

Complete genome sequence of *Stenotrophomonas* sp. strain C-A, isolated from contaminated *Pseudomonas putida* cultures

Marissa N. Roghair Stroud,¹ Larry J. Halverson^{1,2}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT We report the complete *de novo* genome assembly of *Stenotrophomonas* sp. strain C-A, isolated from a contaminated culture of *Pseudomonas putida* KT2440. The genome's annotation may inform how the two organisms interact with one another in mixed culture, and how growth state influences those interactions.

KEYWORDS genome, cell-cell interaction, *Stenotrophomonas*, *Pseudomonas putida*, biofilms

We isolated *Stenotrophomonas* sp. strain C-A from a contaminated culture of *Pseudomonas putida* KT2440 (proposed to be named *P. alloputida* [1, 2]) used for planktonic and biofilm experiments in Reasoner's 2A (R2A) broth. In these studies, we noted the appearance of a second colony morphology when dilution plating and counting colony-forming units. This second colony type was slower-growing, yellowish, and sticky, becoming detectable 24 h after KT2440 colonies were visible. To isolate *Stenotrophomonas* sp. C-A, the contaminated KT2440 culture was grown in R2A broth overnight, then diluted and plated onto Luria agar (LA), which showed two different colony morphologies. Multiple colonies of C-A were isolated and streaked for purity, all exhibiting identical growth properties. C-A grows well on LA, R2A, or 1/2-strength tryptic soy agar mediums, at both room temperature and 30°C.

Stenotrophomonas sp. C-A was grown overnight in LB, washed twice with 1 mM phosphate buffer, and resuspended in Zymo 1× DNA/RNA Shield. This was sent to Plasmidsaurus, who performed DNA isolation, library preparation, and sequencing. DNA was isolated using the Zymo Quick-DNA Miniprep Plus Kit. The library was prepared without amplification using the Nanopore Rapid Barcoding Kit 96 V14. This was sequenced with a primer-free protocol, using R10.4.1 flow cells on a PromethION P24 device, and Dorado v4.3 Super Accurate base-calling model. Read quality control was assessed with a minimum Q-score of 10, and adapter sequences were trimmed using MinKNOW.

Raw fastq files were used for *de novo* genome assembly using a pipeline developed in our lab (3), using default parameters except where otherwise noted. SeqKit v2.10.1 (4) was used to determine the number of raw reads (155,471) and N_{50} (11,539 bp). Reads were filtered using Chopper 0.11.0 (--quality 9 --minlength 1000) (5). Four different assembly programs were used to generate genome assemblies: Flye 2.9.6-b1802 (6), Raven 1.8.3 (7), Miniasm 0.3-r179 (8) with Minipolish v0.2.0 (9), and NextDenovo v2.5.2 (10). A consensus assembly was generated using Tricycler v0.5.6 (11), polished with Medaka v2.0.1, and rotated to start at *dnaA* using Circlator v1.5.5 (12). The final genome was submitted to NCBI for annotation with the Prokaryotic Genome Annotation Pipeline (PGAP v6.10) (13). Completeness and contamination were assessed with EvalG, CheckM, and EvalCon (14, 15), accessed through the Bacterial and Viral Bioinformatics Resource Center Genome Annotation Service (16). The organism was identified as the genus *Stenotrophomonas* but did not match any known species based on a Type Strain Genome

Editor Elinne Becket, California State University San Marcos, San Marcos, California, USA

Address correspondence to Larry J. Halverson, larryh@iastate.edu.

The authors declare no conflict of interest.

See the funding table on p. 2.

Received 16 January 2026

Accepted 11 April 2026

Published 18 May 2026

Copyright © 2026 Roghair Stroud and Halverson. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Server (17) analysis using default parameters; the closest related organism by genome phylogeny was *Stenotrophomonas geniculata* ATCC 19374 (BioSample: [SAMN04207799](#)).

The final assembled genome is 4,770,670 bp with 203.55× coverage and a GC content of 66.24%. There is only one contig ($N_{50} = 4,770,670$). There are 4,411 predicted genes and 4,318 predicted CDS in the PGAP annotation. The genome is estimated to have 100% completeness and 0% contamination.

ACKNOWLEDGMENTS

Bacterial genome sequencing was performed by Plasmidsaurus using Oxford Nanopore Technology with custom analysis and annotation. Ashley Paulsen provided advice and guidance on the assembly and annotation of the genome.

This research was supported by the USDA National Institute of Food and Agriculture AFRI Education and Workforce Development Predoctoral Fellowship (2022-11424) and an Agricultural Microbiomes in Plant Systems and Natural Resources award (2021-67019-34833), as well as the Iowa Agriculture and Home Economics Experiment Station.

AUTHOR AFFILIATIONS

¹Department of Plant Pathology, Entomology, and Microbiology, Iowa State University, Ames, Iowa, USA

²Ames National Laboratory, U.S. Department of Energy, Iowa State University, Ames, Iowa, USA

AUTHOR ORCIDs

Marissa N. Roghair Stroud  <http://orcid.org/0000-0002-2011-3526>

Larry J. Halverson  <http://orcid.org/0000-0002-9164-8691>

FUNDING

Funder	Grant(s)	Author(s)
U.S. Department of Agriculture	2021-67019-34833, 2022-11424	Marissa N. Roghair Stroud Larry J. Halverson

AUTHOR CONTRIBUTIONS

Marissa N. Roghair Stroud, Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Writing – original draft, Writing – review and editing | Larry J. Halverson, Conceptualization, Data curation, Formal analysis, Funding acquisition, Investigation, Methodology, Project administration, Supervision, Writing – review and editing

DATA AVAILABILITY

This whole-genome sequencing project has been deposited at NCBI under BioProject number [PRJNA1373328](#) and BioSample number [SAMN53651247](#). The assembled genome can be found with accession number [CM132916.1](#). The raw reads have been deposited to NCBI's SRA under accession number [SRS27304381](#).

REFERENCES

1. Keshavarz-Tohid V, Vacheron J, Dubost A, Prigent-Combaret C, Taheri P, Tarighi S, Taghavi SM, Moënne-Loccoz Y, Muller D. 2019. Genomic, phylogenetic and catabolic re-assessment of the *Pseudomonas putida* clade supports the delineation of *Pseudomonas alloputida* sp. nov., *Pseudomonas inefficax* sp. nov., *Pseudomonas persica* sp. nov., and *Pseudomonas shirazica* sp. nov. Syst Appl Microbiol 42:468–480. <https://doi.org/10.1016/j.syapm.2019.04.004>
2. Oren A, Garrity G. 2020. List of new names and new combinations previously effectively, but not validly, published. Int J Syst Evol Microbiol 70:4043–4049. <https://doi.org/10.1099/ijsem.0.004244>
3. Paulsen AA, Vang DX, Halverson LJ. 2025. Complete genome sequences of 37 bacteria in a maize rhizosphere synthetic community. Microbiol Resour Announc 14:e0049625. <https://doi.org/10.1128/mra.00496-25>

4. Shen W, Sipos B, Zhao L. 2024. SeqKit2: a Swiss army knife for sequence and alignment processing. *iMeta* 3:e191. <https://doi.org/10.1002/imt2.191>
5. De Coster W, Rademakers R. 2023. NanoPack2: population-scale evaluation of long-read sequencing data. *Bioinformatics* 39:btad311. <https://doi.org/10.1093/bioinformatics/btad311>
6. Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>
7. Vaser R, Šikić M. 2021. Time- and memory-efficient genome assembly with Raven. *Nat Comput Sci* 1:332–336. <https://doi.org/10.1038/s43588-021-00073-4>
8. Li H. 2016. Minimap and minimap: fast mapping and *de novo* assembly for noisy long sequences. *Bioinformatics* 32:2103–2110. <https://doi.org/10.1093/bioinformatics/btw152>
9. Wick RR, Holt KE. 2024. Benchmarking of long-read assemblers for prokaryote whole genome sequencing. *F1000Res* 8:2138. <https://doi.org/10.12688/f1000research.21782.4>
10. Hu J, Wang Z, Sun Z, Hu B, Ayoola AO, Liang F, Li J, Sandoval JR, Cooper DN, Ye K, Ruan J, Xiao C-L, Wang D, Wu D-D, Wang S. 2024. NextDenovo: an efficient error correction and accurate assembly tool for noisy long reads. *Genome Biol* 25:107. <https://doi.org/10.1186/s13059-024-03252-4>
11. Wick RR, Judd LM, Cerdeira LT, Hawkey J, Méric G, Vezina B, Wyres KL, Holt KE. 2021. Trycycler: consensus long-read assemblies for bacterial genomes. *Genome Biol* 22:266. <https://doi.org/10.1186/s13059-021-02483-z>
12. Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294. <https://doi.org/10.1186/s13059-015-0849-0>
13. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
14. Parrello B, Butler R, Chlenski P, Olson R, Overbeek J, Pusch GD, Vonstein V, Overbeek R. 2019. A machine learning-based service for estimating quality of genomes using PATRIC. *BMC Bioinformatics* 20:486. <https://doi.org/10.1186/s12859-019-3068-y>
15. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>
16. Shukla M, Wattam AR, Aleman A, Bhattacharya R, Bowers N, Brettin T, Capria A, Chia N, Cucinell C, Davis JJ, et al. 2025. BV-BRC: a unified bacterial and viral bioinformatics resource with expanded functionality and AI integration. *Nucleic Acids Res* gkaf1254. <https://doi.org/10.1093/nar/gkaf1254>
17. Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>