

Complete genome sequence of pathogenic *Pseudomonas aeruginosa* strain P4 associated with persistent lung infections

Ravali Arugonda,¹ Namrata Bonde,¹ Nicholas Dillon¹

AUTHOR AFFILIATION See affiliation list on p. 2.

ABSTRACT Here we are reporting the circularized whole genome sequence of multi-drug resistant *Pseudomonas aeruginosa* strain P4, a clinical human lung isolate. This strain has previously been used to study antibiotic resistance in *P. aeruginosa*.

KEYWORDS *Pseudomonas aeruginosa*, pathogen, genomes

Multi-drug resistant (MDR) *Pseudomonas aeruginosa* is one of the ESKAPE pathogens and a leading cause of morbidity and mortality (1). In this announcement, we report the assembled complete genome sequence of *P. aeruginosa* strain P4, a clinical sputum sample originally obtained from a tertiary academic hospital in the New York metropolitan area (2). The strain was kindly provided to us by Dr. Victor Nizet (UCSD). Previous studies have been done on P4 in evaluating synergistic antibiotic combinations and antimicrobial peptides (2–4), understanding the effect of host environmental factors on immune responses (5), and in non-antibiotic disinfection strategies (6, 7). Here, we present a complete, circularized genome assembly of P4, comprising a chromosome and one plasmid. This published genome will be useful for studying the metabolism, mechanisms of antibiotic resistance, and virulence of the clinical *P. aeruginosa* strain P4. Genome assembly and annotation metrics are summarized in Table 1.

DNA from P4 was extracted following the Qiagen DNeasy Blood and Tissue kit (Cat#69504) instructions. Adapted from (8), P4 DNA was isolated via first resuspension of pellets from overnight cultures grown in cation-adjusted Mueller Hinton Broth (CA-MHB) at 37°C, 200 rpm in 1× TAE buffer (Tris, Acetic acid, EDTA) followed by centrifugation and resuspension in enzymatic lysis buffer (20 mM Tris-Cl, 2 mM EDTA, 1.2% Triton-X-100) and 50 mg/mL lysozyme. RNase A was then added, and the mixture was incubated at 37°C for 2 h. 25 µL of Proteinase K and 200 µL of AL buffer were added and incubated for 30 min at 56°C on a heat block (VWR). Qiagen kit instructions were followed exactly for the ethanol addition and DNA elution.

Hybrid sequencing was performed by SeqCenter (Pittsburgh, PA). Illumina sequencing libraries were prepared using the tagmentation-based and PCR-based Illumina DNA Prep kit and custom IDT 10 bp unique dual indices with a target insert size of 280 bp. Illumina sequencing was performed on an Illumina NovaSeq 6000 sequencer, producing 2×151 bp paired-end reads. Demultiplexing, quality control, and adapter trimming were performed with bcl-convert (v4.2.4).

Sample libraries were prepared using the PCR-free Oxford Nanopore Technologies (ONT) Ligation Sequencing Kit (SQK-NBD114.24) with the NEBNext Companion Module (E7180L), following the manufacturer's instructions. No additional DNA fragmentation or size selection was performed. Sequencing was carried out on an ONT GridION platform using R10.4.1 flow cells in multiplexed runs, with the 400 bp sequencing mode and a minimum read length threshold of 200 bp. Adaptive sampling was not enabled. Guppy (v6.5.7) was used for super-accurate basecalling, demultiplexing, and adapter trimming.

Editor Catherine Putonti, Loyola University Chicago, Chicago, Illinois, USA

Address correspondence to Nicholas Dillon, Nicholas.Dillon@UTDallas.edu.

The authors declare no conflict of interest.

Received 7 May 2025

Accepted 20 September 2025

Published 24 October 2025

Copyright © 2025 Arugonda et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

TABLE 1 Genomic characteristics of *P. aeruginosa* strain P4, including circularized chromosome and plasmid^a

Genomic features (P4)	Chromosome	Plasmid
Strain ID	P4	
SRA accession no.	SRR33560916 SRR33560917	
GenBank accession no.	CP187226	CP187227
Total no. of raw nanopore reads	239,640	
N ₅₀ of nanopore reads (bp)	11,394	
Total no. of Illumina reads (R1 +R2)	7,369,882	
No. of contigs	1	
Total length (bp)	6,670,990	47,865
Coverage of total genome (×)	319	
GC content (%)	66.25	59.12
No. of predicted genes	6,339	

^aGenomic statistics are based on the hybrid assembly of Nanopore and Illumina sequencing data.

Raw sequenced reads were quality controlled using FastQC (v0.12.1) (9). Trimmomatic (v0.39) (10) trimmed low-quality sequences from raw Illumina reads. Nanofilt (v2.8.0) (11) and porechop (v0.2.4) (12) were used to trim the residual adapter sequence from ONT reads. Illumina and ONT reads were assembled using Unicycler (v0.4.8) (13). Once the assembly was done, contigs were checked for circularization using circlator (v1.5.5) (14). Genome annotation was done using the NCBI Prokaryotic Genome Annotation Pipeline (v6.6) (15). The default parameters were used for all software.

AUTHOR AFFILIATION

¹Department of Biological Sciences, The University of Texas at Dallas, Richardson, Texas, USA

AUTHOR ORCIDs

Ravali Arugonda  <https://orcid.org/0009-0000-8061-9222>

Nicholas Dillon  <http://orcid.org/0000-0003-2351-0700>

AUTHOR CONTRIBUTIONS

Ravali Arugonda, Conceptualization, Data curation, Investigation, Methodology, Writing - original draft | Namrata Bonde, Conceptualization, Data curation, Methodology | Nicholas Dillon, Formal analysis, Funding acquisition, Investigation, Project administration, Validation, Visualization, Writing - review and editing

DATA AVAILABILITY

The complete circularized genome sequence of *P. aeruginosa* strain P4 has been deposited in GenBank under accession numbers [CP187226](#) (chromosome) and [CP187227](#) (plasmid). Raw sequencing reads are available in the Sequence Read Archive (SRA) under accession numbers [SRR33560916](#) (Illumina) and [SRR33560917](#) (Nanopore).

REFERENCES

1. Rice LB. 2008. Federal funding for the study of antimicrobial resistance in nosocomial pathogens: no ESCAPE. *J Infect Dis* 197:1079–1081. <https://doi.org/10.1086/533452>
2. Fair RJ, McCoy LS, Hensler ME, Aguilar B, Nizet V, Tor Y. 2014. Singly modified amikacin and tobramycin derivatives show increased rRNA A-site binding and higher potency against resistant bacteria. *ChemMedChem* 9:2164–2171. <https://doi.org/10.1002/cmdc.201402175>
3. Dillon N, Holland M, Tsunemoto H, Hancock B, Cornax I, Pogliano J, Sakoulas G, Nizet V. 2019. Surprising synergy of dual translation inhibition vs. *Acinetobacter baumannii* and other multidrug-resistant bacterial pathogens. *EBioMedicine* 46:193–201. <https://doi.org/10.1016/j.ebiom.2019.07.041>

4. Holland M, Bjanes E, Nizet V, Dillon N. 2022. Bicarbonate modulates delafloxacin activity against MDR *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *J Antimicrob Chemother* 77:433–442. <https://doi.org/10.1093/jac/dkab421>
5. Siew R, Ou T-L, Dahesh S, Akong K, Nizet V. 2022. Bicarbonate effects on antibacterial immunity and mucus glycobiology in the cystic fibrosis lung: a review with selected experimental observations. *Infect Microbes Dis* 4:103–110. <https://doi.org/10.1097/im9.000000000000101>
6. Banerjee B, Thompson C, Nizet V, Bjånes E. 2024. Bactericidal efficacy of low dose gaseous ozone against clinically relevant multidrug-resistant bacteria. *Front Microbiol* 15:1480433. <https://doi.org/10.3389/fmicb.2024.1480433>
7. Lin L, Kim J, Chen H, Kowalski R, Nizet V. 2016. Component analysis of multipurpose contact lens solutions to enhance activity against *Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Antimicrob Agents Chemother* 60:4259–4263. <https://doi.org/10.1128/AAC.00644-16>
8. Huo W, Adams HM, Zhang MQ, Palmer KL. 2015. Genome modification in *Enterococcus faecalis* OG1RF assessed by bisulfite sequencing and single-molecule real-time sequencing. *J Bacteriol* 197:1939–1951. <https://doi.org/10.1128/JB.00130-15>
9. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. Available from: <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>
10. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
11. De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>
12. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. *Microb Genom* 3:e000132. <https://doi.org/10.1099/mgen.0.000132>
13. Wick RR, Judd LM, Gorrie CL. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
14. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16. <https://doi.org/10.1186/s13059-015-0849-0>
15. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>