

Complete genome sequence of *Stutzerimonas balearica* JWOYS.421, a denitrifying strain isolated from the Hong Kong oyster (*Magallana hongkongensis*)

J. H. Y. Wong,¹ K. M. Leung,¹ G. K. K. Lai,¹ B. D. Russell,^{2,3} S. D. J. Griffin¹

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT *Stutzerimonas balearica* strain JWOYS.421 was recovered from the shell of a Hong Kong oyster (*Magallana hongkongensis*) collected from coastal waters in Hong Kong. Its complete genome, a single circular chromosome of 4,405,581 bp (64.80% G+C), was established through hybrid assembly.

KEYWORDS *Stutzerimonas balearica*, denitrification, Dpd operon, *Magallana hongkongensis*

Genus *Stutzerimonas* was proposed in 2022 for species formerly grouped within the *Pseudomonas stutzeri* complex, whose denitrifying activity had been observed as early as 1895 (1, 2). *Stutzerimonas balearica* is a Gram-negative, mesophilic, rod-shaped facultative anaerobe previously isolated from oil-polluted coastal sediment in the Balearic Islands (3). Exhibiting halotolerance up to 8.5% NaCl, its metabolic versatility has revealed it a promising candidate for marine bioremediation (3–5).

Here, *Stutzerimonas balearica* JWOYS.421 was isolated from the shell of a Hong Kong oyster (*Magallana hongkongensis*) from Lau Fau Shan (22.4682°N, 113.9845°E). One gram of crushed shell was vortexed with 19 mL saline (0.9% w/v) and allowed to settle for ~5 min. A 50 µL aliquot of the clearer supernatant was then added to 2 mL of A+ medium (6), and the mixture incubated overnight with shaking. After a 10,000-fold dilution with saline, 100 µL of the culture was spread onto denitrification medium (DM) agar containing 3.5% w/v NaCl (7) for overnight incubation. Resultant colonies were confirmed to reduce nitrate and nitrite (8) and selected isolates passaged to purity on DM agar. A single colony of JWOYS.421 was streaked onto DM agar before harvesting overnight growth for DNA extraction using Qiagen's DNeasy PowerSoil Pro kit. All agar/broth incubations were at 27°C.

Paired-end short-read libraries (NEBNext Ultra DNA Library Prep Kit, Illumina, Inc.) were sequenced via the NovaSeq 6000 platform using an SP PE250 flowcell and v1.5 Reagent Kit. A total of 9,169,642 raw single reads (mean length 250 bp) (SRX30846274) were quality-filtered and trimmed using TrimGalore! v0.6.7 (stringency:3; -e:0.2), producing 4,573,296 read pairs (mean length 247 bp, total ~1,129.6 Mbp). Long-read libraries, prepared from the same extracted DNA using the Rapid Barcoding Kit SQK-RBK004 without size selection, were sequenced via Oxford Nanopore's Spot-ON Flow Cell (vR9.4.1), MinION sequencer, and MinKNOW v20.06.2 software, with base-calling by Guppy v6.1.55 (high-accuracy mode). The final long-read data set of 288,403 raw reads (mean length 3,694 bp) (SRX30846275) was trimmed by Filtlong v0.2.1 (<https://github.com/rrwick/Filtlong>) (min_length: 2,000; target_bases: 500Mbp; length_weight: 10), selecting 71,777 reads (mean length 9,394 bp, N50 9,910; total ~674.3 Mbp) to scaffold the assembly. Default parameters were used for all software, unless otherwise specified.

Editor Frank J. Stewart, Montana State University, Bozeman, Montana, USA

Address correspondence to S. D. J. Griffin, sgriffin@isf.edu.hk.

The authors declare no conflict of interest.

Received 1 April 2026

Accepted 24 April 2026

Published 18 May 2026

Copyright © 2026 Wong et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Predicted genes^a in *Stutzerimonas balearica* JWOYS.421 linked to denitrification

Gene(s)	Locus (NCBI)	Activity
<i>nasD nasE</i>	ACRYHF_00990, ACRYHF_00900	Soluble NADH-dependent nitrite reductase
<i>ntnC ntrB</i>	ACRYHF_01850, ACRYHF_01855	Nitrogen regulation protein NR(I), NR(II)
<i>narKMXL narGHJI</i>	ACRYHF_03580 to ACRYHF_03565 ACRYHF_03585 to ACRYHF_03600	Nitrate transport and regulation, dissimilatory nitrate reductase
<i>putative dnrE</i>	ACRYHF_04205	Transcription factor regulating respiratory nitrate reduction
<i>norR norCB norD</i>	ACRYHF_04175 ACRYHF_04265 to ACRYHF_04275	Nitric oxide reductase
<i>napEDABC</i>	ACRYHF_06300 to ACRYHF_06280	Periplasmic nitrate reductase
<i>norCBQD</i>	ACRYHF_08815 to ACRYHF_08830	bc-type nitric oxide reductase
<i>nirBD</i>	ACRYHF_09940, ACRYHF_09945	Siroheme-containing nitrite reductase
<i>nasTS</i>	ACRYHF_12175, ACRYHF_12170	Control of assimilatory nitrate/nitrite reduction
<i>nasAB</i>	ACRYHF_12180, ACRYHF_12195	Assimilatory nitrate/nitrite reductase
<i>nirMCFDLGH</i>	ACRYHF_17190 to ACRYHF_17160	Biosynthesis and maturation of the cytochrome cd1 nitrite reductase NirS
<i>nirS</i>	ACRYHF_17195	Nitrite reductase NirS
<i>nirQOP</i>	ACRYHF_17200 to ACRYHF_17210	Reductase regulation
<i>nirJENY</i>	ACRYHF_17215 to ACRYHF_17230	Biosynthesis of the heme d1 cofactor
<i>nosRZDFYL-tata</i>	ACRYHF_17290 to ACRYHF_17260	Nitrous oxide reductase

^aBy NCBI PGAP v6.10 (11) and by BLASTp with *Stutzerimonas stutzeri* A1501 (CP000304.1) (13).

Illumina and MinION data sets were combined using Tricycler v0.5.5 (9), with long-read polishing by Medaka v2.1.0 and short-read polishing by Polypolish v0.6.0 (short-read coverage 256×) (10, 11), yielding a single circular chromosome (4,405,581 bp, 64.80% G+C) rotated to begin *dnaA* (A0A8D3XX96) with mean long-read coverage 153× (judged 99.34% complete with 0.96% contamination by CheckM v1.2.4 [12]), which was submitted to NCBI PGAP v6.10 (13) for annotation and to JSpecies v5.0.2 (14) for taxonomic assignment. *Stutzerimonas balearica* JWOYS.421 shares an average nucleotide identity (ANInu by EZBioCloud [15]) of 98.30% with *Stutzerimonas balearica* DSM 6083 (CP007511.1). Genes linked to denitrification, predicted via PGAP v6.10 and by BLASTp with *Stutzerimonas stutzeri* A1501 (CP000304.1) (16), are shown in Table 1. Nitrite reductase NirS, encoded at ACRYHF_17195, belongs to clade 1a (17, 18), while NirK is not present. A Dpd operon enabling DNA modification with 7-deazaguanine derivatives (19), which is absent in DSM 6083 and A1501, is found at loci ACRYHF_11555 to ACRYHF_11630.

AUTHOR AFFILIATIONS

¹Shuyuan Molecular Biology Laboratory, The Independent Schools Foundation Academy, Hong Kong SAR, China

²The Swire Institute of Marine Science and Area of Ecology and Biodiversity, School of Biological Sciences, The University of Hong Kong, Hong Kong SAR, China

³Institute for Climate and Carbon Neutrality, The University of Hong Kong, Hong Kong SAR, China

AUTHOR ORCID*s*

J. H. Y. Wong  <http://orcid.org/0009-0004-4481-2550>

K. M. Leung  <http://orcid.org/0000-0001-9939-8300>

G. K. K. Lai  <http://orcid.org/0000-0003-4003-4478>

B. D. Russell  <http://orcid.org/0000-0003-1282-9978>

S. D. J. Griffin  <http://orcid.org/0000-0001-9995-926X>

AUTHOR CONTRIBUTIONS

J. H. Y. Wong, Data curation, Formal analysis, Investigation, Methodology, Writing – original draft | K. M. Leung, Investigation, Methodology, Resources | G. K. K. Lai, Data curation, Formal analysis, Methodology, Project administration, Resources, Software, Supervision | B. D. Russell, Conceptualization, Resources | S. D. J. Griffin, Conceptualization, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Visualization, Writing – review and editing

DATA AVAILABILITY

Sequencing data for *Stutzerimonas balearica* JWOYS.421 is available through NCBI under GenBank assembly [GCA_051468625.1](https://www.ncbi.nlm.nih.gov/assembly/GCA_051468625.1) and GenBank accession [CP195794](https://www.ncbi.nlm.nih.gov/assembly/CP195794). Raw reads may be found under [SRX30846274](https://www.ncbi.nlm.nih.gov/sra/SRX30846274) (NovaSeq) and [SRX30846275](https://www.ncbi.nlm.nih.gov/sra/SRX30846275) (MinION). BLASTp comparison data between *Stutzerimonas stutzeri* A1501 and *Stutzerimonas balearica* JWOYS.421 can be found at <https://doi.org/10.6084/m9.figshare.31901167>.

REFERENCES

- Gomila M, Mulet M, García-Valdés E, Lalucat J. 2022. Genome-based taxonomy of the genus *Stutzerimonas* and proposal of *S. frequens* sp. nov. and *S. degradans* sp. nov. and emended descriptions of *S. perfectomarina* and *S. chloritidismutans*. *Microorganisms* 10:1363. <https://doi.org/10.3390/microorganisms10071363>
- Lehmann KB, Neumann R. 1896. Atlas und Grundriss der Bakteriologie und Lehrbuch der speciellen bakteriologischen Diagnostik, p 237–238. In Lehmann JF (ed), *Lehmann's medicinisches Handatlas* Band X, 1st ed. Vol. II.
- Bennasar A, Rosselló-Mora R, Lalucat J, Moore ER. 1996. 16S rRNA gene sequence analysis relative to genomovars of *Pseudomonas stutzeri* and proposal of *Pseudomonas balearica* sp. nov. *Int J Syst Bacteriol* 46:200–205. <https://doi.org/10.1099/00207713-46-1-200>
- Salvà-Serra F, Pérez-Pantoja D, Donoso RA, Jaén-Luchoro D, Fernández-Juárez V, Engström-Jakobsson H, Moore ERB, Lalucat J, Bennasar-Figueras A. 2023. Comparative genomics of *Stutzerimonas balearica* (*Pseudomonas balearica*): diversity, habitats, and biodegradation of aromatic compounds. *Front Microbiol* 14:1159176. <https://doi.org/10.3389/fmicb.2023.1159176>
- Ahmadi M, Jorfi S, Kujlu R, Ghafari S, Darvishi Cheshmeh Soltani R, Jaafarzadeh Haghighifard N. 2017. A novel salt-tolerant bacterial consortium for biodegradation of saline and recalcitrant petrochemical wastewater. *J Environ Manage* 191:198–208. <https://doi.org/10.1016/j.jenvman.2017.01.010>
- Wilhelm SW. 2017. A+ media for marine phytoplankton. Available from: <https://dx.doi.org/10.17504/protocols.io.ibncame>
- Zhang Q-L, Liu Y, Ai G-M, Miao L-L, Zheng H-Y, Liu Z-P. 2012. The characteristics of a novel heterotrophic nitrification-aerobic denitrification bacterium, *Bacillus methylotrophicus* strain L7. *Bioresour Technol* 108:35–44. <https://doi.org/10.1016/j.biortech.2011.12.139>
- Buxton R. 2011. Protocols: nitrate and nitrite reduction test. American Society for Microbiology. Available from: <https://asm.org/protocols/nitrate-and-nitrite-reduction-test-protocols>
- Wick Ryan R, Judd LM, Cerdeira LT, Hawkey J, Méric G, Vezina B, Wyres KL, Holt KE. 2021. Tricycler: consensus long-read assemblies for bacterial genomes. *Genome Biol* 22:266. <https://doi.org/10.1186/s13059-021-02483-z>
- Wick R. R., Judd LM, Holt KE. 2023. Assembling the perfect bacterial genome using Oxford Nanopore and Illumina sequencing. *PLoS Comput Biol* 19:e1010905. <https://doi.org/10.1371/journal.pcbi.1010905>
- Bouras G, Judd LM, Edwards RA, Vreugde S, Stinear TP, Wick RR. 2024. How low can you go? Short-read polishing of Oxford Nanopore bacterial genome assemblies. *Microb Genom* 10:001254. <https://doi.org/10.1099/mgen.0.001254>
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
- Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. *Nucleic Acids Res* 46:D851–D860. <https://doi.org/10.1093/nar/gkx1068>
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>
- Chalita M, Kim YO, Park S, Oh HS, Cho JH, Moon J, Baek N, Moon C, Lee K, Yang J, Nam GG, Jung Y, Na SI, Bailey MJ, Chun J. 2024. EzBioCloud: a genome-driven database and platform for microbiome identification and discovery. *Int J Syst Evol Microbiol* 74:006421. <https://doi.org/10.1099/ijsem.0.006421>
- Yan Y, Yang J, Chen L, Yang F, Dong J, Xue Y, Xu X, Zhu Y, Yao Z, Lin M, Wang Y, Jin Q. 2005. Structural and functional analysis of denitrification genes in *Pseudomonas stutzeri* A1501. *Sci China Ser C* 48:585. <https://doi.org/10.1360/062005-45>
- Jones CM, Stres B, Rosenquist M, Hallin S. 2008. Phylogenetic analysis of nitrite, nitric oxide, and nitrous oxide respiratory enzymes reveal a complex evolutionary history for denitrification. *Mol Biol Evol* 25:1955–1966. <https://doi.org/10.1093/molbev/msn146>
- Menestreau M, Milligan DA, Sennett LB, Bergaust L, Bakken LR, Rowley G, Kjos M, Shapleigh JP, Frostegård Å. 2025. Distinct denitrification phenotypes in closely related bacteria: clues to understanding variations in nitrite accumulation among *Stutzerimonas* strains. *bioRxiv* (Cold Spring Harbor Laboratory). <https://doi.org/10.64898/2025.12.02.691782>
- Thiaville JJ, Kellner SM, Yuan Y, Hutinet G, Thiaville PC, Jumpathong W, Mohapatra S, Brochier-Armanet C, Letarov AV, Hillebrand R, Malik CK, Rizzo CJ, Dedon PC, de Crécy-Lagard V. 2016. Novel genomic island modifies DNA with 7-deazaguanine derivatives. *Proc Natl Acad Sci USA* 113:E1452–E1459. <https://doi.org/10.1073/pnas.1518570113>