

Coding-complete genome of human cytomegalovirus extracted from a nasopharyngeal swab in Brazil

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ABSTRACT We report the coding-complete genome sequence of *Cytomegalovirus humanbeta5* (BR/2023-012844/IEC/NS/2023), recovered from a nasopharyngeal swab collected from a 5-month-old infant with severe acute respiratory syndrome during a respiratory syncytial virus outbreak in Brazil.

KEYWORDS *Cytomegalovirus humanbeta5*, Brazil

Human cytomegalovirus (HCMV) is a member of the family *Herpesviridae*, subfamily *Betaherpesvirinae*, and genus *Cytomegalovirus* (1). Its genome consists of a double-stranded DNA molecule of >200 kbp (2). HCMV genomes are most commonly recovered from blood, urine, and saliva, particularly in congenital infections and in immunocompromised individuals (3–7). Recovering viral genomes from upper respiratory tract samples, such as nasopharyngeal swabs, is uncommon because this anatomical site is not routinely examined for HCMV in clinical or surveillance settings. However, obtaining the viral genome from the nasopharynx may provide key insights into viral shedding, reactivation, and coinfection dynamics. Shotgun metagenomics is well suited for recovering viral genomes directly from clinical samples (3). Notably, as of 1 September 2025, only partial HCMV genome sequences from Brazil were available in GenBank (8).

The nasopharyngeal swab analyzed in this study (2023-012844/IEC) was obtained from a 5-month-old infant hospitalized in an intensive care unit in Macapá, Amapá, located in the Brazilian Amazon. The GPS coordinates for sample collection were 0.0388900 and –51.0663900. The sample was collected in April 2023 by the Health Surveillance Secretariat during a respiratory syncytial virus (RSV) outbreak in Brazil. Viral DNA was extracted using the Extracta Kit – Pathogen DNA and RNA MDx, which employs magnetic microparticle technology, following the manufacturer's instructions. Library preparation and sequencing were performed at the Respiratory Virus Laboratory of the Evandro Chagas Institute. Libraries were constructed using the Illumina Nextera XT DNA Library Preparation Kit (Illumina, San Diego, CA, USA) and sequenced on the Illumina NextSeq 550 System with 2 × 150 bp paired-end reads, generating 39,712,796 read pairs. Quality filtering of reads in FASTQ format was performed using fastp v.0.23.1 (9), and *de novo* assembly was conducted using MEGAHIT v.1.2.9 (10) (Table 1). Taxonomic classification of contigs was carried out using Kraken2 v.2.1.0 (11) with the Standard-16 database, which identified multiple contigs assigned to *Cytomegalovirus*. Classification outputs were visualized using the Pavian R package v.1.2 (12). Manual review and open reading frame prediction were performed in Geneious Prime (13), where multiple overlapping contigs were identified. These contigs, supported by read mapping and alignment with MAFFT (14), allowed reconstruction of a coding-complete genome

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TABLE 1 Sequencing and genome metrics

Metric	Data for BR/2023-012844/IEC/NS/2023
No. of raw reads	39,712,796
No. of reads after trimming	36,229,206
Average read length (bp)	158
Total no. of contigs	912,240
No. of contigs related to <i>Cytomegalovirus humanbeta5</i>	16
Genome size (bp)	231,043
GC content (%)	57.5
Coverage (×)	801.4
N50	466
GenBank accession no.	PX246278

relative to the reference strain (GenBank accession number [PP803491](https://www.ncbi.nlm.nih.gov/nuclseq/PP803491)). Each contig was additionally confirmed by BLASTn (15).

The 2023-012844_CMV/IEC genome exhibited a typical HCMV genomic organization, with a length of 231,043 bp and 178 predicted genes. The mean read coverage was 801.4×, and the GC content (57.5%) was consistent with other *Cytomegalovirus humanbeta5* genomes available in GenBank. The genome displayed high nucleotide identity (>98%) with related HCMV strains available in GenBank ([KJ361950](https://www.ncbi.nlm.nih.gov/nuclseq/KJ361950); [KJ361951](https://www.ncbi.nlm.nih.gov/nuclseq/KJ361951); [KJ426589](https://www.ncbi.nlm.nih.gov/nuclseq/KJ426589); [KT726951](https://www.ncbi.nlm.nih.gov/nuclseq/KT726951); [KY490088](https://www.ncbi.nlm.nih.gov/nuclseq/KY490088); [MT044485](https://www.ncbi.nlm.nih.gov/nuclseq/MT044485)), demonstrating the genetic similarity of the 2023-012844/IEC strain to previously characterized isolates. The shotgun metagenomic approach provided evidence of HCMV genomic material in the nasopharynx of an RSV-positive infant with severe acute respiratory syndrome. As RSV infection can modulate host immune responses (16), further investigations are warranted to elucidate viral coinfections, HCMV reactivation, and the potential impact of congenital HCMV infection.

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AUTHOR CONTRIBUTIONS

Amanda Mendes Silva Cruz, Data curation, Methodology, Software, Writing – original draft | Edivaldo Costa Sousa, Jr, Data curation, Methodology, Software, Validation | Luana Soares Barbagelata, Methodology | Walquiria Aparecida Ferreira de Almeida, Investigation | Fernando Neto Tavares, Funding acquisition, Supervision, Writing – review and editing.

DATA AVAILABILITY

The whole genome sequencing project of sample BR/2023-012844/IEC/NS/2023 was deposited in GenBank under accession number [PX246278](https://www.ncbi.nlm.nih.gov/nuclseq/PX246278). The accession number for the raw shotgun metagenomic sequencing reads in the NCBI Sequence Read Archive is [SRR35176174](https://www.ncbi.nlm.nih.gov/sra/SRR35176174). This data set is associated with BioProject accession number [PRJNA131210](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA131210) and BioSample accession number [SAMN50854041](https://www.ncbi.nlm.nih.gov/biosample/SAMN50854041) (2023-012844/IEC).

ETHICS APPROVAL

The study was approved by the Research Ethics Committee of the Evandro Chagas Institute, Pará, Brazil (approval number 79631424.6.0000.0019).

REFERENCES

- International Committee on Taxonomy of Viruses. 2011. In ICTV ninth report: 2009 taxonomy release. https://ictv.global/report_9th/dsDNA/Herpesvirales/Herpesviridae.
- Murphy E, Shenk TE. 2008. Human Cytomegalovirus, p 1–19. In Human cytomegalovirus. Berlin, Heidelberg.
- Tsai HP, Yeh CS, Lin IT, Ko WC, Wang JR. 2020. Increasing cytomegalovirus detection rate from respiratory tract specimens by a new laboratory-developed automated molecular diagnostic test. Microorganisms 8:1063. <https://doi.org/10.3390/microorganisms8071063>
- Griffiths P, Reeves M. 2021. Pathogenesis of human cytomegalovirus in the immunocompromised host. Nat Rev Microbiol 19:759–773. <https://doi.org/10.1038/s41579-021-00582-z>
- Barroso Hernández S, Rodríguez Sabillón JA, Álvarez López Á, García de Vinuesa Calvo E, Calvo Cano A, Robles Pérez-Monteoliva NR. 2023. Enfermedad por citomegalovirus en los pacientes inmunodeprimidos por glomerulonefritis necrosante paucimune. Medicina Clínica 160:199–202. <https://doi.org/10.1016/j.medcli.2022.07.009>
- Giorgione V, Krajden Haratz K, Gull I, Brusilov M, Birnbaum R, Blecher Y, Malingier G, Kaplan A, Beer G, Kapusta L. 2023. Myocardial function in fetuses with congenital cytomegalovirus infection. Fetal Diagn Ther 50:430–437. <https://doi.org/10.1159/000533280>
- Ozdemir E, Sarac Sivriköz T, Sarsar K, Tureli D, Onel M, Demirci M, Yapar G, Yurtseven E, Has R, Agacıdan A, Kirkoyun Uysal H. 2024. Evaluation of congenital cytomegalovirus infection in pregnant women admitted to a university hospital in Istanbul. Viruses 16:414. <https://doi.org/10.3390/v16030414>
- National Center for Biotechnology Information. 2025. NCBI virus sequences for discovey. Available from: https://www.ncbi.nlm.nih.gov/labs/virus/vssi/#/virus?SeqType_s=Nucleotide&VirusLineage_ss=Human%20betaherpesvirus%205,%20taxid:10359&utm_source=nucore&utm_medium=referral
- Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>
- Li D, Luo R, Liu C-M, Leung C-M, Ting H-F, Sadakane K, Yamashita H, Lam T-W. 2016. MEGAHIT v1.0: A fast and scalable metagenome assembler driven by advanced methodologies and community practices. Methods 102:3–11. <https://doi.org/10.1016/j.jmeth.2016.02.020>

11. Wood DE, Lu J, Langmead B. 2019. Improved metagenomic analysis with Kraken 2. *Genome Biol* 20:257. <https://doi.org/10.1186/s13059-019-1891-0>
12. Breitwieser FP, Salzberg SL. 2020. Pavian: interactive analysis of metagenomics data for microbiome studies and pathogen identification. *Bioinformatics* 36:1303–1304. <https://doi.org/10.1093/bioinformatics/btz715>
13. Kears e M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28:1647–1649. <https://doi.org/10.1093/bioinformatics/bts199>
14. Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol* 30:772–780. <https://doi.org/10.1093/molbev/mst010>
15. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2)
16. Heinonen S, Velazquez VM, Ye F, Mertz S, Acero-Bedoya S, Smith B, Bunsow E, Garcia-Mauriño C, Oliva S, Cohen DM, Moore-Clingenpeel M, Peebles ME, Ramilo O, Mejias A. 2020. Immune profiles provide insights into respiratory syncytial virus disease severity in young children. *Sci Transl Med* 12:eaaw0268. <https://doi.org/10.1126/scitranslmed.aaw0268>