

Complete genome sequence of *Campylobacter* sp. GTC17093 isolated from human skin infection

Jun Yonetamari,^{1,2} Masahiro Hayashi,^{3,4,5} Yoshinori Muto,³ Kaori Tanaka^{1,3,4,5}

AUTHOR AFFILIATIONS See affiliation list on p. 3.

ABSTRACT *Campylobacter*, a microaerophilic, Gram-negative spirillum, has been isolated from both animal and human specimens; however, most identified species are of animal origin. Here, we report the complete genome sequence of a new *Campylobacter* species, consisting of a 1,660,156 bp circular chromosome, isolated from a human skin infection.

KEYWORDS *Campylobacter*, clinical

Campylobacter species are Gram-negative, curved, or spiral-shaped bacteria, with a single polar flagellum at one or both ends or no flagellum (1). *Campylobacter jejuni* and *Campylobacter coli* are the most studied species, while the epidemiology and clinical roles of less common species remain poorly understood (2, 3). The strain GTC17093 used in this study was submitted for genetic analysis by a hospital located in Aichi, Japan. It was isolated from an epidermal cyst on the right chest of an adult patient during incision and drainage. This research was performed in accordance with the Declaration of Helsinki.

The sample was cultured anaerobically on Brucella HK agar plates (Kyokuto Pharmaceutical Industrial, Tokyo, Japan) with 5% sheep blood and incubated at 37°C for 48 h. The strain was subcultured three times under the same conditions. Genomic DNA was extracted from the pellet using Quick Taq HS DyeMix (TOYOBO Co., Ltd., Osaka, Japan) for all genome analyses.

The genome was sequenced using Oxford Nanopore Technologies long-read sequencing and Illumina short-read sequencing (4–6). For long-read sequencing, a library was prepared using the SQK-LSK-110 ligation kit (Oxford Nanopore Technologies [ONT]) without shearing, and small fragments were removed using a Short Read Eliminator XS (Circulomics, Baltimore, USA). Sequencing was performed on a GridION X5 (7) with a FLO-MIN106 flow cell (ONT). Guppy v.7.0.9 (super accuracy mode) was used for base calling, and reads were filtered using NanoFilt v.2.7.1 (8) with “-l 1000 -q 10 -headcrop 50.” For short-read sequencing, a library was prepared using an Illumina DNA Prep (M) Tagmentation Kit, and 2 × 151 bp paired-end sequencing was performed on a NovaSeq 6000 platform (Illumina, USA). Reads were processed using fastp v.0.20.1 (6) with “-q 30 -n 20 -t 1 -T 1.” Read quality was checked using Fastp and NanoPlot v.1.32.1 (8). Sequences were assembled using Unicycler v.0.4.8 (9) with default settings. The assembly was rotated to start with the dnaA gene, and Blobtools v.1.0 (10) was used to confirm data integrity. Average nucleotide identity (ANI) analysis was conducted using PyANI v.0.2.12 (ANIm algorithm) (11).

Genomic data are summarized in Table 1. Genome completeness and contamination were assessed using CheckM (v1.2.2), confirming 100.0% completeness and 0.0% contamination. The DDBJ Fast Annotation and Submission Tool (12) predicted 1,665 coding sequences, nine ribosomal RNAs, 46 transfer RNAs, and one CRISPR. Analysis of the 16S rRNA gene obtained from the genome sequence showed that *Campylobacter*

Editor Vanja Klepac-Ceraj, Wellesley College, Wellesley, Massachusetts, USA

Address correspondence to Masahiro Hayashi, hayashi.masahiro.w8@f.gifu-u.ac.jp.

The authors declare no conflict of interest.

Received 19 December 2024

Accepted 10 April 2025

Published 19 May 2025

Copyright © 2025 Yonetamari et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

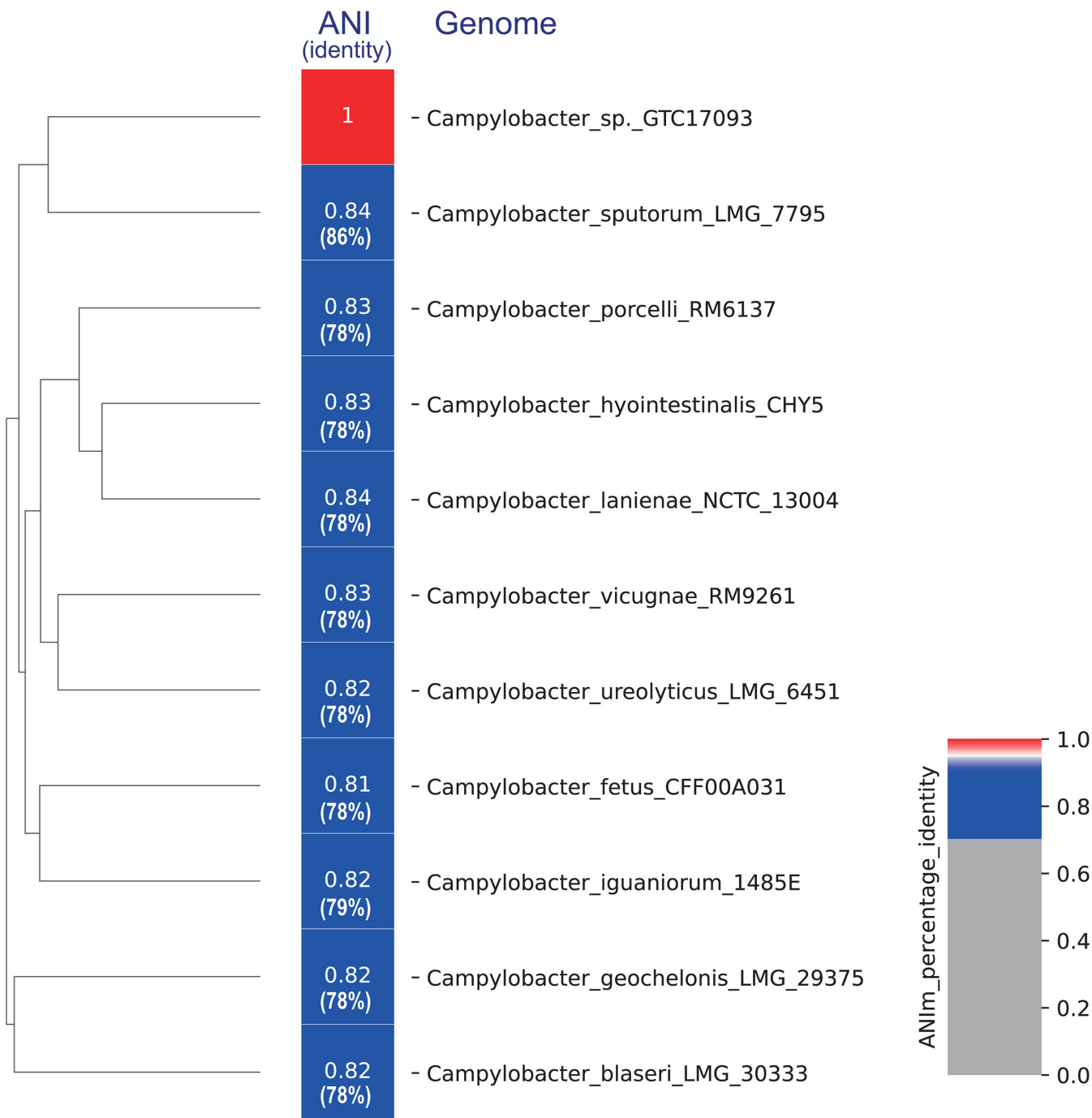


FIG 1 ANI heatmap, generated using PyANI (v.0.2.12) employing the ANIm algorithm, for GTC17093 genome and top 10 closest genomes identified via BLASTN analysis of the whole-genome sequence. The ANIm values are presented in the heat map; sequence identity values obtained via BLASTN analysis are indicated in the brackets.

sputorum LMG 7795 was the closest strain, with 98.1% identity. However, species identification with the genome sequence using GTDB-tk (v.2.3.2) was unsuccessful. BLASTN analysis of the complete genome sequence revealed that GTC17093 is most closely related to several *Campylobacter* species with ANI values between 81% and 84% (Fig. 1). The highest ANI value (84.0%) was observed with *Campylobacter sputorum* LMG 7795 (GenBank accession number: GCA008245005.1). However, the strain could not be decisively classified at the species level.

TABLE 1 Information regarding the complete genome sequence of *Campylobacter* sp. isolated from a human clinical non-diarrheal specimen

	Strain name
Parameter	<i>Campylobacter</i> sp. GTC17093 GTC17093
Illumina sequencing ^a	
No. of reads	1,921,262
Size (kb)	284,813
Avg coverage (x)	172
DRA accession no.	DRR621827
ONT sequencing ^a	
No. of reads	170,285
Size (kb)	1,243,671
Avg read length (bp)	10,808
Avg coverage (x)	749
N50	13,944
DRA accession no.	DRR621828
Assembly	
Estimated genome completeness (%) ^c	100.0
Estimated genome contamination (%) ^c	0.0
Genome structure	one chromosome
DDBJ/GenBank accession no.	AP038832
Genome size (bp)	1,660,156
GC content (%)	30.4
No. of coding sequences ^b	1,665
Number of rRNAs ^b	9
Number of tRNAs ^b	46
Number of CRISPRs ^b	1

^aDRA: DDBJ Sequence Read Archive.^bDFAST and DDBJ fast annotation and submission tools.^cDetermined using CheckM.

ACKNOWLEDGMENTS

We thank Kyoko Hatazaki, Akiko Katano, and Ayako Nagasawa for technical support.

AUTHOR AFFILIATIONS

¹United Graduate School of Drug Discovery and Medical Information Sciences, Gifu University, Gifu, Gifu, Japan

²Division of Clinical Laboratory, Gifu University Hospital, Gifu, Gifu, Japan

³Division of Anaerobe Research, Life Science Research Center, Gifu University, Gifu, Gifu, Japan

⁴Institute for Glyco-core Research iGCORE, Gifu University, Gifu, Gifu, Japan

⁵Gifu University Center for Conservation of Microbial Genetic Resource, Gifu, Gifu, Japan

AUTHOR ORCID^s

Masahiro Hayashi  <http://orcid.org/0000-0001-6317-9077>

AUTHOR CONTRIBUTIONS

Jun Yonetamari, Investigation, Methodology, Writing – original draft, Writing – review and editing | Masahiro Hayashi, Conceptualization, Funding acquisition, Resources, Supervision, Writing – review and editing | Yoshinori Muto, Software, Writing – review and editing | Kaori Tanaka, Writing – review and editing

DATA AVAILABILITY

The genome sequence of *Campylobacter* spp. (GTC17093) was deposited in the DDBJ (<https://www.ddbj.nig.ac.jp/index.html>) under the accession number [AP038832](#). Raw sequence data for GTC17093 were deposited in the DDBJ Sequence Read Archive under accession numbers [DRR621827](#) and [DRR621828](#).

ETHICS APPROVAL

This study was exempt from institutional ethics committee approval.

REFERENCES

1. Kaakoush NO, Castaño-Rodríguez N, Mitchell HM, Man SM. 2015. Global epidemiology of *Campylobacter* infection. Clin Microbiol Rev 28:687–720. <https://doi.org/10.1128/CMR.00006-15>
2. Iraola G, Pérez R, Naya H, Paolicchi F, Pastor E, Valenzuela S, Calleros L, Velilla A, Hernández M, Morsella C. 2014. Genomic evidence for the emergence and evolution of pathogenicity and niche preferences in the genus *Campylobacter*. Genome Biol Evol 6:2392–2405. <https://doi.org/10.1093/gbe/evu195>
3. Liu F, Ma R, Wang Y, Zhang L. 2018. The clinical importance of *Campylobacter* concisus and other human hosted *Campylobacter* species. Front Cell Infect Microbiol 8:243. <https://doi.org/10.3389/fcimb.2018.00243>
4. Sydenham TV, Overballe-Petersen S, Hasman H, Wexler H, Kemp M, Justesen US. 2019. Complete hybrid genome assembly of clinical multidrug-resistant *Bacteroides fragilis* isolates enables comprehensive identification of antimicrobial-resistance genes and plasmids. Microb Genom 5:e000312. <https://doi.org/10.1099/mgen.0.000312>
5. Vu H, Muto Y, Hayashi M, Noguchi H, Tanaka K, Yamamoto Y. 2022. Complete genome sequences of three *phocaeicola vulgatus* strains isolated from a healthy Japanese individual. Microbiol Resour Announc 11:e0112421. <https://doi.org/10.1128/mra.01124-21>
6. Hayashi M, Yonetamari J, Muto Y, Tanaka K. 2024. Complete genome sequence of *Peptostreptococcus porci* isolated from porcine endocarditis in Japan. Microbiol Resour Announc 13:e0020124. <https://doi.org/10.1128/mra.00201-24>
7. European Food Safety Authority and European Centre for Disease Prevention and Control (EFSA and ECDC). 2018. The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2017. EFSA J
8. De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. Bioinformatics 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>
9. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
10. Laetsch DR, Blaxter ML. 2017. BlobTools: interrogation of genome assemblies. F1000Res 6:1287. <https://doi.org/10.12688/f1000research.12232.1>
11. Pritchard L, Glover RH, Humphris S, Elphinstone JG, Toth IK. 2016. Genomics and taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. Anal Methods 8:12–24. <https://doi.org/10.1039/C5AY02550H>
12. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34:1037–1039. <https://doi.org/10.1093/bioinformatics/btx713>