuore, Piacenza, Italy

AUTHOR ORCIDs

Gabriele Bellotti  <http://orcid.org/0000-0001-8521-3256>

Edoardo Puglisi  <https://orcid.org/0000-0001-5051-0971>

AUTHOR CONTRIBUTIONS

Gabriele Bellotti, Conceptualization, Data curation, Formal analysis, Software, Validation, Visualization, Writing – original draft | Marta Bisaschi, Formal analysis, Methodology, Software, Writing – original draft | Edoardo Puglisi, Conceptualization, Project administration, Resources, Supervision, Validation, Writing – review and editing

DATA AVAILABILITY

This Whole Genome Shotgun project, named “Complete genome sequences of *Azospirillum* strains,” has been deposited in the European Nucleotide Archive (ENA) under the project accession [PRJEB100579](https://www.ebi.ac.uk/ena/record/PRJEB100579). Raw reads and assemblies are available under the accession numbers reported in Table 1.

REFERENCES

1. Cassán F, Coniglio A, López G, Molina R, Nieves S, de Carlan CLN, Donadio F, Torres D, Rosas S, Pedrosa FO, de Souza E, Zorita MD, de-Bashan L, Mora V. 2020. Everything you must know about *Azospirillum* and its impact on agriculture and beyond. *Biol Fertil Soils* 56:461–479. <https://doi.org/10.1007/s00374-020-01463-y>
2. Bellotti G, Cortimiglia C, Antinori ME, Cocconcelli PS, Puglisi E. 2025. Comprehensive genome-wide analysis for the safety assessment of microbial biostimulants in agricultural applications. *Microb Genom* 11:001391. <https://doi.org/10.1099/mgen.0.001391>
3. Bellotti G, Sanou P, Sunzini P, Tabaglio V, Cocconcelli PS, Puglisi E. 2024. Draft genome sequences of four plant-associated free-living diazotrophs isolated from the rhizosphere of Malian tomato. *Microbiol Resour Announc* 13:e0056824. <https://doi.org/10.1128/mra.00568-24>
4. De Coster W, D’Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>
5. Lin Y, Yuan J, Kolmogorov M, Shen MW, Chaisson M, Pevzner PA. 2016. Assembly of long error-prone reads using de Bruijn graphs. *Proc Natl Acad Sci USA* 113:E8396–E8405. <https://doi.org/10.1073/pnas.1604560113>
6. Vaser R, Šikić M. 2021. Time- and memory-efficient genome assembly with Raven. *Nat Comput Sci* 1:332–336. <https://doi.org/10.1038/s43588-021-00073-4>
7. Li H. 2016. Minimap and miniasm: fast mapping and de novo assembly for noisy long sequences. *Bioinformatics* 32:2103–2110. <https://doi.org/10.1093/bioinformatics/btw152>
8. Wick RR, Judd LM, Cerdeira LT, Hawkey J, Méric G, Vezina B, Wyres KL, Holt KE. 2021. Tricycler: consensus long-read assemblies for bacterial

- genomes. *Genome Biol* 22:266. <https://doi.org/10.1186/s13059-021-02483-z>
9. Bouras G, Judd LM, Edwards RA, Vreugde S, Stinear TP, Wick RR. 2024. How low can you go? Short-read polishing of Oxford Nanopore bacterial genome assemblies. *Microb Genom* 10:001254. <https://doi.org/10.1099/mgen.0.001254>
 10. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212. <https://doi.org/10.1093/bioinformatics/btv351>
 11. Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL, Gillespie JJ, Gough R, Hix D, Kenyon R, et al. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. *Nucleic Acids Res* 42:D581–D691. <https://doi.org/10.1093/nar/gkt1099>
 12. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>
 13. Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>