

Coding-complete genome sequence of a largemouth bass virus isolate from *Micropterus salmoides* collected in China

Hengkang He,¹ Hao Huang,¹ Yang Zhang,¹ Xiaogang Yang,¹ Meisheng Yi,^{1,2} Kuntong Jia^{1,2}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT We report the coding-complete genome sequence of a largemouth bass virus isolate obtained from diseased largemouth bass (*Micropterus salmoides*) in China. The genome is 99,378 bp in length with a GC content of 52.1% and contains 102 predicted coding DNA sequences.

KEYWORDS largemouth bass virus, *Micropterus salmoides*, sequencing

Ranaviruses are double-stranded DNA viruses in the family *Iridoviridae* that infect a wide range of fish species (1, 2). Largemouth bass virus (LMBV) is a *Ranavirus* pathogen of largemouth bass (*Micropterus salmoides*) (3). Here, we report the coding-complete genome sequence of the LMBV isolate SYSUMS-2025.

The LMBV isolate whose coding-complete genome sequence is reported here (GenBank accession number: [PX879566](#)) was derived from diseased largemouth bass collected in Guangdong Province, China (23.479°N, 113.323°E), in 2025. Since no live animals were experimentally manipulated or sacrificed for this study, ethical approval was not required. Spleen tissues from diseased fish were homogenized in phosphate-buffered saline (PBS; pH 7.4), clarified by centrifugation, and inoculated onto the mandarin fish (*Siniperca kneri*) brain cell line (passage 31) for virus isolation as previously described (2). Subsequently, the virus was propagated in this cell line, harvested after cytopathic effects were observed, and concentrated via ultrafiltration. Total nucleic acids used for PCR and sequencing were extracted using the FastPure Viral DNA/RNA Mini Kit Pro (Vazyme), and the viral identity was confirmed by PCR amplification of the major capsid protein (MCP) gene using LMBV-specific primers (F: 5'-GCGGCCAACCAGTTTAACGCAA-3'; R: 5'-AGGACCCCTAGCTCTGCTTGAT-3') with 2× Taq PCR Master Mix (Vazyme, China). PCR cycling conditions were 95°C for 3 min; 25 cycles of 95°C for 30 s, 55°C for 30 s, and 72°C for 30 s; and 72°C for 3 min.

Whole-genome sequencing was performed using Oxford Nanopore Technologies (ONT) and Illumina platforms. A sequencing library was prepared using the SQK-NBD114.96 Kit (Oxford Nanopore Technologies). Sequencing was conducted on a PromethION platform using an R10.4.1 Flow Cell. Base calling, adapter trimming, and quality filtering were performed using Guppy (v6.5.7) (4). A total of 27,522 raw reads were generated, with an N50 read length of 9,607 bp. Host-derived reads were removed by mapping ONT reads to the *Siniperca kneri* reference genome (GenBank accession no. [GCF_020085105.1](#)) using minimap2 (v2.26-r1175) (5). The final assembly achieved an average genome coverage of 1,893.4×. For Illumina sequencing, DNA libraries were prepared using the VAHTS Universal Plus DNA Library Prep Kit for Illumina V2 (Vazyme), and paired-end sequencing (2 × 150 bp) was performed on an Illumina NovaSeq 6000 platform. After quality control and host read removal, 226.12 Mb of Illumina data were retained. Read trimming and adapter removal were performed using fastp (v0.20.0) with a Q20 threshold (6). *De novo* genome assembly was performed using ONT long reads in Flye (v2.9.2-b1786) (7). The draft assembly was subsequently polished using Illumina

Editor John J. Dennehy, Queens College
Department of Biology, Queens, New York, USA

Address correspondence to Meisheng Yi,
yimsh@mail.sysu.edu.cn, or Kuntong Jia,
jiakt3@mail.sysu.edu.cn.

The author declares no conflict of interest.

See the funding table on p. 3.

Received 28 February 2026

Accepted 16 March 2026

Published 20 April 2026

Copyright © 2026 He et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](#).

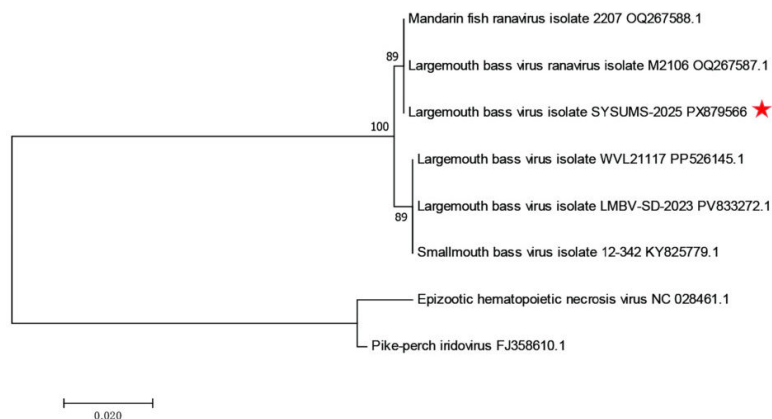


FIG 1 Maximum-likelihood phylogenetic tree based on amino acid sequences of the major capsid protein (MCP) of largemouth bass virus. Reference sequences were retrieved from the National Center for Biotechnology Information (NCBI) GenBank database using BLAST searches (accessed January 2026). Multiple sequence alignment was performed using MUSCLE implemented in MEGA7 (v7.0.26). The phylogenetic tree was inferred using the maximum-likelihood method with 1,000 bootstrap replicates in MEGA7 and visualized in the same software. Epizootic hematopoietic necrosis virus and pike-perch iridovirus were used as outgroups. The isolate SYSUMS-2025 obtained in this study is indicated by a red asterisk.

reads with Pilon (v1.24) (8). Coding DNA sequences were predicted and annotated using Prokka (v1.14.6) (9). Default parameters were used unless otherwise specified.

The coding-complete genome of the LMBV isolate was assembled into a single linear contig of 99,378 bp, with a GC content of 52.1%. A total of 102 coding DNA sequences were predicted. BLASTn analysis using default parameters showed that the genome shared up to 99.89% nucleotide identity with previously reported LMBV genomes, including strain M2106 (GenBank accession no. [OQ267587.1](https://www.ncbi.nlm.nih.gov/nuclseq/OQ267587.1)) (10). Phylogenetic analysis based on MCP amino acid sequences in MEGA7 showed that the SYSUMS-2025 isolate clustered within the LMBV clade together with other known LMBV isolates (Fig. 1) (11).

ACKNOWLEDGMENTS

This study was supported by the Natural Science Foundation of Guangdong Province (2023B1515120074), the Guangdong Province Special Support Plan Youth Top Talent Project (NIQN2024002), the Scientific and Technological Planning Project of Guangzhou City (2025B03J0032; 2023B03J1267), the National Natural Science Foundation of China (32473189, 32173001, and 32273115), and the National Natural Science Foundation of China and Israel Science Foundation Joint Scientific Research Program (32561143026).

AUTHOR AFFILIATIONS

¹School of Marine Sciences, Sun Yat-Sen University, Guangzhou, China

²Guangdong Provincial Key Laboratory of Marine Resources and Coastal Engineering, Guangzhou, China

AUTHOR ORCIDs

Hengkang He  <http://orcid.org/0009-0000-9260-5776>

Meisheng Yi  <http://orcid.org/0000-0003-1794-2734>

Kuntong Jia  <http://orcid.org/0000-0001-7674-9454>

FUNDING

Funder	Grant(s)	Author(s)
Natural Science Foundation of Guangdong Province	2023B1515120074	Meisheng Yi
Guangdong Special Support Plan	NIQN2024002	Kuntong Jia
Scientific and Technological Planning Project of Guangzhou City	2025B03J0032, 2023B03J1267	Kuntong Jia
National Natural Science Foundation of China	32473189, 32273115, 32173001	Meisheng Yi Kuntong Jia
National natural science foundation of china Israel science foundation joint scientific research program	32561143026	Kuntong Jia

AUTHOR CONTRIBUTIONS

Hengchang He, Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Software, Validation, Visualization, Writing – original draft, Writing – review and editing | Hao Huang, Conceptualization, Formal analysis, Investigation, Methodology, Visualization, Writing – original draft | Yang Zhang, Formal analysis, Investigation, Methodology, Software, Validation, Visualization | Xiaogang Yang, Data curation, Formal analysis, Investigation, Methodology, Resources, Software, Visualization, Writing – original draft | Meisheng Yi, Conceptualization, Funding acquisition, Supervision, Writing – original draft, Writing – review and editing | Kuntong Jia, Conceptualization, Formal analysis, Funding acquisition, Supervision, Writing – original draft, Writing – review and editing

DATA AVAILABILITY

The complete genome sequence of largemouth bass virus has been deposited in GenBank under accession number [PX879566](#). The Oxford Nanopore long-read sequencing data and Illumina short-read sequencing data have been deposited in the Sequence Read Archive under accession numbers [SRR36829122](#) and [SRR36864002](#), respectively, under BioProject [PRJNA1402781](#) and BioSample [SAMN54605172](#).

REFERENCES

- Duffus AL, Waltzek TB, Stöhr AC, Allender MC, Gotesman M, Whittington RJ, Hick P, Hines MK, Marschang RE. 2015. Distribution and host range of ranaviruses. In Gray MJ, Chinchar VG (ed), *Ranaviruses*. Springer, Cham.
- Zhang H, Dong J, Yan Y, Liu S, Ye X, Gao F, Sun C. 2023. Development of a highly permissive mandarin fish (*Siniperca chuatsi*) kidney cell line for mandarin fish ranavirus using a single-cell cloning method. *Cells* 13:18. <https://doi.org/10.3390/cells13010018>
- Zilberg D, Grizzle JM, Plumb JA. 2000. Preliminary description of lesions in juvenile largemouth bass injected with largemouth bass virus. *Dis Aquat Org* 39:143–146. <https://doi.org/10.3354/dao039143>
- Oxford Nanopore Technologies. 2023. Guppy basecalling software. Oxford Nanopore Technologies, Oxford, UK.
- Li H. 2018. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34:3094–3100. <https://doi.org/10.1093/bioinformatics/bty191>
- Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>
- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. *Nat Biotechnol* 37:540–546. <https://doi.org/10.1038/s41587-019-0072-8>
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. *PLoS One* 9:e112963. <https://doi.org/10.1371/journal.pone.0112963>
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>
- Yu XD, Ke F, Zhang QY, Gui JF. 2023. Genome characteristics of two ranavirus isolates from mandarin fish and largemouth bass. *Pathogens* 12:730. <https://doi.org/10.3390/pathogens12050730>
- Kumar S, Stecher G, Tamura K. 2016. Mega7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol* 33:1870–1874. <https://doi.org/10.1093/molbev/msw054>